Animal and Plant Health Agency

Certification scheme fruit plants

Explanatory guide to black currants, red currants, white currants and gooseberries (*Ribes*)

October 2023

Contents

1. Introduction

This document is a guide to *Ribes* certification taken from The Plant Health (Amendment etc.) (EU Exit) Regulations 2020 and The Plant Health (Phytosanitary Conditions) (Amendment) (EU Exit) Regulations 2020

2. Applications

The scheme is open to any grower in England and Wales who can meet the general conditions for entry and comply with the specific conditions of entry. Applications for entry of material to be submitted through the approved administrator, presently APHA. Please request an application form from FPCS.admin@apha.gov.uk

Growers will need to apply to APHA and be registered as producers under the scheme.

3. Labelling / sealing / supplier and variety registration

Refer to separate documents covering labelling, sealing, supplier and variety registration.

4. Grades and stock eligibility

New plantings:

GRADE	PARENT MATERIAL
Pre-basic mother plant	Any variety certified at Pre-basic or a candidate Pre-basic
Basic 1	Pre-basic
Basic 2	Pre-basic, Basic 1
Basic 3	Pre-basic, Basic 1, Basic 2
Certified	Pre-basic, Basic 1, Basic 2, Basic 3

Established stool beds:

GRADE	PARENT MATERIAL
Basic 1	Basic 1 grade in the previous year Beds eligible for a maximum of 6 years at Basic 1

Basic 2	Basic 1 or Basic 2 grade in the previous year Beds eligible for a maximum of 6 years at Basic 2
Basic 3	Basic 1, Basic 2 or Basic 3 in the previous year Beds eligible for a maximum of 6 years at Basic 3
Certified	Basic 1, Basic 2, Basic 3 or Certified in the previous year Beds have unlimited life at Certified provided all scheme conditions are met

5. Freedom from quarantine diseases

Crops must not be grown on land known to be infected with the following soil-borne diseases: Rhizomania, Strawberry red core or Verticillium wilt disease of hops or which is under notice for Potato Wart disease or Potato Cyst Nematode.

Growers who become aware of or suspect the presence of any quarantine disease must notify the Plant Health Inspector immediately.

6. Soil sampling requirements

Application for soil sampling should be made through the approved administrator.

Basic 1, Basic 2 and Basic 3

Soil sampling of the proposed field is required for freedom from the soil living virus vector nematodes prior to planting.

Longidorus elongatus

Longidorus macrosoma

Xiphinema diversicaudatum

Fields found to be infested cannot be used for planting unless one of the following requirements has been complied with:

a) Field treated with an approved soil fumigant

b) A soil bait test has been carried out for the relevant viruses. If the result is negative for virus the site can be utilised.

An alternative to soil sampling is crop rotation where no nematode host crop has been grown at the intended planting site for the last 5 years. For *Ribes* the relevant host crops are:

Grapevine, *Fragaria spp*., *Ribes spp., Rubus spp*., cherries, plums, apricot, peach, almond and Japanese plum and their rootstocks, poplar, walnut and olive trees, hops and elder/elderberry.

Certified Soil sampling is not required.

7. Isolation distances for field grown material

Stocks entered must be isolated by at least the distance shown in the following table. (metres)

	Basic 1	Basic 2	Basic 3	Certified	Approved - Health ♯	CAC	Fruiting
Basic 1	2	2	2	10	50	1500	1500 *
Basic 2	2	2	2	10	50	1000	1000 *
Basic 3	2	2	2	2	50	1000	1000 *
Certified	10	10	2	2	2	1000	1000 *
Approved-Health ♯	50	50	50	2	2	50	400 *
CAC	1500	1000	1000	1000	50	1	400
Fruiting	1500*	1000*	1000*	1000*	400*	400	0

* Gooseberries and jostaberries 100 metres from all other *Ribes*

8. Spacing

Stool beds: Not less than 2 metres between rows.

Nursery rows: Not less than 1 metre between rows.

For glasshouse production separation must be sufficient to ensure varieties of the same grade do not mix. Field isolation distances apply between different grades; exceptionally approval to reduce isolation distances may be granted from PHSI requiring additional precautions and standard operating procedures.

9. Aphid proof gauze house production and isolations

Specific conditions apply to the construction of an aphid proof structure for certification. Contact PHSI for full details. All material grown in the structure must be entered for certification. The propagator must, with consultation with PHSI, have a Standard Operating Procedure in place detailing the operation of the gauze house.

		Isolation for plants outside the gauze house (metres)				
		Basic 1 and Basic 2	Basic 3 and Certified	Approved -Health ♯	CAC	Fruiting
Plants grown inside gauze house	Pre-basic	100	100	200	200	500
	Basic 1 and Basic 2	2	2	100	100	250
	Basic 3 and Certified	2	2	50	50	100

10. Roguing

Limited roguing is permissible after inspection with prior approval of the APHA Plant Health Inspector. Records must be kept and made available of the stocks rogued, the reason for roguing and numbers of plants removed.

11. Bulking up

Stool bed extension (bulking up) is permissible providing the material used is eligible and bulking up takes place within the same field. Where stool bed numbers are increased using cuttings from the original planting then the whole stool bed must take the same age as the original planting. Growers must keep records of these procedures and make them available to inspector upon request.

12. Gapping up

Gapping up is permissible providing that the material used is eligible and prior approval from APHA Plant Health has been obtained. Growers must keep records and make them available if requested to do so.

13. Number of inspections

Pre-basic	Two inspections per year
Basic	One inspection only
Certified	One inspection only

Conduct the inspection at the optimum time depending on the season, usually between mid-June and mid-July.

14. Validity of certificates

Plants from certified crops may be described as being certified at the appropriate grade until 31 May in the year following certification.

15. Summary of pest and disease tolerances, sampling and testing

Testing

Pre-basic

Each pre-basic mother plant shall be sampled and tested four years after acceptance as a pre- basic mother plant and with subsequent intervals of four years for pests in Annex II and in the case of doubt for pests in Annex I.

Basic and Certified

Sampling and testing shall be carried out in the case of doubts concerning the presence of pests listed in Annexes I and II

Inspection

Pre-basic, Basic and Certified plant material shall be visually inspected and found free from pests and diseases listed in Annex I Part A and Annex II. Freedom can be met by removal of infected plants and /or by biological, physical or chemical treatments if applicable.

Pre-basic, Basic and Certified plant material infested by pests and diseases in Annex I Part B shall not exceed the tolerance levels indicated. Sampling and testing will be required if in doubt to the presence of those pests and diseases. Tolerances can be met by removal of infected plants and / or by biological, physical or chemical treatments if applicable.

Annex I

Annex I Part A		**Pre-basic, Basic and Certified**
Insects and mites		
Dasyneura tetensi	Black currant leaf midge	Nil tolerance at all grades
Ditylenchus dipsaci	Stem and bulb eelworm	
Pseudaulacaspis pentagona	White peach scale	
Quadraspidiotus perniciosus	San Jose scale	
Tetranycus urticae	Two spotted spider mite	
Cecidophyopsis ribis	Black currant gall mite	

Fungi *Sphaerotheca mors-uvae* Gooseberry mildew *Microsphaera grossulariae* Diaporthe strumella (Phomopsis ribicola)	Nil tolerance at all grades

Annex I Part B	**Pre-basic**	**Basic 1-3**	**Certified**
Nematodes			
Aphelenchoides ritzemabosi	0	0.05	0.5
Viruses			
Aucuba mosaic and black currant yellows combined	0	0.05	0.5
Vein clearing and vein net of black currant, Gooseberry vein banding	0	0.05	0.5

Annex II

Annex II	**Pre-basic, Basic and Certified**
Viruses as appropriate for the species concerned Arabis mosaic virus (ArMV) Black currant reversion virus (BRV) Cucumber mosaic virus (CMV) Gooseberry vein banding associated viruses Strawberry latent ringspot virus (SLRSV) Raspberry ringspot virus (RpRSV)	Nil tolerance at all grades

16. Record keeping (critical points plan)

The supplier must maintain relevant information to monitor the key points in the production process of all stocks entered for certification.

These include:

- Location and number of plants
- Timing of their cultivations
- Propagation operations
- Packaging, storage and transportation operations.

The information should remain available for at least three years and made available to PHSI upon request.

17. Requirements for pre-basic plant material

17.1 Eligible material

Any new or established variety or candidate material of potential new varieties can be entered. The progeny of pre-basic stock is eligible as parent material to produce basic grade cuttings or for planting stool beds for entry at basic 1.

17.2 Growing conditions

Candidate pre-basic mother plants must be kept under insect proof conditions and physically isolated from pre-basic mother plants until all tests have been successfully completed.

Pre-basic plants must have been maintained in a suitably designed insect proof gauze house containing only *Ribes* pre-basic plants. See section 9.

Strict precautions should be taken to prevent the introduction of any pest or disease in Annexes I and II.

All mother plants must be grown singly in sterilised growing medium and in individually labelled containers.

17.3 Pests and diseases

Mother plants must have been individually tested and found free from the diseases listed in section 18 every four years using the indicator plants or test methods described.

New plants to be entered into the pre-basic house must have been tested and found free from all diseases listed in section 18.

Any plants found to be infected with the diseases listed in section 18 or exhibiting suspicious symptoms should be removed immediately.

17.4 Documentation

The Person Responsible for the production of the plants must provide documentary evidence to show that the material has been produced under the conditions described above and that all the necessary tests were carried out and no evidence of infection was found.

This evidence must be provided to the purchaser of the pre-basic material before it can be used as parent material to produce basic 1 stool bed.

17.5 Trueness to type

Pre-basic material will be subject to trueness to type verification.

18. Required methods of testing for diseases for pre-basic *Ribes*

Black currant *(R. nigrum*): Reversion agent	Graft inoculation to black currant cv. Baldwin or Ojebyn
Vein banding agent	Graft inoculation to red currant cv. Jonkeer van Tets, black currant cv. Amos Black, hybrid *Ribes* EM 1385/81 or on gooseberry cv. Leveller
Strawberry latent ringspot nepovirus Arabis mosaic nepovirus Cucumber mosaic cucumovirus Raspberry ringspot nepovirus	Mechanical inoculation to test plants of *Chenopodium quinoa, Cucumis sativus* or *Nicotiana clevelandii* as applicable. They should be confirmed serologically.

Red currant (*R. rubrum*) and white currant *(R.sylvestre*): Reversion agent	Graft inoculation to black currant cv. Baldwin or Ojebyn
Vein banding agent	Graft inoculation to red currant cv. Jonkeer van Tets, black currant cv. Amos Black, hybrid *Ribes* EM 1385/81 or on gooseberry cv. Leveller
Strawberry latent ringspot nepovirus Raspberry ringspot nepovirus Arabis mosaic nepovirus	Mechanical inoculation to test plants of *Chenopodium quinoa, Cucumis sativus* or *Nicotiana clevelandii* as applicable. They should be confirmed serologically.

Gooseberry (*R. uva–crispa*): Vein banding agent	Graft inoculation to red currant cv. Jonkeer van Tets, black currant cv. Amos Black, hybrid *Ribes* EM 1385/81 or on gooseberry cv. Leveller
Arabis mosaic nepovirus Raspberry ringspot nepovirus	Mechanical inoculation to test plants of *Chenopodium quinoa*. They should be confirmed serologically.

Notes:

1. For graft inoculation tests, one indicator plant should be used for each virus being tested for. Tests for reversion should be observed for two years and tests on several shoots from large bushes are necessary because of the uneven distribution of this agent in the plant. For other pathogens tests should be continued until the following spring. Alternative indicator plants may be acceptable depending on the country of origin of the material, APHA should be consulted in such cases.

2. For mechanical inoculation tests on herbaceous indicators, plants should be observed for up to 4 weeks. Identification of specific viruses will require serological tests applied to extracts from the herbaceous indicators.

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Any enquiries regarding this publication should be sent to APHA.

http://www.gov.uk/apha