Animal &
Plant Health
Agency

Poultry Health Scheme (PHS) handbook

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PHS membership includes you making an authorisation under the Data Protection Act for release of relevant information to all those affected by the establishment's health status.

Contents

1. Introduction

This handbook contains information for:

- Prospective and participating poultry flock owners and exporters
- Animal and Plant Health Agency (APHA) staff
- Official Veterinarians (OVs) and private veterinarians providing their services to prospective and participating poultry flock owners and exporters

The Poultry Health Scheme (PHS) is based on the requirements of Commission Delegated Regulation (EU) 2019/2035 (hereafter, 'the regulation'), setting out rules for establishments keeping terrestrial animals and hatcheries, and the traceability of certain kept terrestrial animals and hatching eggs and supplementing Regulation (EU) 2016/429 of the European Parliament and of the Council. In the EU and Northern Ireland, this regulation repealed and replaced Council Directive 2009/158/EC (hereafter, 'the directive'), on animal health conditions governing intra-Union trade, and imports from third countries of, poultry and hatching eggs.

Although the objective and the scope of the PHS has not changed, the regulation added some additional details to the requirements. The main differences are:

- the existing requirement to operate a microbiological control programme at the hatchery has been specified to at least 60 samples to be taken every 6 weeks (see Annex I)
- hatchery sampling for Salmonella is required to complement the surveillance on farms (see Annex I)
- an additional initial sampling was added to the Mycoplasma testing schedule (see Annex I)
- clinical examination is now required as part of the disease surveillance
- all diagnostic testing for the purpose of the PHS must be carried out at official laboratories designated by the competent authority. Laboratory designations are on a pathogen and test specific basis.
- some records must **be** retained for longer.

The requirements are listed in Regulation 2019/2035. Granting approval of hatcheries is described in Article 7 and approval of establishments (farms) is laid down in Article 8 of the Regulation. The requirements for biosecurity measures, disease surveillance, microbiological control programme, facilities, personnel, and supervision by the Competent Authority (CA) are listed in Annexes I and II of the same Regulation.

The PHS and this document intend to reflect the EU regulatory requirements for the registration and approval of premises intending to export breeding and productive poultry, day old chicks and hatching eggs from Great Britain to the European Union or Northern Ireland.

Some non-EU countries also require the premises of origin of some poultry consignments to be part of a ‘government supervised poultry health scheme’, in which case the reference in the relevant Export Health Certificate (EHC) means the PHS.

Read about how to join the PHS, renew or cancel your membership on GOV.UK.

2. Birds that are covered

The PHS only covers poultry premises – hatcheries, farms and egg distribution centres (EDCs). PHS-approved premises must only house poultry.

For the purpose of the PHS, ‘poultry’ is defined as birds that are reared or kept in captivity for the production of meat, eggs for consumption, other products, restocking supplies of game birds, or for the purpose of breeding of birds used for the types of production for meat or eggs.

Where appropriate, PHS premises can be defined as a separate part of a larger establishment. In that case, there must be clear separation and effective biosecurity between the PHS and the other parts of the premises.

3. Managing your membership

Membership of the PHS in Great Britain is open to individuals or companies operating at specified premises. If you operate premises at more than one location, you need to register each premise separately.

The categories of membership of the PHS are:

- farm
- hatchery (including egg distribution centres)
- combined farm and hatchery

APHA must approve membership of the PHS. Read about how to join the PHS, renew or cancel your membership on GOV.UK.

Fees: registration and membership

APHA review the PHS fees annually.

You must pay a registration fee when you apply to join the PHS.

You must also pay the membership fee when you are accepted into the PHS, then annually before your membership expires. APHA will send a reminder 4 to 6 weeks before your renewal fee is due.

Initial inspection visit

An initial inspection of your premises will be carried out by an APHA veterinarian. This inspection will establish whether your premises and its management procedures meet the requirements of Regulation 2019/2035 (as amended). You can find these requirements for farms and/or hatcheries in the relevant sections of this handbook.

If the inspection of your premises, operations or required test results are not satisfactory, APHA will provide written notification outlining the reasons for you not meeting the required standards. Where you can take actions to meet membership requirements, you will be informed of this, and you will have a limited time to make the changes needed to obtain membership. APHA will inform you if a further visit is required, depending on the nature of the corrective actions. You will need to pay another APHA inspection fee if a vet needs to inspect your premises again.

When the inspection of your premises, operations and required test results are satisfactory, your membership will be approved, after required fees have been paid.

Initial testing

Depending on the species of poultry and the origin of the birds, you may need to undertake testing for Salmonella and Mycoplasma before you can obtain membership of the scheme. For example, if the poultry are not originating from an approved PHS premises, you will need to have surveillance testing done (species dependent) with satisfactory results.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory's UKAS number or name here: UKAS accredited organisations.

Along with your test results, you must keep systematic records, detailed and comprehensive standard operation procedures (SOPs) and, in the case of hatcheries and

egg distribution centres, a description of and in-house guidance of procedures for the microbiological control programme as referred to in Article 7 of Regulation 2019/2035. These must be available for inspection.

When your membership starts

Your membership lasts for one year.

As long as any required test results are satisfactory, and the membership fee and inspection fees have been paid, then your membership will start from the date of the satisfactory inspection visit. If there are pending test results, the membership will start once satisfactory test results are received.

If poultry are not present at the initial approval visit, your membership will begin once poultry have been placed at the premises.

PHS membership and differing residency periods may be required by the competent authorities of countries of destination. Such requirements are specified in the relevant Export Health Certificate. You can find Export Health Certificates on the Form Finder pages of GOV.UK.

Membership number

APHA will send you your PHS membership number and certificate by post after your membership starts.

Your PHS approval number will mirror your county parish holding (CPH) number. Approved establishments will be added to a PHS list of members on GOV.UK.

If you do not already have a CPH number for the site where your poultry are kept, contact:

- England - contact the Rural Payments Agency (RPA) on 03000 200 301
- Wales - use manage my CPH on Rural Payments Wales online
- Scotland - contact Scottish Government Rural Payments and Services

Getting new birds and hatching eggs

From your membership start date, birds and hatching eggs may only enter your flock, hatchery or egg distribution centre from:

- PHS establishments in England, Wales and Scotland or from establishments approved under the equivalent Northern Ireland Poultry Health Assurance Scheme (NIPHAS)

- approved establishments in EU member states - equivalent to PHS

- establishments in non-EU countries in accordance with current import requirements and imported with the relevant import health certificate as published on GOV.UK for poultry and hatching eggs over 20 units, i.e. in compliance with PHS requirements

4. Before you can export: 6-week wait period

As a rule, newly approved premises cannot export to the EU and Northern Ireland until they have been members of the PHS for at least 6 weeks. You can only export eggs and day-old birds to the EU or Northern Ireland after the 6-week wait period at the hatchery and the flock(s) of origin has finished.

Sampling and testing must be done in these 6 weeks under the rules of the PHS (See Annex 1). For Mycoplasma, testing by bacteriology is allowed but not recommended as it takes 21 days of culture to report a negative result.

You can send eggs to PHS approved hatcheries or egg distribution centres within Great Britain during this period.

Resident flocks: 6-week wait period

If you have resident flocks when the APHA vet visits and other requirements satisfy PHS membership, the 6-week period starts from the day of the APHA vet visit if the resident flocks have been tested twice for the relevant pathogens. The time between tests must be 14 to 42 days with negative results.

APHA will issue your letter of membership confirmation when they receive the second set of negative results.

In this case, you can export 6 weeks after the satisfactory vet visit.

No stock: 6-week wait period

If you do not have stock at the time of the APHA vet visit and all other requirements satisfy PHS, the 6-week period starts from the day of the APHA vet visit if the flocks to be placed have come from a PHS member that has been a member themselves for longer than 6 weeks. You will not need to test the birds to become approved.

If the flocks you get are not from a PHS member, you need to have two tests for the relevant pathogens that must be 14 to 42 days apart, with negative results. In this case, you can export 6 weeks after you populate the premises.

5. Annual inspections

A satisfactory veterinary inspection must take place once a year, on or before the anniversary of the previous inspection. APHA will write to you to advise you if the inspection is overdue.

Following the inspection, when APHA has received your renewal form and fee, they will send you a renewal letter confirming your membership is valid for the next year. Your membership anniversary date remains the same.

If APHA do not receive an inspection report for your premises by your renewal date, then your membership will be suspended with effect from the missed annual re-approval date and ultimately will be revoked if there is a further delay in APHA receiving your inspection report.

Official veterinarian (OV) annual inspection requirements

If the member opts for an OV inspection instead of APHA inspections, the veterinary surgeon must be an OV authorised for the export of poultry. To avoid conflict of interest whilst acting in their official capacity, they must not work full-time for the member party and respect their professional obligations.

APHA can supply a list of eligible OVs. If you choose the OV option, it is your responsibility to arrange for the OV to make the annual inspection. The OV of your choice should send the completed inspection report to APHA.

You will need to pay the fees of the OV for the inspection. They are separate from the PHS membership fees.

Your membership could be suspended or revoked if we receive an unsatisfactory report.

If your membership is revoked, an APHA vet (not an OV) will need to inspect your premises before your membership can be reinstated. You will be required to pay the registration fee and veterinary inspection fee again, and a new membership certificate will be issued upon re-approval.

If your membership is suspended and you have resolved the issues, an APHA vet might need to visit (subject to the veterinary inspection fee) to inspect your premises before APHA can lift the suspension. Any non-compliance must be resolved by APHA inspections.

6. Suspension and revocation of memberships

Suspension of PHS membership

APHA will suspend your membership if any of these apply:

- you no longer meet the conditions laid down in Articles 7 and 8 to Regulation (EU) 2019/2035
- there is a pending disease investigation on your premises for Avian Influenza or Newcastle disease
- you have received poultry or hatching eggs from an establishment with a suspected or confirmed case of Avian Influenza or Newcastle disease, pending the completion of disease investigation
- your premises has been identified as a destination after a tracing from establishments where Avian Influenza or Newcastle disease has been confirmed
- the results of surveillance at your premises may indicate presence of Salmonella pullorum or S. gallinarum or in turkeys pathogenic S. arizonae, or Mycoplasma gallisepticum or M. meleagridis (subject to the relevance for the particular poultry species, while further tests are performed)
- you have not completed measures required by a notice served by an OV or APHA vet if the appropriate disease surveillance (if required) is not carried out at the establishment or the establishment does not satisfy the requirements of Regulation (EU) 2019/2035 (until you complete the measures)
- your membership has lapsed beyond the 365-day period and APHA has not received your renewal application form or membership fees - membership will be revoked after 30 days from the renewal date
- a vet has not inspected your premises and given a satisfactory report before your annual membership expires

The duration of the suspension will be determined by APHA (in consultation with the inspecting veterinarian) depending on the reason for suspension.

Revocation of your PHS membership

APHA will revoke your membership if any of these apply:

- Avian Influenza or Newcastle disease is confirmed on your premises

- Salmonella pullorum, S. gallinarum, pathogenic turkey S. arizonae strains, Mycoplasma gallisepticum or Mycoplasma meleagridis is confirmed on your premises (subject to the relevance for your poultry species)

- you did not take the action after an OV or APHA vet served a second notice, requiring you to implement the appropriate disease surveillance or to bring the establishment in line with the requirements of Regulation 2019/2035, outlined in this handbook

- membership lapsed beyond the 365-day period and APHA has not received your renewal application or membership fee 30 days after the renewal date

- a vet has not inspected the premises and submitted a satisfactory report within 3 weeks of due date

7. Restoring your membership

You must demonstrate compliance with the rules of the scheme for APHA to grant approval, including allowing and paying for an additional APHA visit where appropriate and proportionate to the breach. For example, in the case of an AI outbreak in a PHS farm, the epidemiology report will give the cue to check on farm biosecurity before reapproval.

APHA may reinstate your membership if it was revoked for one or more of the below reasons:

1. Confirmation of Avian Influenza or Newcastle disease - APHA may restore your membership 21 days after completion of the cleansing and disinfection protocol following depopulation.

2. Infection caused by S. pullorum, S. gallinarum, pathogenic turkey S. arizona strains - APHA may restore your membership after negative results have been recorded in 2 rounds of suitable sampling of the new flock. APHA will advise on the number and type of samples and the methodology according to the circumstances of the case. You should perform sampling at intervals of at least 21 days, starting after depopulation and disinfection of the infected premises, and the effectiveness of disinfection has been verified by checking the standard of the cleansing and disinfection protocol and, where appropriate, by suitable tests on dried surfaces. APHA staff may need to carry out the sampling and testing and advise on the effectiveness of disinfection.

3. Following the presence of Mycoplasma gallisepticum or Mycoplasma meleagridis in relevant poultry species, negative tests have been recorded from 2 separate samples performed in a flock’s premises either:

i) following depopulation of the entire flock, then cleansing and disinfection, and repopulation - two tests have been performed with an interval of at least 21 days with negative results

ii) two samples and tests have been completed with an interval of at least 60 days - APHA will advise on the number and type of samples to be taken

For the purpose of the PHS, which aims to control for certain Salmonella and other pathogens, 'disinfection' requires at least a Defra-approved disinfectant at the appropriate dilution rate.

4. Non-compliance with the rules of Regulation 2019/2035, subject to notices served by an APHA veterinarian or OV. APHA may restore membership when they agree that you are complying with all the Regulation requirements.

5. Your membership has lapsed beyond the 365-day period and a satisfactory annual visit has not been completed. APHA may restore your membership after a satisfactory inspection is completed, and you have paid a further registration fee.

If your membership is restored following revocation, you must wait 6 weeks from the re-approval date before you can export.

8. List of approved establishments and advertisements

You can find a list of Poultry Health Scheme members on GOV.UK.

The regulation requires central competent authorities to make the list of approved establishments publicly available.

You can draw attention to your PHS membership in advertising, as long as the advertising correctly describes your membership status. APHA reserves the right to require the withdrawal of any advertising material it considers to be inaccurate.

9. Requirements for approved Poultry Health Scheme (PHS) members

Farms must demonstrate compliance with the rules of the PHS and Regulation 2019/2035 regarding:

- biosecurity (point 1 of Part 4 of Annex I to Regulation 2019/2035)
- surveillance (point 2 of Part 4 of Annex I to Regulation 2019/2035)
- facilities (point 3 of Part 4 of Annex I to Regulation 2019/2035)

Hatcheries must demonstrate compliance with the rules of the PHS and Regulation 2019/2035 regarding:

- biosecurity (point 1 of Part 3 of Annex I to Regulation 2019/2035)
- surveillance (point 2 of Part 3 of Annex I to Regulation 2019/2035 and Parts I and II to Annex II to Regulation 2019/2035)
- facilities (point 3 of Part 3 of Annex I to Regulation 2019/2035)
- staff (point 4 of Part 3 of Annex I to Regulation 2019/2035)
- supervision by the competent authority (point 5 of Part 3 of Annex I to Regulation 2019/2035)

Egg Distribution Centres (EDCs) are considered hatcheries under Article 4(47) of Regulation (EU) 2016/429. EDCs must be formally approved under the PHS in accordance with Article 7 of Delegated Regulation (EU) 2019/2035. Approval is granted based on compliance with biosecurity, hygiene and traceability standards.

10. Biosecurity

You must maintain a high level of protection against disease in PHS approved establishments by using effective biosecurity measures.

Biosecurity prevents disease causing agents entering or leaving places where they can pose a risk to:

- farm animals
- other animals
- humans
- the safety and quality of a food product

The same principles apply at the farm, in hatcheries and in egg distribution centres. The aim is minimising the risk of disease spreading to, from and between animals or separated groups and preventing cross-contamination of eggs, goods and premises.

Biosecurity in the poultry industry

Large groups of poultry kept in intensive units can significantly increase the risk of spreading avian diseases. This risk should be carefully considered, especially on mixed farming operations.

Special attention should be given to the presence of game birds, backyard poultry flocks and wild bird populations, as these can act as agents for introducing diseases onto PHS premises. Poultry are susceptible to a range of zoonotic pathogens – those transmissible

to humans – including specified Salmonella types and Campylobacter spp., highlighting the importance of robust biosecurity and monitoring protocols.

Poultry premises must implement and maintain the following Standard Operating Procedures (SOPs) as a minimum:

- **biosecurity protocols** to prevent disease introduction and spread
- **cleaning and disinfection** routines for all equipment and storage areas
- **pest control** measures to prevent contamination
- **transport hygiene** standards for incoming and outgoing vehicles
- **traceability systems** to track egg batches from source to destination

PHS premises must be officially supervised and subject to periodic inspections to ensure compliance with PHS requirements.

Records of hygiene testing, SOP implementation and bird and/or egg traceability must be maintained and made available upon request.

Additional testing for Salmonella pullorum/gallinarum/arizonae and Mycoplasma gallisepticum/meleagridis is required on farms and hatcheries. This testing is not required at egg distribution centres (EDCs) due to minimal handling of eggs at these premises.

For the purpose of the PHS, which includes testing for specified Salmonella and other pathogens, 'disinfection' requires at least a Defra-approved disinfectant at a concentration appropriate to the purpose. ,

The below Codes of Practice, along with guidance provided by APHA during official farm visits, provide specific advice that highlights the importance of:

- careful cleansing and disinfection of units before stocking and restocking
- effective control of farm pests
- separation of the unit from other farm activity
- rigorous hygiene measures when entering and leaving the unit (for example, separate clothing and footwear)
- making sure all inputs such as feed, water and bedding are handled and stored in such a way to ensure they are free of contamination

Codes of practice and assurance schemes in the poultry industry

- European rules for Salmonella control
- Biosecurity and preventing welfare impacts in poultry and captive birds
- Code of Practice for the welfare of laying hens and pullets
- Code of Practice for the prevention and control of Salmonella in commercial egg laying flocks

- Code of Practice for the prevention and control of Salmonella in chickens reared for meat on farm
- Code of Practice for the prevention and control of Salmonella in breeding flocks and hatcheries
- Code of Practice for Lion eggs

You can find more information on disease prevention for livestock farmers on GOV.UK.

11. Farm biosecurity requirements

On an approved farm, the hatching eggs must be:

- collected frequently - at least once a day and as soon as possible after laying
- cleaned and sanitised as soon as possible, unless sanitisation takes place at a hatchery in Great Britain
- placed in new or cleaned and disinfected packaging

If your establishment houses poultry species of the orders Galliformes and Anseriformes at the same time, you must make and maintain a clear separation between them.

You must include appropriate sanitary breaks after cleaning and disinfecting operations and before a new flock of poultry arrives in the facilities.

Staff must wear appropriate work clothing and visitors must wear protective clothing. Everyone must follow the hygiene rules drawn up by the operator.

An appropriate wastewater collection system must be in place to prevent introduction and spread of disease. Concerns regarding the disposal of wastewater may be highlighted to the relevant authority.

12. Hatcheries and egg distribution centre biosecurity requirements

Poultry hatching eggs must come from either PHS approved establishments keeping breeding poultry, other PHS hatcheries, or equivalent approved sources in the case of imported eggs as per the import requirements specified in the health certificate.

Unless the establishment of origin sanitised the eggs, you must clean and sanitise eggs either:

- between their arrival at the hatchery and incubation
- at the time of their dispatch

You must clean and disinfect the:

- incubators and all equipment that has come into contact with eggs or chicks, after hatching
- packaging materials after each use, unless they are single use

Appropriate systems must be in place to ensure the collection of wastewater place to prevent introduction and spread of disease. Concerns regarding its disposal may be highlighted to the relevant authority.

All visitors and staff working in the hatchery must be provided with appropriate protective clothing and are required to follow all relevant biosecurity protocols. In addition, staff must adhere to the hygiene code of conduct at all times.

13. Routine testing

You might need to carry out routine testing for certain Salmonella and Mycoplasma. This will depend on the type of poultry you keep, their origin, and whether they are present on site at the time of the initial APHA inspection.

Any positive PHS test result must be reported immediately to the APHA PHS team (Customer Service Centre – One Health). Either the Private Veterinary Surgeon or the poultry producer is responsible for notifying APHA.

Advice on sampling in case of a relevant Salmonella disease breakdown can be obtained from the APHA Salmonella disease consultant.

All test regimes are subject to regular review. APHA will notify you of any changes in writing. The current testing requirements are laid out in Annex I of this handbook. Table 1 includes a summary.

Table 1: Summary of testing requirements

	Salmonella pullorum	Salmonella gallinarum	Salmonella arizonae	Mycoplasma gallisepticum	Mycoplasma meleagridis
Turkeys	Required	Required	Required	Required	Required
Fowl	Required	Required	Not required	Required	Not required
Ducks	Required	Required	Not required	Not required	Not required
Pheasants, guinea fowl, partridges and quail	Required	Required	Not required	Not required	Not required
Geese, pigeons, ratites	Not required	Not required	Not required	Not required	Not required

Samples must be collected in accordance with the PH30 Samples Collection Guidance. A veterinary surgeon is responsible for providing oversight, including ensuring appropriate training is delivered, verifying that the samples are collected correctly, and reviewing the results.

For each PHS-approved premises, at the time of initial approval and at each annual renewal, the supervising veterinarian must submit a written sampling protocol and a signed declaration, to the approving or renewing veterinarian (APHA or OV). This submission must confirm that the agreed testing protocols have been followed and that the test results demonstrate compliance with the PHS sampling and testing regime.

The OV or APHA vet carrying out the annual PHS inspection must not have any conflict of interest, as the scheme is intended to support export certification and international trade compliance.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory's UKAS number or name here: UKAS accredited organisations.

You must pay for all sampling and testing associated with PHS membership.

APHA can advise on any need for additional testing of your flock or the environment of your establishment. For example, when resident birds are present on the premises before approval and following confirmation of any of the diseases covered by the PHS.

14. Clinical examination

An OV or RCVS registered Private Vet Surgeon (PVS) must clinically examine flocks during each laying or productive period at the best time for detecting the disease in question. You must keep a record of these examinations.

For breeding flocks, the point of lay is the best time for detecting disease. This is when the birds are more stressed and would most likely show signs of disease for Salmonella and Mycoplasma.

Examination of birds can be carried out at the rearing premises or at the PHS premises of destination. In the latter case the vet at origin will need to be provided with evidence that examination will be carried out at destination before they give approval.

For productive flocks, the birds must be examined for disease at least once during the productive period.

In all cases, the examination must be performed at least once per production cycle or, at a minimum, once annually to cover the risk period.

The owner or agent must notify the OV of any variation in hatchability or the presence of Salmonella and Mycoplasma in the context of the Poultry Health Scheme. If necessary, samples for diagnosis must be submitted without delay to the APHA approved laboratory. The owner or agent must notify APHA immediately following signs of notifiable diseases including the presence of Newcastle disease and Avian Influenza so a veterinary enquiry can be made.

15. Facilities: PHS approved farms

The facility setting and layout must be compatible with the type of production undertaken.

Facilities must:

- help prevent the introduction of disease
- allow you to control disease if it is introduced
- have clear separation between different types of poultry and flocks
- have good hygiene conditions
- allow you to monitor the health of the flocks

Equipment must:

- be compatible with the type of production pursued
- allow effective cleaning and disinfection of the facilities and vehicles used for transporting poultry and eggs

Farm operations

You must use measures to protect against the introduction of disease. Base your rearing techniques on high biosecurity principles and procedures including, where feasible, operate to the all-in and all-out principle. You must clean and disinfect that part of the establishment after depopulation.

You can refer to the guidance available in Section 9 of the PHS handbook or the information on GOV.UK - Disease prevention for livestock and poultry keepers - GOV.UK

The premises must contain only poultry and hatching eggs:

- from PHS establishments
- from establishments in EU member states similarly approved in accordance with Regulation 2019/2035
- imported from third countries in accordance with Retained Regulation 798/2008

You must define hygiene rules in writing and agree them with the inspecting APHA vet or OV.

All personnel and visitors must wear appropriate protective clothing.

You must keep buildings, pens, and equipment in good repair.

Your standard operating protocols (SOPs) must contain a written protocol for all sampling and testing procedures. This must include the name of the veterinarian responsible, test type and official laboratory used.

Record keeping

After disposal of your flock, you must keep records for at least 3 years. These records must include:

- when you moved stock onto and off the premises
- the production performance of the flock
- morbidity and mortality rates with causes
- any laboratory tests and their results
- medicines and vaccines you have given
- place of origin of the poultry
- visitors and deliveries to the establishment
- chemicals used in any protocol – include the name of the product, application date, concentration and targeted pathogen

Where medicines have a withdrawal period, you must keep a record of treatment dates and required withdrawal period for at least 5 years.

16. Facilities: PHS approved hatcheries and egg distribution centres

The hatchery must be physically and operationally separate from rearing facilities.

The layout must allow the following operations to be separated from each other:

- egg storage and grading
- egg disinfection
- pre-incubation
- incubation for hatching
- sexing and vaccination of day-old chicks
- preparation and packaging of chicks and goods for despatch

You must make sure that:

- buildings are protected against wild birds and rodents
- walls and floors are made of hard-wearing, waterproof and washable materials
- natural or artificial lighting, air flow and temperature systems are appropriate to the operation they are used for
- disposal of hatchery waste is hygienic

- equipment is smooth and has waterproof surfaces

The same principles apply to egg distribution centres with regards to facilities.

Hatchery operations

Hatcheries must operate on a one-way circuit for eggs and mobile equipment to prevent cross-contamination and maintain effective biosecurity.

Hatching eggs must only be:

- from PHS establishments
- from establishments in European Member States similarly approved in accordance with Regulation 2019/2035
- imported from Third Countries in compliance with Retained Regulation 798/2008

You must define hygiene rules in writing and agree them with the inspecting OV or APHA vet.

All personnel and visitors must wear appropriate protective clothing.

You must keep all buildings and equipment in good repair.

You must disinfect:

- eggs – between the time of their arrival at the hatchery and the incubation process or at the time of their dispatch for trade within Great Britain or export to the EU or Northern Ireland unless they have been previously sanitised at the breeding establishment of origin
- incubators – regularly
- hatchers and equipment – after each hatch
- chick boxes, containers and delivery vehicles – after each delivery or collection

The same principles apply to egg distribution centres with regards to operations.

Personnel at PHS approved hatcheries

Personnel must have appropriate skills and knowledge of hygiene techniques needed to prevent the spread of poultry diseases. This could be via specific training or through equivalent practical experience.

All visitors and staff working in the hatchery must be provided with appropriate protective clothing and are required to follow all relevant biosecurity protocols. In addition, staff must adhere to the hygiene code of conduct at all times.

The same principles apply to egg distribution centres with regards to biosecurity.

Record keeping

You must keep records for each flock for at least 3 years. Where possible, these records must include:

- place of origin of eggs
- date of arrival
- hatchability
- any abnormalities
- any laboratory tests and their results
- number and destinations of eggs not incubated or disposed of
- destinations of day-old chicks
- visitors and deliveries to the establishment
- medicine and vaccines you have given

For egg distribution centres, all the above must be recorded except for hatchability and medicine and vaccine use.

Where medicines have a withdrawal period, you must keep a record of treatment dates and their withdrawal period for at least 5 years.

17. Veterinary medicines

The Veterinary Medicine Regulations 2013 requires farmers to keep the records detailed in this section.

Owners and keepers of food-producing animals must keep records of all medical products acquired for those animals for 5 years.

When you acquire a medical product for a food-producing animal you must record the:

- name of the product
- batch number
- date of acquisition
- quantity acquired
- name and address of the supplier

When you, or a vet administers the medicine, you or the vet must record the:

- name of the product
- date of administration
- quantity administered
- withdrawal period
- identity of the animal you treated, for example, flock code or ID

If you dispose of a veterinary medicine, you must record:

- the date of disposal
- the quantity of product disposed of
- how and where you disposed of it

You can keep these records on paper or as a computerised record. There is no specified format. If you only keep an electronic copy of medicine records, you must supply a printed copy if requested by an inspector.

18. Vaccinations

You must only use vaccines that have a marketing authorisation issued by the Veterinary Medicines Directorate (VMD).

The use of vaccines imported under a VMD issued Special Import Certificate (SIC) or, autogenous vaccines manufactured and used with the approval of VMD, can be considered to have the equivalent of a marketing authorisation in the UK.

You must maintain records of all medicine usage, including vaccines. These records must be available for inspection.

Vaccination for Salmonella serotypes

Salmonella vaccination programmes must not:

- interfere with serological detection in the context of field investigation
- result in false-positive tests

You must not use live Salmonella vaccines in the framework of national control programmes:

- in breeding or productive poultry during their reproductive or laying stage unless the safety of their use has been demonstrated and they are authorised for such purpose
- where the manufacturer does not provide an appropriate method to distinguish bacteriologically wild-type strains of Salmonella from vaccine strains

Further information on veterinary medicines can be found at the Veterinary Medicines Directorate page.

19. Compliance with other legislation

As a PHS member, you must still comply with GB requirements and import requirements imposed by EU or non-EU importing countries, such as:

- UK legislation governing the production and marketing of poultry and poultry products
- Any European regulations for the trade with EU (such as Regulation (EU) 2016/429 and subsequent Commission delegated regulations)
- Any requirements for exports to any other importing non-EU country

20. Export certification by Official Veterinarians

All parties involved must treat any information shared with a members' private veterinarian as "Commercial-in-Confidence".

Private certification of poultry export consignments can only be carried out by an OV trained and authorised by APHA for export poultry work.

Members must make their own arrangements for all laboratory samples and tests required under the PHS, and for veterinary inspections, tests etc. required for trade.

Export certification usually requires a health examination of the birds for export and/or flocks of origin by an OV within a particular timeframe prior to export. In some cases, this requirement for the flocks may be waived if allowed by the receiving country (assessment of production records and health status of the flock still needed) provided the establishment is subject to routine monthly health inspections by an OV. It is the member's discretion to arrange and pay for these routine inspections to be able to make use of this option where it is permitted. Appropriate records of such inspections should be held at the establishment.

21. Processing of samples and testing methods

Samples collected must be subject to either:

i. serological testing in the case of birds sampled on farm

or

ii. bacteriological testing either as an alternative or in addition to serological testing referred to in point (i); however, samples for bacteriological testing must **not** be taken from poultry or eggs from poultry that have been treated with antimicrobial medicinal products during the three weeks prior to testing

Confirmatory testing must be capable of differentiating live vaccinal strains from field strains, i.e. serological responses to S. pullorum and S. gallinarum infection must be different from serological responses due to the use of Salmonella enteritidis vaccine, where this vaccine is used. Such vaccination must therefore be avoided if serological monitoring is to be used. **In cases where vaccination is used, testing shall be done using bacteriological methods.**

Note: There is currently no test that can differentiate between the response to Salmonella pullorum and gallinarum carrying status and vaccination for these serotypes.

Samples collected for bacteriological testing must be processed as follows:

- direct enrichment in selenite-cysteine broth for faecal/meconium and intestinal samples or other appropriate media suitable for direct enrichment of samples where competing flora is expected, followed by plating on a suitable plating agar, such as Brilliant Green Agar (BGA)

Other methods are permitted, but are less suitable for PHS-relevant Salmonella, such as:

- non-selective pre-enrichment followed by selective enrichment in soya-based Rappaport-Vassiliadis (RVS) broth or Müller- Kauffmann Tetrathionate-Novobiocin broth (MKTTn) for samples (such as embryos dead-in-shell) where competing flora is expected to be minimal

- direct plating of aseptically collected diseased tissues from infected birds on to a minimally selective agar, such as MacConkey agar

- Importantly, S. pullorum and gallinarum are not typically detectable in the modified semi-solid Rappaport Vassiliadis (MSRV) medium used for monitoring of zoonotic Salmonella spp. in the Salmonella National Control Programme, so must not be used to test for these organisms (but MSRV is suitable for the detection of Salmonella arizonae, see below)

22. Test results: when a flock is considered positive

A flock is considered positive when, following the positive results of the routine testing performed in accordance with the above, a second positive test of an appropriate type confirms presence of the disease agent. APHA must be notified.

23. General testing requirements

Samples must be collected and submitted either for either serological or bacteriology testing. Samples must be taken from farm or hatchery, or for hatchery and EDC hygiene control.

At hatcheries, sampling must represent every flock of origin present in a hatcher on the day of sample collection.

At farms, sampling must represent each flock present on the premises.

For the purpose of the PHS, ‘flock’ is defined as all poultry of the same health status kept on the same establishment or in the same enclosure and constituting a single epidemiological unit. In housed poultry this will include all birds sharing the same airspace.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory’s UKAS number or name here: UKAS accredited organisations.

The PHS member must pay the cost of all sampling and testing associated with the PHS.

24. Testing schedule for environmental (microbiological)

Quality control programme for purpose of environmental hygienic controls is required for hatcheries and egg distribution centres only, and must be in writing and agreed with the inspecting OV or APHA vet, and shall include the following:

a) environmental samples must be collected from the hatchery and egg distribution centres for bacteriological examination to monitor hygiene conditions. Swab samples to conduct aerobic plate counts (or similar tests) would be acceptable for this purpose.

b) samples must be taken at least every 6 weeks and each sampling must include a minimum of 60 samples. For smaller hatcheries, cumulative sampling over the 6-week period to a total of at least 60 samples are acceptable (to be agreed between the operator and the inspecting officer). Sampling must focus on surfaces and equipment that come into contact with egg trays and packaging.

25. Testing schedule for Salmonella pullorum or gallinarum

Routine testing on farm

Relevant species: fowl, turkeys, ducks, guinea fowl, partridges, pheasant, quails

Frequency/timing: Breeding flocks must be sampled at the point of lay. Productive poultry must be sampled during the production period and at least once a year.

Testing at the mid-stage premises may be omitted, if routine PHS testing is carried out at the destination premises, and the OV or APHA vet has documentary evidence confirming this.

For example, if flocks from a rearing premises are sampled at the destination breeding or production premises later in their life cycle, the OV responsible for approving the rearing premises must be provided with documentary evidence of routine PHS testing from the destination premises at the time of (re-)/approval. If such evidence cannot be provided, the flocks must be tested at the rearing premises. Sampling matrix:

Samples must be taken from each flock in the establishment keeping poultry, as appropriate:

- for serological testing:
    - For breeding, blood samples taken at or near point of lay taken at the rate shown in the sampling table in Annex 2.
    - For production, blood samples taken at least once a year during the production period
- for bacteriological testing:
    - post-mortem tissues, especially liver, spleen, ovary, oviduct and ileo-caecal junction
    - swabs from the cloaca of live birds, in particular from those birds that appear sick or that have been identified as highly seropositive
    - environmental samples (pooled caecal/faecal dropping samples) - these can be tested in pools of up to 10 individual droppings

The disease surveillance programme must be fully integrated between farm and hatchery. If testing is not carried out on farm, it is the responsibility of the farm operator to ensure that appropriate arrangements are in place for sampling and testing at the hatchery.

Testing methods:

- Preferred method: Serological testing is the recommended approach for on-farm surveillance of S. pullorum and gallinarum
- Alternative (or in addition) method: If conditions interfere with the validity of serological tests, bacteriology may be used as an alternative

For newly approved flocks carrying out their initial testing during the 6-week initial period (tests between 14 and 42 days), serological testing is preferred. The following sample types may be used:

- Blood samples for serological testing

  or

- Bacteriological testing of:
  - Faecal samples collected 4 weeks prior to any movement of birds or eggs off the premises (refer to sampling table in Annex 2 for the required number of samples)

    and

  - Cull chicks following placement on farm (up to a maximum of 60 chicks)

Samples for bacteriological testing must not be collected from poultry or eggs from poultry that have received antimicrobial medicinal products within the three weeks preceding the sampling or laying date.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory's UKAS number or name here: UKAS accredited organisations.

Routine testing at the hatchery

Relevant species: fowl, turkeys, ducks, guinea fowl, partridges, pheasant, quails

Frequency/timing: Samples shall be collected and examined at least once every 6 weeks.

Sampling matrix: The testing shall include at least:

- one pooled sample of meconium and down from each chick hatcher

  and

- a sample of **(20 in total)**:
    - either 10 second grade chicks and 10 dead-in-shell (embryos) chicks representing **every flock of origin** present in a hatcher on the day of sample collection

      or

    - 20 second grade chickens representing **every flock of origin** present in **a** hatcher on the day of sample collection

Test types available: S. pullorum or S. gallinarum

Test type	Additional information
Serology (monitoring only)	Rapid Serum Agglutination Test Tube Agglutination Test Other tests as developed e.g. ELISA, following validation as fit for purpose according to relevant ISO standards with UKAS accreditation
Bacteriology (monitoring and confirmation)	Culture using direct enrichment using selenite broth/ Brilliant Green Agar test is suitable for all samples BPW/RVS/BGA test suitable for non-faecal contaminated samples (for example dead-in shells and cull chicks). BPW-MSRV-based tests (as described by ISO 6579-1:2017 and used for the National Control Program (NCP)) are not suitable for S. Pullorum or S. Gallinarum but will detect the turkey pathogenic Salmonella arizonae O18 strain.
Molecular tests	Other tests as developed e.g. PCR, following validation and demonstration as fit for purpose according to relevant ISO 16140 series standards

Serological, bacteriological and molecular tests must be carried out using recognised validated methods.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory's UKAS number or name here: UKAS accredited organisations.

26. Testing schedule for Salmonella arizonae

Routine testing on farm

- Relevant species: turkeys only.
- Frequency/timing: Each flock must be tested at least once a year during the production period – the initial testing being at or near the point of lay.
- Samples: **bacteriology only**. There are no suitable serological tests. **Boot swabs** collected for testing under the Salmonella National Control Program (NCP) and the Poultry Health Scheme (PHS) are considered suitable for detection of the target S. arizonae strain. All samples must be tested in an official laboratory designated by the competent for the purpose of S. arizonae testing for the PHS. A list of official laboratories currently approved to undertake diagnostic testing for the purpose of the PHS can be found on GOV.UK at PHS Laboratories.

Routine testing at the hatchery

Samples shall be collected and examined at least once every 6 weeks in the hatchery. The testing shall include at least:

- one pooled sample of meconium and down from each chick hatcher

  and

- a sample of **(20 in total)** either:
  - 10 second grade chicks and 10 dead-in-shell (embryos) chicks representing **every flock of origin** present in a hatcher on the day of sample collection

    or

  - 20 second grade chicks representing **every flock of origin** present in a hatcher on the day of sample collection

Test types available: Salmonella arizonae

Test type	Additional information
Serology	• No test available
Bacteriology	• Culture • BPW-MSRV-based (as described by ISO 6579-1:2017 and used for NCP) test is suitable for S. arizonae, BPW – RVS enrichment or direct selenite enrichment methods can also be used.
Molecular tests	• Acceptability to be determined – no suitable validated test currently.

27. Testing schedule for Mycoplasma gallisepticum

Routine testing

- Relevant species: fowl, turkeys
- Frequency/timing for breeding poultry:
    - Fowl: at 16 weeks of age, at point of lay and then every 90 days during lay.
    - Turkeys: at 20 weeks of age, at point of lay and then every 90 days during lay.
- For productive poultry, samples must be collected during productive period, every 90 days.
- Testing method: either bacteriology and/or serology, and/or molecular testing. Culture is allowed but not recommended as it takes 21 days of culture to report a negative result.
- All samples must be tested in an official laboratory designated by the competent authority for the purpose of M. gallisepticum testing for the PHS. A list of official laboratories currently approved to undertake diagnostic testing for the purpose of the PHS can be found on GOV.UK at PHS Laboratories.
- Statistical sampling of each flock, as per table in Annex 2.
- Birds should be sampled at random from each part of the flock.
- Surveillance for clinical signs and lesions of Mycoplasma infection must be ongoing.

Samples for testing (both for routine monitoring and following suspicion of disease)

As appropriate –

- Serum
- Sperm
- Day old bird – air sac swabs from culls
- Tracheal, choanal, or air sac swabs
- On suspicion of infection, samples should be taken to confirm disease as follows:
    - from flocks, samples as per table in Annex 2
    - from day old chicks or poults
    - for examination for airsacculitis
    - samples from carcases according to veterinary advice and/or discussion of sampling options with APHA

Sample numbers: Statistical sampling

For the purposes of the PHS, a 'flock' is defined as meaning all poultry of the same health status kept on the same premises or in the same enclosure and constituting a single epidemiological unit. In housed poultry this will include all birds sharing the same airspace. Refer to the table in Annex 2 to determine the number of samples required – based on flock size – to achieve 95% confidence in detecting a 5% within-flock prevalence.

Test types available: Mycoplasma gallisepticum

Test type	Additional information
Serology (monitoring only)	Rapid Serum Agglutination test Western blot ELISA Haemagglutination inhibition
Bacteriology (for monitoring purposes)	Swabs from live birds or carcases; suitable types to include swabs taken from the trachea, the choanae or the cloaca

Bacteriology (for confirmatory purposes)	Swabs from live birds or swabs or tissues from carcases, especially air sacs from day-old chicks with lesions
Molecular tests (monitoring and confirmation)	Swabs from live birds or swabs/ tissues from carcases, especially air sacs from day-old chicks with lesions

Serological, bacteriological and molecular tests must be carried out using recognised validated methods.

All sample analysis for the PHS must be carried out at officially recognised PHS laboratories. These have been designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK. To check whether the specific test for the relevant pathogen is accredited, you can search with the laboratory's UKAS number or name here: UKAS accredited organisations.

The PHS member must pay the cost of all sampling and testing associated with the PHS.

28. Testing schedule for Mycoplasma meleagridis

Routine testing

- Relevant species: turkeys
- Frequency/timing: at 20 weeks of age, at point of lay and then every 90 days during lay.
- For productive poultry samples must be collected during production, every 90 days.
- Samples: either bacteriology and/or serology and/or molecular testing
- All samples must be tested in an official laboratory designated by the competent for the purpose of M. meleagridis testing for the PHS. A list of official laboratories currently approved to undertake diagnostic testing for the purpose of the PHS can be found on GOV.UK at PHS Laboratories.
- Statistical sampling of each flock, as per the table in Annex 2.
- Birds should be sampled at random from each part of the flock.
- Surveillance for clinical signs and lesions of Mycoplasma infection must be ongoing.

Samples for testing (both for routine monitoring and following suspicion of disease)

As appropriate –

- Serum
- Day old poults - air sac swabs from culls
- Tracheal, choanal, cloacal or air sacs swabs and swabs from reproductive tract (including oviduct, penis and semen of adult birds, as appropriate).
- On suspicion of infection, samples should be taken to confirm disease as follows:
    - from flocks, samples as per table in Annex 2
    - from day old poults for examination for airsacculitis
    - samples from carcases according to veterinary advice and/or discussion of sampling options with APHA

Annex 1 – Summary of PHS testing schedules

1. Farm testing schedule (Salmonella pullorum/gallinarum)

Applies to: all poultry species

Testing methods	Samples required	Number and frequency	Observations
Serology (preferred on farm) RSA; positives checked with SAT	Blood	**60 samples** per flock **At point of lay** (breeding poultry) **At least once/year during production period** (production poultry)	Species other than Galliformes may disclose high proportion of false-positive reactions to serological testing. Bacteriological testing either as an alternative or in addition to serological testing. Bacteriology testing to be used in flocks vaccinated against S. enteritidis (uncapable to differentiate serological response). Soya based Rappaport-Vassiliadis (RVS) medium not suitable for culture of S. pullorum/gallinarum. For **confirmatory testing**: Culture of post-mortem tissues (liver, spleen, ovary, etc.). For newly approved flocks: 2 tests within the first 6 weeks (at interval of 14-42 days). Clinical examination at least once during each laying or productive period.
Bacteriology (e.g. culture in selective enrichment medium) Selenite/BGA culture	Post-mortem (tissues from liver, spleen, ovary, oviduct and ileo-caecal junction) Environmental (pooled caecal/faecal droppings in pools of 10 or boot swabs) Cloacal swabs		

2. Hatchery testing schedule (salmonella gallinarum/pullorum)

Applies to: all poultry species

Testing method	Samples required	Number and frequency	Observations
Bacteriology (e.g. culture using selective enrichment methods) Selenite/BGA culture **or** BPW/MSRV/XLD	Meconium Down Second grade chicks Dead-in-shell chicks	**1 pooled sample of meconium and down** (per hatcher) AND **10 second grade chicks + 10 dead-in-shell chicks** OR **20 second grade chicks** **At least once every 6 weeks**	Sampling must be integrated with source/destination farm

2. Farm testing schedule (Salmonella arizonae)

Applies to: turkeys

Testing method	Samples required	Number and frequency	Observations
Bacteriology (e.g. culture in selective enrichment medium) Selenite/BGA culture **or** BPW/MSRV/ XLD & second medium with different biochemical indicator for S arizonae only	Post-mortem (tissues from liver, spleen, ovary, etc) Environmental (faecal droppings in pools of 10 or boot swabs) Cloacal swabs	**60 samples** per flock **At point of lay** (breeding poultry) **At least once/year** (production poultry)	Bacteriological only (serological tests are not suitable for S. arizonae) Samples tested using BPW-MSRV-based (as described by ISO 6579-1:2017 are suitable for S. arizonae detection) For **confirmatory testing**: Culture of post-mortem tissues (liver, spleen, ovary, etc.). For newly approved flocks: 2 tests within the first 6 weeks (at interval of 14-42 days). Clinical examination at least once during each laying or productive period.

3. Hatchery testing schedule (Salmonella arizonae)

Applies to: turkeys only

Testing method	Samples required	Number and frequency	Observations
Bacteriology (e.g. culture using selective enrichment methods) Selenite/BGA culture **or** BPW/MSRV/XLD& second medium with different biochemical indicator for S. arizonae only	Meconium Down Second grade chicks Dead-in-Shell chicks	**1 pooled sample of meconium and down** (per hatcher) AND **10 second grade chicks + 10x dead-in-shell chicks** OR **20 second grade chicks** **At least once every 6 weeks**	Sampling must be integrated with source/destination farm

4. Farm testing schedule (Mycoplasma gallisepticum)

Applies to: chickens and turkeys

Testing methods	Samples required	Number and frequency	Observations
Serology RSA / ELISA or any other validated test	Blood	**Statistical sampling (up to 60 samples)** per flock **Chickens (breeding):** At 16 weeks, point of lay and every 90 days during lay **Chickens (production):** Every 90 days **Turkeys (breeding):** At 20 weeks, at point of lay and every 90 days during lay **Turkeys (production):** Every 90 days	Bacteriological and molecular testing either as an alternative or in addition to serological testing. For **confirmatory testing**: Submission of carcases as per expert advice, in addition to routine sampling. Clinical examination at least once during each laying or productive period.
Bacteriology Bacteriological tests must be recognised validated methods	Sperm Swabs from trachea, choanae or cloaca Post-mortem tissues (especially air sac swabs from day-old culls)		
Molecular testing Molecular tests must be recognised validated methods			

5. Farm testing schedule (Mycoplasma meleagridis)

Applies to: turkeys only

<table>
<tr><th>Testing methods</th><th>Samples required</th><th>Number and frequency</th><th>Observations</th></tr>
<tr><td>Serology

RSA / ELISA or any other validated test</td><td>Blood</td><td rowspan="3">Statistical sampling (up to 60 samples) per flock

Turkeys (breeding): At 20 weeks, at point of lay and every 90 days during lay

Turkeys (production): Every 90 days</td><td rowspan="3">Bacteriological and molecular testing either as an alternative or in addition to serological testing.

For confirmatory testing: Submission of carcases as per expert advice, in addition to routine sampling.

Clinical examination at least once during each laying or productive period.</td></tr>
<tr><td>Bacteriology

Bacteriological tests must be recognised validated methods</td><td>Sperm

Swabs from trachea, choanae or cloaca

Post-mortem tissues (especially air sac swabs from day-old culls)

Swabs from oviduct and penis</td></tr>
<tr><td>Molecular testing

Molecular tests must be recognised validated methods</td></tr>
</table>

Annex 2 – Number of samples: Statistical sampling

For the purposes of the PHS, a ‘flock’ is defined as meaning all poultry of the same health status kept on the same premises or in the same enclosure and constituting a single epidemiological unit. In housed poultry this will include all birds sharing the same airspace.

Flock size	Number of samples required to provide 95% confidence of detecting 5% within-flock prevalence
1 to 19	All birds
20 to 29	20
30 to 39	25
40 to 49	30
50 to 59	35
60 to 89	40
90 to 199	50
200 to 499	55
500 or more	60

Annex 3 – Definitions

Term	Definition for the purpose of the PHS
APHA	Animal and Plant Health Agency, the executive agency of Defra providing delivery of field services and laboratory functions.
Breeding establishment	Premises producing hatching eggs for the production of breeding or productive poultry.
Breeding poultry	All poultry that is 72 hours old or more, intended for the production of hatching eggs.
Biosecurity	The sum of management and physical measures designed to reduce the risk of the introduction, development and spread of diseases to, from and within: (a) an animal population, or (b) an establishment, zone, compartment, means of transport or any other facilities, premises or location.
Day-old chicks	All poultry less than 72 hrs old.
Department	In England, the Department for Environment, Food and Rural Affairs (Defra). In Scotland, the Scottish Government. In Wales, the Welsh Government.
Depopulation	The destruction, subject to all the necessary health safeguards including disinfection, of all poultry and products at the outbreak site which are infected or suspected of being contaminated.
Disinfectants	For the purpose of pathogen control under the PHS, disinfectants must be at least Defra-approved.
Establishment	A facility or part of a facility which occupies a single site and is devoted to one or more of the following activities: (a) breeding establishment

	(b) rearing establishment (c) hatchery
Flock	All poultry of the same health status kept on the same establishment or in the same enclosure and constituting a single epidemiological unit. In housed poultry this will include all birds sharing the same airspace
Fowl	Birds of the species Gallus gallus
Hatchery	An establishment which collects, stores, incubates and hatches eggs for the supply of: (a) hatching eggs; (b) day–old chicks or hatchlings of other species;
Hatching eggs	All eggs laid by poultry or captive birds, intended for incubation.
Membership fee	The fee payable for annual membership to the PHS
NIPHAS	Northern Ireland Poultry Health Assurance Scheme
OV	Official Veterinarian, a private veterinary surgeon to whom the Department has delegated authority to inspect PHS premises and to carry out export health certification.
Poultry	As defined in Article 4 of Regulation 2016/429: “birds that are reared or kept in captivity for the production of a) meat; b) eggs for consumption; c) other products; d) restocking supplies of game birds; e) the purpose of breeding of birds used for the types of production referred to in points (a) and (b).

Productive poultry	All poultry 72 hours old or more, reared for the production of meat and/or eggs for consumption (or for restocking supplies of game).
PHS	Poultry Health Scheme
Ratites	Includes ostriches, emus, rheas, cassowaries, kiwis and tinamous.
Rearing establishment	(a) either a breeding poultry establishment rearing breeding poultry prior to the reproductive stage; or (b) a productive poultry rearing establishment rearing egg-laying productive poultry prior to the laying stage
Registration fee	A fee payable to APHA when first joining the PHS or when re-joining after membership has been revoked
S. gallinarum	Salmonella enterica, subspecies enterica, serovar Gallinarum biovar Gallinarum
S. pullorum	Salmonella enterica, subspecies enterica, serovar Gallinarum biovar Pullorum
S. arizonae	Salmonella enterica subspecies arizonae, serovar 18:z4, z32 (O18 Pathogenic arizonae serovar of turkeys)

Annex 4 - Useful links and contacts

Organisations and schemes	Contact information
APHA PHS contacts in Great Britain	Telephone 03000 200 301 or email CSCOneHealthPHS@apha.gov.uk
APHA Centre for International Trade: Carlisle	Telephone: 03000 200 301 or email general enquiries CITCarlisle@apha.gov.uk Centre for International Trade: Carlisle Eden Bridge House Lowther Street Carlisle CA3 8DX
British Egg Industry Council (BEIC)	British Egg Industry Council (BEIC)
British Poultry Council (BPC)	British Poultry Council (BPC)
British Veterinary Poultry Association	British Veterinary Poultry Association
Defra Approved Disinfectants	Disinfectants approved for use in England, Scotland and Wales
'Form Finder' page containing Export Health Certificates and associated documents	Find an export health certificate - GOV.UK
GB Import Health Certificates	Find a model health certificate for exports to GB - GOV.UK
Salmonella NCP	Salmonella National Control Programme Laying Hens and Flocks Broiler Flock Chickens

Organisations and schemes	Contact information
	Chicken Breeders Turkey Breeders Fattening Turkeys
Northern Ireland Poultry Health Assurance Scheme (NIPHAS)	NIPHAS
PHS Official Laboratories	Official laboratories designated by the competent authority to undertake testing for the purpose of the PHS. Laboratories must have UKAS accreditation for each specific test procedure used for PHS purposes. You can find a list of the PHS Laboratories on GOV.UK.
UK Accreditation Service (UKAS)	UKAS
Veterinary Medicine Directorate (VMD)	Veterinary Medicines Directorate
World Health Organisation for Animal Health (WOAH)	WOAH - World Organisation for Animal Health

Legislation	
Commission Regulation (EC) No 798/2008	Regulation 798/2008 is retained in the UK following exit from the European Union for import purposes [SI 2020/1462 Part 3 Regulation 53]
Commission Delegated Regulation (EU) 2019/2035	Regulation (EU) 2019/2035
Veterinary Medicine Regulations	The Veterinary Medicine Regulations 2013