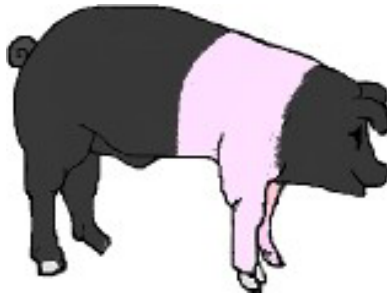




Animal &  
Plant Health  
Agency



# Great Britain pig quarterly report: disease surveillance and emerging threats

Volume 36: Quarter 2 of 2026 (April to June)

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# Introduction and overview

This report reviews potential disease threats for the second quarter of 2026 (April to June). See the [annexe on GOV.UK](#) for a full explanation of how data are analysed. Submissions to and diagnoses made through the Great Britain (GB: England, Wales and Scotland) scanning surveillance network can be interrogated using the interactive pig [disease surveillance dashboard](#). Diagnostic submissions are voluntary and subject to several sources of bias; data cannot be extrapolated to determine disease significance or prevalence. This report is compiled using data available at the time of writing. It contains disease findings gathered from APHA, Scotland's Rural College (SRUC) Veterinary Services and surveillance pathology partners, as well as intelligence gathered through the Pig Expert Group networks.

There is [guidance available for veterinarians](#) on sampling and testing pigs affected with different disease presentations. Veterinarians are encouraged to contact their regional Veterinary Investigation Centre (VIC) to discuss disease investigations with Veterinary Investigation Officers at APHA and SRUC.

## Unusual diagnoses or presentations

### Porcine circovirus 3 associated with congenital tremor

Six live preweaned piglets were submitted to the Thirsk VIC from two separate farrowing batches to investigate congenital tremors (CT) affecting primarily gilt litters on a large indoor breeding unit. Pigs were born tremoring and recovered if provided with additional care. The clinical and epidemiological features did not raise suspicion of classical swine fever (CSF).

All six pigs were normothermic; bright, alert and responsive; and had continuous whole-body tremors which lessened at rest. One piglet was also splay-legged. There were no cranial nerve deficits and no gross lesions were detected at post-mortem examination, other than one piglet with multifocal pinpoint haemorrhages over the kidney cortices. Fresh samples of brain and spinal cord were aseptically collected for molecular analysis. The remainder were fixed in formalin.

Histopathology on the brains and spinal cords of five piglets revealed moderate, diffuse, hypomyelination of spinal cord (confirmed by Luxol-fast blue staining – Figure 1) which was considered consistent with CT type AII. There was also mild, multifocal, non-suppurative encephalomyelitis.

Enhanced surveillance for disease associated with PCV3 is in place at APHA, using histopathology on pig hearts as an initial screen to detect non-suppurative myocarditis and/or periarteritis in fetuses, pigs or plucks received by APHA VICs for postmortem

examination. Where such lesions are detected, further investigation is progressed for detection of involvement of PCV2 by immuno-histochemistry (IHC) or PCV3 by *in situ* hybridisation (ISH). In these piglets, a non-suppurative myocarditis was additionally identified which raised suspicion of PCV3-associated disease.

Although atypical porcine pestivirus (APPV) has been confirmed in almost all cases of CT type All examined at APHA since discovery of the virus in 2015, APPV was not detected by PCR on brains and spinal cords from any of the six pigs. ISH (RNAScope) for PCV3 in the five examined pigs showed positive labelling of PCV3 RNA which correlated strongly with the demyelinated white matter tracts in the spinal cord. There were also a few smaller foci of staining in the cerebellar white matter and grey matter of spinal cord sections. ISH for PCV3 was also positive in hearts and lymph node. PCR for PCV3 on the brains and hearts was positive in all pigs, with low Ct levels (14.1 to 24.1) indicating high viral loads. Next generation sequencing (NGS) of central nervous system (CNS) and heart tissue from two pigs detected only PCV3 and no other viruses. Immunohistochemistry for PCV2 was negative. Given that APPV is typically detected in the majority or all CT type All-affected piglets (Arruda and others, 2016; Cagatay and others, 2019), it would be unusual for *in utero* APPV infection to no longer be detectable in any of the CT-affected piglets if APPV had been the cause in this outbreak.

PCV3 is a relatively recently discovered pig virus which, since 2016, has been described in pigs in countries including the United States (US), China, Poland, Italy, Spain and the UK (Collins and others, 2017; Palinski and others, 2017), with no zoonotic concern. PCV3 has been detected in samples from diseased and healthy pigs. Two main disease manifestations have been identified in a relatively low number of APHA submissions each year; PCV3-associated fetopathy and PCV3-associated systemic disease in postnatal pigs. Whilst PCV3 fetopathy outbreaks have been diagnosed, diagnoses of systemic disease due to PCV3 in postnatal pigs have been sporadic and tend to affect a single pig per batch submitted.

PCV3 has been detected in two previous cases of CT type All submitted to APHA. In one, APPV was detected by NGS and PCR and PCV3 by PCR only, with high Ct values indicating low viral loads. Therefore, the APPV was considered to be the cause of the CT ([quarter 4 2018 report](#)). In the other, APPV was not detected by PCR but PCV3 was detected by NGS and PCR, with no other viruses ([quarter 1 2023 report](#)). As PCV3 was present at relatively low viral loads and no myocarditis or multisystemic inflammation was identified in the affected pig, the criteria for a diagnosis of PCV3-associated disease were not reached and the PCV3 detected was of uncertain clinical significance.

In the literature, PCV3 has rarely been associated with CT and there may be [only one other report of its detection in CT-affected piglets without APPV](#) (Mora-Díaz and others (2020)). The authors detected PCV3 in brain and pooled systemic tissues by PCR with low Ct values in piglets with CT from one farm; tissues tested negative for APPV by PCR. ISH confirmed PCV3 presence in neurons, cardiomyocytes and smooth muscle of arterioles.

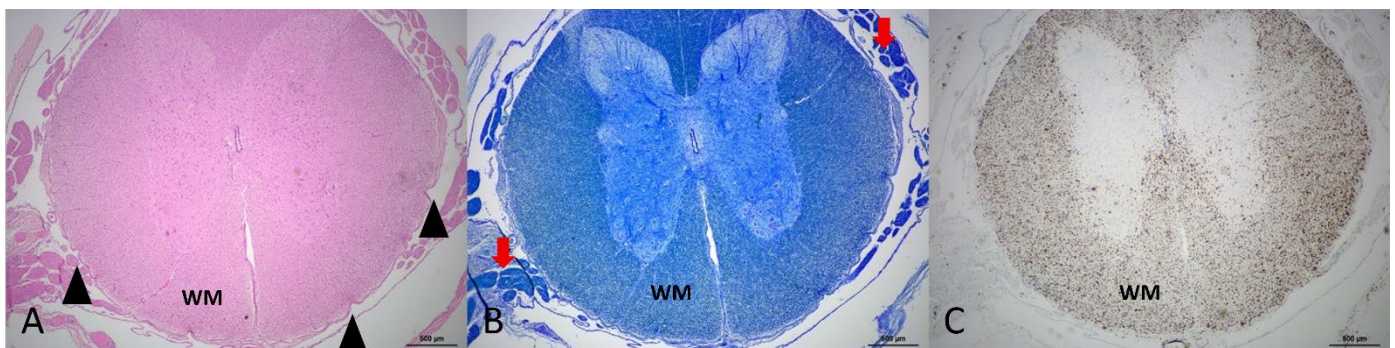
Arruda and others (2019) detected PCV3 by ISH and PCR in CNS tissues from piglets with CT from two farms in the US, in both cases, alongside APPV. Chen and others (2017) [detected PCV3 in piglets with CT in China](#) alongside APPV in five of the seven PCV3-positive samples and histopathological findings were not described. PCV3 has not been definitively established as a cause of CT as experimental infections of pregnant sows with PCV3 to produce CT-affected piglets have not been described (Cobos and others, 2025).

This case at APHA may be the one of the first in which: solely PCV3 has been detected in all tested piglets from an outbreak of CT type All with typical hypomyelination and nonsuppurative encephalitis; and PCV3 was confirmed in association with lesions by ISH and in high viral loads. We welcome information from others with similar findings to [pigexpertgroup@apha.gov.uk](mailto:pigexpertgroup@apha.gov.uk).

A previous diagnosis of PCV3-associated fetopathy was established on the farm in 2021 and of PCV-3 associated systemic disease in 2022, with concurrent diagnoses in both cases. Genetic analysis of the viruses from each of these PCV3 diagnoses is in progress.

Other causes of congenital tremor include *in utero* infection with certain viruses (one being CSF virus, for which suspicion [must be reported as suspect notifiable disease](#)), recessive genes or teratogens. More information about investigation CT is provided in a [previous surveillance item](#) and a [podcast](#). An [APHA presentation provides key features of PCV3 as well as APHA surveillance findings up to June 2021](#). The results of testing an archive of just over 800 sera from pigs sent to abattoirs in England in 2023-24 by PCR for PCV2, PCV3 and PCV4 at APHA (funded by AHDB) were described in the [April to June 2025 pig disease surveillance quarterly report](#).

**Figure 1: A section of spinal cord from a piglet with congenital tremors associated with PCV3. A: H&E. The white matter (WM) is diffusely pale staining and shrunken, giving a puckered outline (black arrow heads) to the spinal cord cross section. B: Luxol Fast Blue (LFB). Reduced staining of the spinal cord WM compared to the more intensely staining nerve roots (red arrows), indicating hypomyelination. C: ISH for PCV3. Positive brown immunolabelling present throughout the WM. Bar = 500µm.**



## **Salmonella Choleraesuis detected on a pig farm in England**

*Salmonella enterica* subspecies *enterica* serovar Choleraesuis (S. Choleraesuis) was isolated from an intestinal swab submitted to a private laboratory in March 2026 from a closed, breed-to-finish pig farm in England. Prior to this detection, no S. Choleraesuis isolations were reported to APHA in pigs since one from a pig at slaughter in 2015 which was part of an abattoir survey and not attributable to a specific farm. The last VIDA (Veterinary Investigation Diagnosis Analysis) diagnosis for salmonellosis due to S. Choleraesuis was in 2009.

[Salmonella is a reportable pathogen in the UK](#); APHA's *Salmonella* reference laboratory must be informed of suspected isolations of *Salmonella* in animals and provided with a culture of the organism by GB laboratories. In this case, the private laboratory submitted a suspect *Salmonella* isolate to APHA for definitive identification under the [Zoonoses Order \(1989\)](#).

S. Choleraesuis is a serotype adapted to pigs and, in naïve pigs, is considered to cause more severe disease than other serovars of *Salmonella*, most frequently leading to septicaemic disease. Historical information indicates that the S. Choleraesuis has been present long-term on the premises (since at least 2018), which has been continuously occupied with pigs for decades. At the time of APHA visits to the farm, the clinical effect of disease due to S. Choleraesuis in the pigs was assessed as very mild. The long-term infection in this closed herd with no pig introductions may have led to endemic stability.

The Food Standards Agency (FSA) assessed the risk to human health through consumption of pork originating from the herd to be low. Slaughterhouses and cutting plants operate under strict hygiene requirements that are monitored by the FSA. This, in addition to the handling instructions supplied on the products, provides confidence that the risk of contamination has been reduced to a level that protects the consumer.

Compared to the more commonly isolated serotypes S. Enteritidis and S. Typhimurium, each year, there are a small number of reported cases of human disease due to S. Choleraesuis in the UK. There were 42 isolations of S. Choleraesuis from human clinical samples at the UKHSA Gastrointestinal Bacteria Reference Unit between 2004 and September 2026. Genetic comparison of the isolate from this farm with clinical UK human isolates since 2014 and an isolate from a dog in 2025 was undertaken by the UK's Health Security Agency (UKHSA) and APHA. Isolates were not genetically similar.

UKHSA led on public health actions as part of this incident. In April 2026, UKHSA undertook a public health risk assessment to understand the level of occupational risk for farm workers and abattoir workers coming into contact with pigs from this farm. The risk assessment concluded that an occupational risk to farm workers and abattoir workers was present but was likely to be minimal as biosecurity and hygiene measures were already in place. UKHSA shared an information leaflet with farm workers and undertook passive

monitoring following the *S. Choleraesuis* detection on the farm. No human illness suspicious of salmonellosis has been described in people in contact with the farm's pigs.

APHA continues to assist the farm and their veterinarians in ensuring that actions are in place to protect human health, minimise spread within and off the farm and monitor for changes in pig health.

Laboratories in GB (including veterinarians undertaking microbiological culture at veterinary practices) are reminded of their [legal obligation to report suspect isolations of \*Salmonella\* to APHA](#). Veterinarians submitting samples to non-UK laboratories are encouraged to inform APHA of detections of reportable pathogens from farms in GB, as these findings are valuable for pathogen surveillance. Information on control of *Salmonella* on pig farms can be found on [GOV.UK](#). Resources on biosecurity for pig farms is available through [AHDB](#).

## Two cases of isoimmune disease in piglets

Two unrelated cases of immune-mediated damage to blood components or their precursors in bone marrow have been recently diagnosed in pigs by APHA, with differing presentations.

In the first, three euthanised two-day old piglets with skin discolouration and haemorrhage were submitted to the Thirsk VIC. Just one litter of piglets was affected on an indoor breeding unit. Affected piglets were not pyrexia and mortality across the farm had not increased, enabling concerns about swine fevers to be allayed.

The piglets had haemorrhages in the subcutis and musculature, over serosal surfaces and kidney cortices, as well as within the gastrointestinal tract. Piglets also had large, pale and friable livers. Histopathology of the bone marrow revealed mildly increased cellularity and adequate numbers of megakaryocytes. Haematology was not possible to confirm the presence of thrombocytopenia as piglets were submitted dead. Given the case presentation and lack of evidence for alternative explanations such as disseminated intravascular coagulation due to septicaemia, a diagnosis of isoimmune thrombocytopenic purpura (ITP) was proposed. ITP is a coagulation disorder in which circulating platelets are destroyed due to colostrum-derived maternal antibodies formed against foetal platelets. These piglets died in the first phase of disease. In the second phase of disease (late-onset ITP), there is destruction of bone marrow megakaryocytes.

In the second case, one 19-day-old piglet was found to be acutely lethargic with increased respiratory effort, discoloured ears and blotchy skin. It died shortly afterwards. Just one piglet was affected in a litter of 16 and no other litters were affected in this indoor, breed-to-finish herd.

The piglet was submitted to the Shrewsbury VIC for PME which revealed widespread, purple discolouration of the skin (Figure 1) and subcutis, as well as yellow discolouration of

the sclera. There were also haemorrhages over the serosal and mucosal surfaces of the gastrointestinal tract, mesentery and over the heart and kidneys; an increased quantity of red peritoneal fluid; watery blood; and generalised lymphadenopathy with dark red lymph nodes.

Histopathology of the bone marrow revealed a profound reduction in haematopoietic tissue affecting all cell lines (severe trilineage hypoplasia). The resultant thrombocytopaenia can explain the haemorrhagic diathesis. Erythrophagocytosis present in the bone marrow, liver and spleen indicated haemolysis as the cause of anaemia and jaundice. Given the age of the pig, a sporadic isoimmune condition affecting both circulating erythrocytes and bone marrow precursors was considered likely, with negative results from diagnostic testing for septicaemia, *Mycoplasma suis*, and pathogenic *Leptospira* serovars. The isoimmune condition in this pig differs from ITP where solely circulating platelets are affected.

Isoimmune conditions in piglets occur when piglets ingest colostral antibodies that their sow has produced against antigens present on foetal blood cells, which were inherited from the sire and to which the sow is incompatible. As sows are sensitised to these foetal antigens during an earlier pregnancy, ITP is not seen in gilt litters. Such conditions are now rare, probably due to the widespread use of artificial insemination (AI) for which boars are rotated frequently, making it less likely that a sow would be inseminated with the same boar semen in successive pregnancies. The variability of affected piglets within one litter is likely due to variable colostral antibody intake, differences in inherited antigens and because semen for AI may be pooled from several boars, meaning that the parentage of individual piglets within the litter can differ. Where isoimmune conditions occur following natural mating, the sow should not be served again with the same boar. More information on ITP is available on the [NADIS website](#).

**Figure 2: Purple discolouration and haemorrhages in the pale, slightly yellowed skin of a pig affected with isoimmune disease.**



# Horizon scanning

## Novel disease syndrome in pigs associated with a parvovirus variant in Europe

The Pig Expert Group has published [a new information note](#) requesting vets attending pigs to look out for a new disease syndrome seen on a growing number of pig farms in the Netherlands and also recently described on a single breeding farm in Denmark.

The syndrome is characterised by a combination of bilateral exophthalmos, generalised skin reddening, reduced growth and/or hair loss, plus, in some pigs, wrinkled skin, thickened neck, apparent strabismus, impaired growth and respiratory signs (hyperpnoea, tongue protrusion). Signs can affect up to 80% of pigs in 50% of the litters at the same time, but morbidity may be lower than this. Direct mortality tends to be low.

Disease mainly affects young piglets from around two weeks of age up to around 10 weeks of age. Gilts and finishers have also been seen to have bulging eyes. Outbreaks have lasted an average of 4 months (range 3-6 months). Consequences can include peri-weaning diarrhoea or greasy pig disease and batches become uneven with poorer quality finishers. No common or specific risk factors have yet been identified that trigger the disease.

A new parvovirus (PV) variant has been detected in high amounts in tissues from all affected pigs tested to date in both countries, and not in healthy pigs. This new PV variant differs from known porcine PVs and is most closely related to a PV detected in fox faeces and spleens in the Netherlands, Croatia and Denmark. It is not known how or when the virus was introduced into pigs in the Netherlands or Denmark. No disease was associated with the virus in the foxes in which it was detected and its pathogenic role in wildlife remains uncertain. No human illness, or concerns about human illness or food safety, are described in relation to the new PV variant in the Netherlands or Denmark, and there is no zoonotic concern in relation to endemic porcine PV present in UK pigs. Commercial vaccines for endemic porcine PV are not expected to provide immunity in pigs.

A definitive causal relationship between the new PV variant and this disease syndrome has not yet been established; further epidemiological studies and experimental trials are needed to confirm its role. In the meantime, pig keepers and vets in Europe are being alerted to the clinical signs described and encouraged to investigate any suspected cases, in order to identify any cases beyond the Netherlands and Denmark.

This syndrome is not known to have occurred in pigs in Great Britain (GB) and, to date, there is no information to know if the novel PV described occurs in pigs, foxes or other wildlife in GB. Live pigs and pig semen can be imported into GB from authorised EU countries or territories, subject to the applicable animal health requirements and

certification. However, very little is known at present about risk factors, transmission and risk pathways.

APHA encourages vets who see pigs in England or Wales with clinical signs resembling this disease syndrome to [contact their nearest APHA Veterinary Investigation Centre or surveillance pathology partner](#) and discuss with the duty vet. Based on information from cases in the Netherlands and Denmark, the key features expected in multiple pigs are:

- Bilateral exophthalmos (bulging eyes)
- Strabismus (eyes point in different directions)
- Generalised skin reddening
- Skin wrinkling
- Alopecia (hair loss or breakage)
- Reduced growth

The first two clinical signs are unusual and striking, and their occurrence in multiple pigs with some description of skin pathology is likely to qualify a case as a suspect for investigation. It is most likely that affected pigs for submission will be sucking piglets or weaners, but age is not a criterion for submission if the key clinical signs are present in other ages of pig. If you have any comments or queries, please contact [PigExpertGroup@apha.gov.uk](mailto:PigExpertGroup@apha.gov.uk).

## Porcine sapovirus research in the United States

[Swine Health Information Center \(SHIC\)-funded research on porcine sapoviruses \(PoSaVs\)](#) has successfully characterised and isolated contemporary PoSaV strains in the US. PoSaVs are genetically diverse and are widely distributed in pigs in pig-producing countries. PoSaV has been associated with diarrhoea and weight loss in pigs around and after weaning. Eight PoSaV genogroups have been identified, with genogroup GIII being most prevalent in US pig farms. Results from [a recent study](#) identified cell lines that allowed PoSaV propagation; samples known to be positive for PoSaV were passaged in three cell lines and PoSaV was successfully isolated in swine testicular and porcine kidney (LLC-PK1) cells. Characterisation of several recent GIII strains showed that they shared between 87.5% and 87.7% nucleotide identity in the viral capsid protein with the prototype GIII PoSaV Cowden strain which was detected 40 years ago. These developments are beneficial for future studies on PoSaV.

Wang and others (2020) previously reported detection of PoSaVs in pigs of varying ages with clinical diarrhoea in different US states, using next-generation sequencing and genetic characterization (Wang and others, 2020). The authors also detected other enteric viruses in all but one of the cases investigated.

Piglets experimentally infected with PoSaV may develop vomiting and/or diarrhoea. In the field, subclinical infection appears to occur and coinfection of PoSaV with other enteric

viruses is common, thus its contribution to clinical disease is uncertain. However, PoSaV infection has been increasingly identified in recent years in commercial US pig farms among young piglets with mild diarrhoea. [PoSaV was also detected](#) in cases of diarrhoea and poor growth with minimal mortality in piglets in Manitoba and Saskatchewan, Canada, in late 2022 to 2023.

[Past investigation at APHA](#) supports the likelihood that PoSaV is widespread in UK pigs. PoSaV was found in a high proportion of a small number of enteric samples from postweaned pigs from 2021-22 tested by metagenomics in an AHDB-BBSRC-funded project (see below - Dastjerdi and others, 2026). The proportion of positive samples detected at APHA were fairly equal between diarrhoeic and healthy groups of age-matched pigs. APHA found that the PoSaV viral load was higher in affected compared to healthy pigs, but the difference was not statistically significant. A [SHIC factsheet](#) describes the subclinical nature of many infections due to PoSaV and notes that the prevalence of PoSaV is often similar in pigs with and without diarrhoea, making the value of diagnostic testing questionable in diarrhoeic pigs. APHA has the capability to detect PSaV through next generation sequencing, should this be needed. The literature on PoSaVs will be kept under review.

## Information sources on global notifiable disease

APHA's International Disease Monitoring (IDM) team monitor any major, notifiable or new and emerging animal disease outbreaks worldwide. [See IDM's outbreak assessments which detail such outbreaks](#). Monthly IDM summaries are also included in the [disease surveillance items in the Veterinary Record](#).

Visit the .GOV.UK website for information on the latest situation on [foot-and-mouth disease](#) and [African swine fever \(ASF\)](#). The [European Commission also publishes information on ASF](#) and maps are available showing the current [EU ASF restriction zones](#). The Food and Agriculture Organisation (FAO) Emergency Prevention System for Animal Health (EMPRES-AH) produces regular ASF disease [situation updates for ASF in Asia and the Pacific](#). The [Swine Health Information Centre \(SHIC\) global reports](#) includes a detailed round-up of ASF in their global disease monitoring report each month.

AHDB issued a [reminder to pig producers](#) in England of the threat of ASF to the national pig herd. AHDB offers resources for ASF contingency planning, including webinars, workshops, podcasts and advice on contingency planning. Information on what food items or products of animal origin may be brought into the UK is found via [.GOV.UK](#) and [Food Standards Agency](#).

An [on-line guide with images](#) of the clinical signs and pathology of ASF is available to veterinarians and pig keepers. This notes that, at the start of an outbreak, deaths may initially just involve one or two pigs. Significantly increased mortality may only develop later once the virus has spread further in a group.

[Authors of a study which present a protocol to assess external biosecurity on commercial indoor pig farms in Spain](#) have emphasised the importance of strict biosecurity between domestic pigs and wildlife, particularly for wild boar in the prevention of ASF (Sebastián-Pardo and others, 2026). They identified that the presence of wild boar near the farm and fomites were key risk areas for possible entry of ASF.

Veterinarians and pig keepers must show vigilance and be familiar with the clinical signs of all notifiable diseases, so that suspicions are reported immediately. In England, this is by calling the Defra Rural Services Helpline on 03000 200 301. In Wales, contact 0300 303 8268 and in Scotland, contact your local APHA [Field Services Office](#). For information on notifiable diseases in animals, including disease controls, visit [.GOV.UK](#).

## Ongoing scanning surveillance initiatives

### New 'Dissecting Animal Diseases' APHA podcast episodes

APHA have released two new '[Dissecting Animal Diseases](#)' podcast episodes involving discussions around diagnostic investigations completed at APHA. Listen to the next two episodes, '[the different faces of oedema disease](#)' and '[swine fevers excluded in feral wild boar found dead](#)'. Follow the podcast on [Spotify](#) or [Apple podcasts](#) to never miss an episode.

### Virome of post-weaned diarrhoeic pigs and healthy cohorts in England

Members of APHA's Pig Expert Group co-authored [a publication](#) on work jointly funded by AHDB and BBSRC (Dastjerdi and others, 2026). [In the study](#), researchers used metagenomic sequencing to compare the presence, load and diversity of viruses in 36 enteric samples from pigs affected with post-weaning diarrhoea and age-matched healthy pigs on commercial farms in England.

More than ten viral families were identified from the samples and co-infection with four viruses (astrovirus, enterovirus, kobuvirus and smacovirus) was detected in all samples from both affected and healthy pigs. No notifiable viruses were detected, particularly no porcine enteric coronaviruses (which includes porcine epidemic diarrhoea virus), nor porcine teschovirus-1 (the cause of Teschen disease). Although some viruses were detected at a higher viral load in the samples from pigs with diarrhoea than healthy pigs (astro, entero, sapelo, sapo, posa, adeno and toro-viruses), the differences were not statistically significant.

The authors outline the complex nature of the viral element in the enteric microbiome of post-weaned pigs. They note that the involvement of some of the detected viruses in post-weaning diarrhoea is unclear and requires further study.

## **Brachyspira hyodysenteriae – swine dysentery**

*Brachyspira hyodysenteriae* is the cause of swine dysentery. Whole genome sequencing (WGS) and minimum inhibitory concentration (MIC) testing by broth microdilution is undertaken on a representative *B. hyodysenteriae* isolate from a submission from each premises (where successfully isolated and provided to APHA). This is completed at no charge to the submitting veterinarian, under APHA's pig disease scanning surveillance and 'Monitoring of Antimicrobial Resistance in Bacteria from Animals and their Environment' projects. WGS enables multilocus sequence typing (MLST). MLST is a tool for characterisation of isolates of a bacterial species by analysing sequence data of seven conserved genes in each *B. hyodysenteriae* isolate. This results in a combination of alleles known as a sequence type (ST) for each isolate. The multilocus sequence types of *B. hyodysenteriae* isolates from pigs in GB, as well as the genes or SNPs associated with reduced antimicrobial susceptibility that they possess, are represented on the [B. hyodysenteriae MLST dashboard](#).

AHDB's webpages on [biosecurity](#) and [swine dysentery](#), including the [#MuckFreeTruck](#) campaign, contain comprehensive information on appropriate biosecurity before, during and after a visit to a pig holding. Farms which are signed up to the pig industry's [Significant Diseases Charter](#) (which is now a requirement for Red Tractor assured farms) must report a diagnosis of swine dysentery to the Charter. Alerts are then issued to participants of the Charter to raise awareness about swine dysentery outbreaks. The Pig Expert Group recently collaborated with key representatives from the pig sector to publish an article describing [prevention, diagnosis and management of swine dysentery for the general farm animal vet](#) (Scott and others, 2025). In [a recent webinar](#), Dr Judith Rohdes presented on the current challenges for swine dysentery surveillance.

## **Porcine enteric coronavirus surveillance**

APHA carries out enhanced surveillance for porcine epidemic diarrhoea (PED) virus, transmissible gastroenteritis virus (TGEV) and porcine deltacoronavirus (PDCoV). Diagnostic submissions from cases of diarrhoea and/or enteropathy in pigs (non-suspect PED) submitted to APHA have been routinely tested by PCR for PED virus and transmissible gastroenteritis virus (TGEV) on a weekly basis. None have been positive for PEDV or TGEV in 2008 diagnostic submissions tested under AHDB Pork funding from June 2013 to June 2026. This enhanced surveillance has included testing for porcine deltacoronavirus (PDCoV) since February 2023 under the same funding and no PDCoV has been detected in the UK to date. The last diagnosis of PED and of TGE recorded in the GB national diagnostic database ([Veterinary Investigation Diagnosis Analysis](#) [VIDA]) was in 2002 and 1999, respectively. Porcine epidemic diarrhoea (PED) due to any PED virus strain remains notifiable in England and Scotland and [suspicion of disease](#), or confirmation of infection, [must be reported](#) (Defra, 2015; Scottish Government, 2016).

## Porcine reproductive and respiratory syndrome

Porcine reproductive and respiratory syndrome (PRRS) remains one of the most significant endemic viral infections in UK pigs. The APHA's [interactive PRRS dashboard](#) provides surveillance and diagnostic data from the GB scanning surveillance network for submissions diagnosed with PRRS from 2012 and has been updated to include data for 2024. All diagnoses made through the GB surveillance network have been due to PRRSV-1, with no PRRSV-2 detected in British pigs to date. The Pig Expert Group published an [information note](#) on preventing the introduction of exotic PRRSV strains into GB in imported live pigs or semen.

As part of PRRSV surveillance at APHA, ORF5 gene sequencing is undertaken under pig disease surveillance funding on the sample with the lowest Ct value (likely highest viral load) in each PCR-positive submission to APHA. This monitors diversity in the PRRSV detected and assesses for introduction or development of novel or genetically diverse PRRSV-1 strains into GB. Sequencing completed so far in 2026 has not detected any suspected new introductions. Viruses in which the ORF5 gene sequence has 98.5% or greater similarity to one of the live PRRSV vaccines are termed “vaccine-like”. As the ORF5 sequence analysis is based on just 4% of the genome, vaccine-like viruses are analysed further by sequencing part of the nonstructural protein 2 (nsp2) to help identify any potential recombinants.

## Swine influenza

Pigs with respiratory disease in the UK can be tested for swine influenza virus at no charge to the submitting veterinarian through the [Government-funded swine influenza surveillance project at APHA](#).

Samples are initially tested for the presence of influenza A Matrix (M) gene RNA. Following a positive detection, molecular assays are applied to determine the hemagglutinin (HA) and neuraminidase (NA) subtype of swine influenza A virus. This is useful for veterinarians considering vaccination of pigs and may help investigation of epidemiological links. Apart from [one H3N2 detection](#), the subtypes detected in the last year were all H1 viruses, belonging to three main genetic clades: H1N1 that emerged in the 2009 pandemic (Clade 1A.3.3.2); H1N2 viruses of the 1B.1.1.X clade that is unique to GB and was linked to the human case in Nov 2023 and; H1 viruses of the 1C.2.2 or ‘Eurasian avian-like’ clade that were prevalent in Europe before 2009 and have re-emerged in recent years. No H3N2 viruses were detected in GB pigs between 1997 and late 2025 when H3N2 was detected in one herd.

## Seneca Valley virus

In 2022, [APHA confirmed vesicular disease due to Seneca Valley virus \(SVV\) on five commercial breeding pig premises in one geographical area of England between June and](#)

[September](#). No cases of vesicular disease in pigs due to Seneca Valley virus were identified in GB in 2023, 2024 and 2025 and none to date in 2026. more information about the SVV outbreaks in pigs in England was [recently published](#) Armson and others (2026).

Key facts about SVV are provided [here](#). Kikuti and others (2025) have described [SVV surveillance data for US pig breeding herds from 2015 to 2025](#) and SHIC published an [informative webinar on SVV](#).

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