



Animal &  
Plant Health  
Agency

# United Kingdom Variety List Trials: Trial Procedures for Official Examination of Value for Cultivation and Use (VCU) Harvest 2022

## Field peas – spring and winter sown

Reviewed March 2022

### Changes from Harvest 2021 VCU procedures

1. C.6 Records, C.3.6.5: 1-9 scale added instead of day/month/year
2. D.1.1.4, Disease: 'must be reported as a percentage' removed from paragraph
3. Appendix 2 – Seed treatment: no treatment from 1 June 2021.
4. p25, Appendix 4 – Growing Trial Operators: additional trial operator: NIAB, North Yorkshire
5. Appendix 5 – Controls: Yield controls:
  - Prophet to Bluetime
  - Mascara to Karpate
  - Small blue taken out no comparison needed to Greenwood
6. Appendix 8 – 1-9 percentage scale removed

## Contents

Section A – General information .....	1
A.1 Purpose .....	1
A.2 Scope.....	1
A.3 Responsibilities .....	1
[OBJ]A.4 Summary of growing trials, tests and assessment procedures.....	4
[OBJ]Section B – Seed handling procedures .....	6
B.1 Responsibilities .....	6
B.2 Seed handling procedures .....	6
[OBJ]B.3 Authentication of VCU seed .....	7
[OBJ]Section C – Growing trial procedures.....	8
C.1 Responsibilities.....	8
C.2 Site suitability.....	8
C.3 Sowing the trial .....	8
C.4 Husbandry .....	10
C.5 Harvesting .....	11
[OBJ]C.6 Records.....	13
[OBJ]Section D – Disease testing procedures .....	17
D.1 Assessment of natural infection.....	17
D.2 Assessment of disease in inoculated trials .....	17
[OBJ]Section E – Quality test procedures .....	19
E.1 Responsibilities .....	19
E.2 Quality assessment methodology for obligatory and additional tests.....	19
[OBJ]Section F – Trial design and data handling procedures .....	21
F.1 Plan validation and storage.....	21
F.2 Data recording.....	21

F.3 Other tests and trials .....	21
Appendix 1 – Approved Trial Organisers/ Operators.....	22
Spring sown peas.....	22
Winter sown peas.....	22
Appendix 2 – Seed treatment and fungicide products for use on NL trials .....	23
Appendix 3 – Seed despatch deadline dates .....	24
Appendix 4 – Growing Trial Operators and Trial Locations .....	25
Spring sown peas.....	25
Winter sown peas.....	25
Pathology Trials Operator .....	25
Appendix 5 – Control varieties for VCU assessments .....	26
Spring peas .....	26
Winter peas .....	26
Appendix 6 – Dates by which records should be submitted .....	27
To Trials Organiser.....	27
Plot records to Data Handling Operator .....	27
Plot samples to Quality Testing Operator.....	27
Appendix 7 – Moisture content.....	28
Determination for yield .....	28
Oven method.....	28
Conditioning and electronic moisture meter method .....	28
Moisture meters .....	29
Appendix 8 – Disease assessment for pea .....	30
Downy mildew ( <i>Peronospora viciae</i> ) .....	30
Bacterial blight and mycosphaerellae .....	31

Appendix 9 – Phenological growth stages and BBCH-identification keys of pea ( <i>Pisum sativum</i> L.) .....	32
Principal growth stage 0: germination .....	33
Principle growth stage 1: leaf development.....	33
Principal growth stage 3: Stem elongation (main shoot) .....	33
Principal growth stage 5: Inflorescence emergence .....	33
Principal growth stage 6: flowering.....	33
Principal growth stage 7: development of fruit .....	34
Principal growth stage 8: ripening .....	34
Principal growth stage 9: senescence .....	34
Pea growth stages .....	35

# Section A – General information

## A.1 Purpose

A1.1 This document sets out the approved procedures to be used for growing trials, tests and assessments as required by the current **Protocol for Official Examination of Value for Cultivation and Use for Field Peas**.

## A.2 Scope

A.2.1 These procedures apply to all varieties of field peas.

## A.3 Responsibilities

### A.3.1 Procedures Development Group

A.3.1.1 The Procedures Development Group is responsible for reviewing these procedures annually and making any amendments for which it has responsibility, in accordance with the provisions of the **VCU Protocol**.

### A.3.2 Organisers and operators

#### A.3.2.1 Trials Organiser

British Society of Plant Breeders Ltd (BSPB)  
BSPB House  
114 Lancaster Way Business Park  
Ely  
Cambs.  
CB6 3NX

Tel No: 01353 653846  
Mobile No: 07747 567351  
Email [jeremy.widdowson@bspb.co.uk](mailto:jeremy.widdowson@bspb.co.uk)

A.3.2.2 The Trials Organiser is responsible for ensuring all **VCU Protocol** and **Procedures** requirements are followed and liaison with all Operators carrying out trials for National List purposes, including supply of seed and data handling.

#### A.3.2.3. Pathology Trials Operator

The Pathology Trials Operator appointed by APHA is responsible for carrying out inoculated trials for the assessment of disease in accordance with the **VCU Protocol** and these **Procedures**.

#### A.3.2.4 Data Handling Operator

The Data Handling Operator identified by the Trials Organiser is responsible for trial design and data validation in accordance with the **VCU Protocol** and associated **Procedures**.

#### A.3.2.5 Growing Trial Operators, Seed Handling Operators and Quality Testing Operators.

The Trials Organiser is responsible for potential Growing Trial Operators and Quality Testing Operators to carry out trials and tests as determined by the Procedures Development annual review in accordance with the **VCU Protocol**, and these **Procedures**. The Trials Organiser is also responsible for finding Seed Handling Operators who are able to carry out seed handling. Seed Handling Operators prepare trial seed for sowing on behalf of any Growing Trial Operator in accordance with the **VCU Protocol** and these **Procedures**.

A.3.2.6 A list of all approved Organisers and Operators is shown in [Appendix 1](#).

#### A.3.3 VCU Protocol and Procedures non-compliance

A.3.3.1 Where these procedures use the word “must or will” for any action then failure to carry out this action will result in non-compliance. Where the word “should” is used for any action then this is the method to be followed unless there are clear reasons not to, which can be justified by the operator as technically sound.

A.3.3.2 The Trials Organiser will forward any reports on **VCU Protocol or Procedures** non-compliance to APHA within 1 week of receipt. The Trials Organiser will obtain authorisation from APHA for any actions, including those necessary to remedy non-compliances, which are not within the requirements of the **VCU Protocol**. Such actions must be recorded as a non-compliance. Where emergency action is required and APHA staff are not available (e.g. evenings / weekends) the Trials Organiser should act, but report this to APHA at the earliest opportunity. Where GMOs are concerned the arrangements are as detailed in section 3.4.

#### A.3.4 Procedures for GM varieties

A.3.4.1 The National Authorities and Trials Organiser will develop procedures for GM varieties if an application for a GM candidate variety is received.

#### A.3.5 Processing of seed

A.3.5.1 The Trials Organiser is responsible for organising the processing of seed of candidate varieties submitted by the applicant, and seed of control, or other reference varieties, in accordance with the requirements set out in these **Procedures** and the current **VCU Protocol**. The Trials Organiser will ensure that any seed treatments or additives are approved for the purpose. Seed treatment products for use in NL trials are listed in [Appendix 2](#).

#### A.3.6 Dispatch of seed

A.3.6.1 The Trials Organiser will arrange for seed to be dispatched by the agreed deadlines to the Growing Trial Operators, and, for authentication, to address(es) supplied by APHA. Dates are given in [Appendix 3](#).

### **A.3.7 Monitoring of Growing Trial Operators and Seed Handling Operators - documentation**

A.3.7.1 The Trials Organiser will take any necessary action to enforce deadline dates and quality standards for required documentation.

A.3.7.2 The Trials Organiser will ensure Growing Trial Operators and Seed Handling Operators have access to all current protocols relevant to them and that they are notified of any amendments.

### **A.3.8 Seed quantities**

A.3.8.1 The Trials Organiser will determine the quantity of seed required for all VCU tests and trials in each annual series, including authentication, and will notify the applicant of quantities and delivery addresses.

### **A.3.9 Labelling of seed**

A.3.9.1 The Trials Organiser is responsible for ensuring all seed is clearly labelled with variety name/breeders' reference and AFP number.

### **A.3.10 Seed quality**

A.3.10.1 Seed submitted for VCU testing should meet the standards for the final generation of seed given in the appropriate seed regulations, in respect of germination, analytical purity and content of other seeds and any other impurities.

## A.4 Summary of growing trials, tests and assessment procedures

A.4.1 The number of trials and site locations are as detailed in Appendix 4.

A.4.2 Control varieties are listed in [Appendix 5](#).

A.4.3 The specified control varieties for quality assessments are grown only if there are candidates of the same type in the trial. The quality control varieties are not yield comparators.

A.4.4 The Trials Organiser is responsible for informing the Growing Trial Operators of the additional characters, which must be recorded as and when requested by applicants, and any samples that may be required for analysis.

### A.4.5 VCU Characters which may be assessed

**Bold = Obligatory**

*Italics = Additional if requested by the applicant*

Type of character	Reference	Description
Yield	Section C	<b>Plot yield</b> <b>Moisture content</b>
Behaviour with respect to factors in the physical environment.	Section C	<b>Standing ability</b> <b>Winter hardiness (winter peas only)</b> <i>Straw length</i> <i>Relative Maturity</i>
Resistance to harmful organisms	Section D	<b>Downy mildew resistance</b> <b>Bacterial blight resistance (winter peas only)</b> <b><i>Mycosphaerella</i> resistance (winter peas only)</b>
Quality characters	Section E	<b>Colour/grain type/TGW</b> are assessed by the DUS test centre <i>Protein Content</i>



#### **A.4.5.1 Further measurements**

The following must be measured or recorded in all trials, following procedures in Section C

**Sowing date**

**Plant population**

**Harvest date**

**Pre-harvest shedding**

**Bird damage**

**Combine losses**

**Plot size**

## **Section B – Seed handling procedures**

### **B.1 Responsibilities**

B.1.1 Seed Handling Operators/Growing Trial Operators are responsible for carrying out the following seed handling procedures.

### **B.2 Seed handling procedures**

B.2.1 Seed Handling Operators will receive a sowing list from the Trials Organiser, along with instructions as to which seed treatments or additives may be used. Seed treatment products for use in NL trials are listed in Appendix 2.

B.2.2 Seed Handling Operators/Growing Trial Operators must record receipt of seed from applicants by checking it off against the sowing list as it arrives. The Trials Organiser and Applicant should be notified of any damage to the packaging, loss of seed or identification problems within one working day of receipt.

B.2.3 Each Seed Handling Operator (or Growing Trial Operator if handling the seed) must retain a 200 gram untreated sample of the seed submitted of every variety in the trial, for authentication by the DUS test centre. However, if the seeds are small, the weight may be reduced further, provided the minimum number of seeds is 500.

B.2.4 Seed Handling Operators/Growing Trial Operators must record use of treatment chemicals in accordance with best practice and in full observance of all manufacturers' recommendations and relevant statutory obligations.

B.2.5 Any seed treatment equipment used must be fit for the purpose, properly calibrated, set up and operated in accordance with the manufacturer's recommendation.

B.2.6 Cross contamination must be avoided by ensuring equipment is clean between weighing and treatments.

B.2.7 A record must be kept of chemicals used and date of treatment.

B.2.8 Seed treatment should take place as near to the drilling date as possible.

B.2.9 Once seed has been treated, it must be kept safely until required for drilling and quality control. Each seed handling operator must retain a 100 gram sample of treated seed until one month after harvest.

## **B.3 Authentication of VCU seed**

B.3.1 The Trials Organiser will notify the minimum quantity required for authentication to Growing Trial Operators/Seed Handling Operators annually. Authentication samples are not required from Growing Trial Operators who receive seed from another Seed Handling Operator. All samples for authentication must be retained until harvest.

B.3.2 All samples must be kept under suitable conditions for the authentication procedures required and must be clearly labelled and sealed.

B.3.3 APHA will select samples from Growing Trial Operators/Seed Handling Operators for authentication at the DUS test centre.

B.3.4 Growing Trial Operators/Seed Handling Operators must send requested samples to the DUS test centre by the date specified by APHA.

B.3.5 Where there is more than one Seed Handling Operator, APHA will select samples for authentication from at least two Seed Handling Operators.

B.3.6 If the level of off types recorded in DUS tests or VCU authentication of a candidate exceeds 10%, the VCU tests will be considered invalid.

# Section C – Growing trial procedures

## C.1 Responsibilities

C.1.1 The Growing Trial Operators are responsible for conducting the trials according to these procedures.

## C.2 Site suitability

C.2.1 The Growing Trial Operator is responsible for providing a suitable site, which meets the following criteria:

C.2.2 Previous cropping must be appropriate for a field pea crop to be grown. For reasons of pests and diseases, the site should not have grown peas or other host plants i.e. field beans, broad beans, dwarf beans, vetches, tares or lupins over at least the preceding 4 years. Field peas must not be grown immediately following other legume crops. No trials should be grown in any field with a known history of *Sclerotinia*

C.2.3 Soil type should be typical of those on which field peas are grown locally. Soil fertility and texture should be uniform across the site. The soil should be sufficiently uniform to avoid variation in the growth of the trial

C.2.4 The trial should be sited away from trees, hedges, headlands and other features, which are likely to cause uneven growth or encourage grazing damage from wild fauna

C.2.5 The trial area should be cultivated in the direction of ploughing and drilled across the direction of ploughing and cultivation, such that each plot receives similar wheeling compaction. Cultivations should follow best local practice.

## C.3 Sowing the trial

### C.3.1 Plot size

C.3.1.1 The harvested plot area per variety must not be less than 15 m<sup>2</sup> for trials with 4 replications and 25 m<sup>2</sup> per replicate for trials with 3 replications. Plots must be drilled to a greater length than required and cutback to the required length prior to harvest. The plot width for calculating the harvested area is measured from outer row to outer row, plus half the inter-plot gap on either side. The allowance for the inter-plot gap must be no greater than 0.45 m.

### C.3.2 Plant population

C.3.2.1 The trials should be sown at a seed rate calculated to achieve a plant population of approximately 70 plants/m<sup>2</sup>. Seed should be treated against damping off and seed-borne infection of *Aschochyta* spp if an appropriate treatment is available. The Growing Trial Operator will determine the exact plant population as appropriate to local conditions. The following formula will be used to calculate the seed rate for a given thousand seed weight: -

$$\text{Seed rate (kg/ha)} = \frac{(\text{Target population} \times \text{Thousand seed weight}) \times 100}{(\text{Establishment \%} \times \text{Germination \%})}$$

The likely establishment % should be judged carefully depending on soil conditions and seedbeds. Growing Trial Operators are responsible for achieving the correct target populations.

### C.3.3 Trial layout

C.3.3.1 The Trials Organiser following consultation with APHA produces provisional sowing lists. The Trials Organiser, will make final sowing lists available along with the trial plans produced by the Data Handling Operator.

C.3.3.2 The trial must be sown according to the plan produced by the Data Handling Operator and may be an incomplete block design. In an incomplete block design, each replicate is split into a number of sub-blocks. The sub-blocks within a replicate must be sown adjacent to each other, as must plots within a sub-block. Any splitting of replicates must be between sub-blocks and not through sub-blocks. Varieties can be moved within a sub-block but must not be moved from their sub-block. Varieties must not be moved around within the plan e.g. if drilling errors occur. If plots are moved out of their original sub-block, they will have to be treated as missing plots. If there are any queries, please contact the Trials Organiser.

C.3.3.3 Buffer plots may be required in some instances; e.g. where there is a significant height difference between a variety or varieties. The Trials Organiser will advise if this is the case.

C.3.3.4 If there is a need to replace a planned variety e.g. if varieties are withdrawn, affected plots must be sown with any of the standard control varieties. Any such replacements must be agreed with the Trials Organiser. The control varieties are listed in Appendix 5.

### C.3.4 Drilling

C.3.4.1 Drills to be set up, calibrated and used only when conditions are right.

C.3.4.2 Care must be taken with drill settings and drilling speed to ensure satisfactory and uniform establishment and plant population from plot to plot. It is also important to ensure that there is no carry over of seed between plots.

C.3.4.3 At least **one row** of discard plot must be drilled on either side of the trial with the same drill and at the same time that the trial is drilled.

C.3.4.4 Precautions must be taken to avoid any missing rows. Any missing rows or parts of rows must be noted in the trial diary and reported to the Trials Organiser within one month of emergence.

### **C.3.5 Confirmation of trial layout**

C.3.5.1 After full establishment and within two months of sowing (autumn sown trials) or one month of sowing (spring sown trials) the Growing Trial Operator must confirm that the trial has been sown to plan or give full details of any changes to plan. This should be done by clearly highlighting the changes in the electronic plan and returning it to the Data Handling Operator.

- Return a completed site data 1 sheet including the following information:
- Site location details including how to get to the field.
- Sketch showing the layout of the trial in the field, in relation to other trials and showing access roads, gates, etc. The location of these features should utilise the navigation platform [What3Words.com](https://www.what3words.com)
- Trial sketch showing plot numbers and variety codes and/or names
- A short post-establishment report of the condition of the trial.

## **C.4 Husbandry**

### **C.4.1 Agronomy**

C.4.1.1 Where not specified in these procedures agronomy should follow best local practice, advisory and regulatory guidelines. Application of fertilisers and sprays should be uniform. It is normally best to apply these across the direction of the plots. Application wheelings should not run through the harvested plot area.

### **C.4.2 Fertiliser Application**

C.4.2.1 It must take into account inherent fertility, previous cropping, winter rainfall, the best local practice. All fertiliser applications should take account of the AHDB Nutrient Management Guide (RB209), the corresponding advisory publications in England, Wales, Scotland and Northern Ireland and past trialling experience.

### **C.4.3 Herbicides**

C.4.3.1 Any sensitivity to herbicides to be reported to the Trials Organiser.

### **C.4.4 Pest and Disease Control**

#### **C.4.4.1 Pest Control**

If necessary, appropriate means must be used to prevent or minimise damage by field mice, birds and other vertebrate and insect pests.

#### C.4.4.2 Disease control

All seed treatments applied should be according to Appendix 2.

Field pea trials are normally managed. In exceptional circumstances it may be necessary to deviate from the programme; eg additional sprays may be required during periods of extremely high disease pressure, or reduced rates may be required for drought stressed trials under low disease pressure. The Trials Organiser must be consulted before taking such a decision. Winter field pea trials should receive a fungicide according to Appendix 2.

#### C.4.5 Irrigation

C.4.5.1 Irrigation is only permitted to facilitate establishment.

#### C.4.6 Pathways

C.4.6.1 Internal pathways should be made before flowering and after the risk of pest damage has passed at approximately growth stage 16.

## C.5 Harvesting

#### C.5.1 Timing of harvesting

C.5.1.1 Date of harvesting will be determined by the Growing Trial Operator based on crop maturity and local weather conditions.

C.5.1.2 Plots should be trimmed to their final length prior to growth stage 16. Plot boundaries are parted prior to combining, if necessary. The plot dimensions must be measured prior to harvesting. If it is necessary to reduce the size of any plot at harvest, give clear details on the yield file. Individual harvested plot lengths must be recorded. Harvest date should be timed when the trial is ripe and reflect local practice. To minimise losses the combine should be driven into the direction of lodging so that the pods are on the combine table before the haulm is cut.

#### C.5.2 Harvesting method: Direct combining

C.5.2.1 Desiccation is frequently unnecessary, given satisfactory weed control, and should only be used as a last resort since it will affect relative maturities. The Trials Organiser should be consulted if desiccation is considered necessary.

#### C.5.3 Samples

C.5.3.1 No samples ex harvest are routinely required except for moisture determination when using the oven method. If other samples are required, they will be notified to the Growing Trial Operator by the Trials Organiser. It is essential that all samples:

- are representative of the variety/plot from which they are taken with minimal contamination. When sampling on-combine, it is essential to minimise the risk of contamination of grain from the previous plot.
- are taken from the same source.
- contain the weight of grain requested.

C.5.3.2 Moisture content samples must be assessed from every yield plot in the trial by the Growing Trial Operator. See appendix 7 for electronic and dry matter samples.

C.5.3.3 All bagged samples must be kept in good condition at a moisture content and temperature appropriate for safe long-term storage. They should be clearly marked both inside and outside the container/bag.

C.5.3.4 Samples may not be required from every variety, the Trials Organiser will provide details of which varieties require samples, the quantities required and the tests to be carried out.

C.5.3.5 Sample drying should be undertaken using a cold/warm air drier to reduce moisture content to 15% or below.

C.5.3.6 All plot samples must be labelled with the trial identification number, variety name/breeders' reference, AFP number, plot number and Growing Trial Operator identification number.

C.5.3.7 If Protein content is to be measured the Trials Organiser will request the appropriate sample. A sample of 500g from each plot must be taken at the time of plot weighing and sealed in a polythene bag. All harvest samples should be sent to the Quality Testing Operator as soon as practicable after harvest or after the completion of any drying where this is necessary. Notification of dispatch should be faxed or emailed to the Trials Organiser at the same time.

C.5.3.8 Where additional quality tests are requested by applicants, the Trials Organiser will provide appropriate instruction and labels. The samples should be dispatched to the appropriate Quality Testing Operator as soon as practical after harvest, or after completion of drying where necessary.

#### **C.5.4 Submission of data**

C.5.4.1 Appendix 6 lists the records, with deadlines, to be sent to the Trials Organiser. Diary sheets and other field records should be returned to the Trials Organiser within 5 working days

C.5.4.2 All plot records should be transmitted to the Data Handling Operator following the deadlines set out in Appendix 6. The Growing Trial Operator should ensure that data are free from errors before transmission. After scrutiny, copies of results will be returned to the Growing Trial Operator for action as agreed by the Trials Organiser.

C.5.4.3 All samples should be sent to the Quality Testing Operator following the deadlines set out in Appendix 6.



## C.6 Records

C.6.1 There are four components:

1. **Diary** Field notes of trial status.
- 2.\* **Site data part 1** Including full location details:
  - 1) a map of site location showing nearby settlements and roads
  - 2) a sketch showing the layout of trials in the field with access points and
  - 3) trial layout, showing plot numbers and variety codes/names.
- 3.\* **Site data part 2** Details of agrochemical applications and irrigation.
4. **Plot records** Plot data.

\*Template available from Trials Organiser

C.6.1.1 An entry in the Diary sheet should be made on every trials visit and any observations relevant to variety performance should be recorded. If the trial is in good condition, with no problems, this should be recorded.

### C.6.2 Plot records

C.6.2.1 Plot data may be recorded direct onto a data logger using a system approved by the Trials Organiser or recorded on paper then entered and validated onto a computer using an approved system. A system of ensuring that data are recoverable, in the event of loss of original data, must be implemented, e.g. copy and safe storage. Whichever method is used, individual plot data will only be accepted by the Data Handling Operator in an approved format using the variety names and units as listed in sections C and D.

C.6.2.2 All observations should be checked at the time of recording to ensure that they lie within acceptable limits for the character recorded. Observations that have been designated as exceptional by the recorder should be identified with a note on the approved data file or hard copy medium describing the possible causes together with a recommendation for their exclusion or inclusion in the trial analysis.

C.6.2.3 Plot numbers on record sheets must correspond with the numbering on the field plan.

C.6.2.4 If a character is not recorded or is missing the Growing Trial Operator should indicate in the diary or on the recording sheet the reason why it has been excluded.

C.6.2.5 Where a plot record is missing the Growing Trial Operator should enter “\*” in the approved data file or hard copy medium and, unless the non-recording of the plot has already been agreed with the Trials Organiser, append a note to the file explaining why a missing value has been entered for that plot. The Growing Trial Operator should not enter “0” for missing plots.

C.6.2.6 Specific plot records should be made as counts or on the scales shown for each character. Only the character names as listed may be used. All records should be returned to the Data Handling Operator as soon as possible after they are completed.

C.6.2.7 All records should be returned as soon as reasonably possible and when complete for the whole trial. Indicative deadlines are given in Appendix 6. All records must be returned by the final deadlines.

### C.6.3 Procedures for recording Characters

The following procedures must be followed for measuring all characters to be used in NL decision making.

#### C.6.3.1 PLOT YIELD AND MOISTURE CONTENT (OBLIGATORY) (kg)

The plot seed yield must be recorded, and returned with details of harvested plot dimensions, the growth stage and moisture content to the Data Handling Operator within 5 days of harvesting the trial.

The following information must accompany the yield data:

The moisture content % of the harvested grain, determined either by oven or an approved electronic method. See Appendix 7.

Plot length: the plot length harvested in metres.

Plot width: the width of the harvested plot in metres from outer row to outer row plus half of the inter-plot gap on either side. The allowance for the inter-plot gap should be no greater than 0.45m.

If these are not the same for every plot, a separate record must be submitted

#### C. 6.3.2 STANDING ABILITY (OBLIGATORY) (1-9)

1 very poor  
9 very good

This must be recorded at harvest time i.e. maturity. Poor standing ability in peas is generally a combination of progressive sinking and leaning of the haulm.

#### C.6.3.3. WINTER HARDINESS (for Winter Sown peas only) (OBLIGATORY) (1- 9)

This should be recorded after any period of adverse weather on the scale 1-9 where 1 = total loss of plant and 9 = no damage.

#### C.6.3.4 STRAW LENGTH (ADDITIONAL) (cm)

Record average plot height after the end of flowering before leaning or lodging takes place (if practical take 3 measurements along the length of the plot). If lodging has occurred, choose a representative area of the plot, lift a number of plants against the measuring pole and record an average height.

C.6.3.5 *RELATIVE MATURITY* (ADDITIONAL) (1-9)

Relative Maturity should be judged with a visual estimate of crop senescence and moisture content 18 to 20%, where;

- 1 = latest maturing
- 9 = earliest maturing

This should be recorded when each plot is harvested.

C.6.3.6 **SOWING DATE** (OBLIGATORY) (Day/Month/Year)

This is recorded in Part 1 of the Site Information Form.

C.6.3.7 **PLANT POPULATION** (OBLIGATORY) (1-9)

Where there is evidence of poor establishment, at a level which will affect results, plant counts should be taken soon after full emergence.

- 9 = Target population

Records will be converted and stored as number of plants per m<sup>2</sup>.

C.6.3.8 **HARVEST DATE** (OBLIGATORY) (Day/month/year)

C.6.3.9 **BIRD DAMAGE** (OBLIGATORY) (1-9)

- 1 severe damage
- 9 no damage

Records of bird damage which affects the yield of the trial should accompany the yield data.

C.6.3.10 **COMBINE LOSSES** (OBLIGATORY) (1-9)

This must be recorded.

9 = no combine losses. Combine losses should be assessed if the losses are thought sufficient to exclude the yield data from results. Indicate the estimated number of peas lost per m<sup>2</sup> for the lowest score given on the 1 to 9 scale.

C.6.3.11 **SHEDDING** (OBLIGATORY) (1-9)

This must be recorded.

9 = no shedding. Shedding should be assessed if the losses are thought sufficient to exclude the yield data from results. Indicate the estimated number of peas lost per m<sup>2</sup> prior to harvest for the lowest score given on the 1 to 9 scale.

#### **C.6.4 Site factors**

Any observations of the trial, which may be having an effect on the plots, must be recorded.

Where varietal differences are seen in pest or disease attack, records should be made in accordance with the procedure in Section D for disease.

Records for other scores should be taken as % plants affected or on a 1 to 9 scale. Include definitions for each rating on the 1 to 9 scales.

#### **C.6.5 Trial inspection**

All trials will be inspected by the Trial Inspection and Technical Validation Operator and, in some cases, it may be necessary to visit on more than one occasion. To aid inspection in the absence of the trial officer, as a minimum plot 1 should be clearly labelled.

The requirements for Growing Trial Operators in respect of inspections are as follows:

1. To give reasonable access to trials to inspectors and provide full location and site details (if not already given with site data 1).
2. To supply the inspector with information (for example sprays applied etc) within seven days of a request.
3. To co-operate with the inspector in making any non-routine assessments required to establish the validity of the trial (for example population counts).
4. To carry out any action agreed in consultation with the inspector. In particular, it is important that any requirement to shorten plots is undertaken. The data on plots that the trials operator and inspector agree to exclude should not be submitted.

## Section D – Disease testing procedures

### D.1 Assessment of natural infection

The Growing Trial Operator is responsible for carrying out these procedures.

#### D.1.1 Diseases recorded

D.1.1.1 The precise timing for assessment is best judged in relation to the development of disease in the trial, with the aim being to achieve the assessment, which shows the most differentiation between varieties. In practice, this usually means that two or three sequential assessments are necessary. If disease infection persists, numerical records should be made throughout the season.

D.1.1.2 Diseases should be recorded routinely when the level of infection on the most affected variety is over 5% of the leaf or stipule/tendrill area.

D.1.1.3 Downy mildew should be assessed using the key given in Appendix 8. It must be recorded and reported as a percentage

D.1.1.4 Bacterial blight and *Mycosphaerella* should be recorded in winter sown trials only using the key in Appendix 8.

All replicates in the trial should be recorded.

Assessments must be made on a “whole plot” basis, i.e. by making an overall assessment of the average % infection on all plants in a small (approx. 1m<sup>2</sup>) area of the plot and repeat at a minimum of 3 points in each plot. Where primary foci of high infection occur, these must be averaged over the whole plot; e.g. a primary focus of 50% infection occupying 5% of the plot must be recorded as 50% x 5% = 2.5%.

### D.2 Assessment of disease in inoculated trials

The Pathology Trials Operator is responsible for carrying out these procedures.

#### D.2.1 Diseases recorded

D.2.1.1 The disease recorded is downy mildew.

D.2.1.2 The pathogen is maintained on seedlings. Conidia are increased to provide inoculum for mixtures of current susceptible varieties sown in polythene tunnels. These are irrigated to promote cycles of infection of downy mildew.

### D.2.1.3 Inoculation

The resulting infected material is incorporated into the soil of the polythene tunnel area as it senesces, thus introducing a high and uniform level of oospore inoculum into the soil. The resulting area is planted directly with trial plots (1.5m lines, 4 replicates per variety) the following year, and collections of downy mildew isolates are made from naturally occurring field infections whenever possible and added to the seedling maintenance system. Tunnel areas are “updated” with new populations on a rotational system. It is not generally possible to use one tunnel area for more than three consecutive years, and new areas need to be developed as required.

D.2.1.4 Assessment of developing systemic and secondary infections are assessed using the key in Appendix 8.

# Section E – Quality test procedures

## E.1 Responsibilities

The Quality Testing Operators are responsible for conducting the approved quality tests according to these procedures.

## E.2 Quality assessment methodology for obligatory and additional tests

### E.2.1 Preparation of samples prior to quality analysis

E.2.1.1 Samples should be:

- Relatively weed free
- Free from excessive numbers of broken grains
- Bright and of good colour
- Well filled and free from visual sprouting.

E.2.1.2 Sample cleaning

The samples should be cleaned to remove combining debris such as straw, chaff, and unthreshed pods and weed seeds. The cleaning may be by hand or with hand-held or mechanical sieves.

E.2.2 Quality tests

#### E.2.2.1 COLOUR/GRAIN TYPE/TGW (OBLIGATORY - to be done by the DUS centre)

The variety must be classified by the DUS Test Centre into seed categories as follows. The DUS Test Centre will also determine TGW.

<b>Blues</b>	White Flowered, with round, smooth, Blue/Green grain
<b>Whites</b>	White Flowered, with round, smooth, White/Yellow grain
<b>Marrowfats</b>	White Flowered, with irregular shaped/dimpled, Green grain
<b>Maples</b>	Coloured Flowered, with Brown/Flecked grain

This categorisation is used to confirm the entry made by the applicant in the variety application Technical Questionnaire.

## **E.2.2.2 PROTEIN CONTENT DETERMINATION**

### **E.2.2.2.1 Hammer milling of grain prior to analysis**

The mill must be a hammer mill fitted with a 1mm screen. 300g of sample is milled and the material must be totally removed from the receptacle. The sample must be spread thinly, either with a printer's roller or with a wide blade spatula. The sample must be re-formed into a pile and the process repeated four times. After mixing, a representative sub-sample must be taken in the following manner:-

- A sample jar of 250ml capacity should be filled in small stages re-mixing the bulk between stages and blending each stage within the jar.
- The sample jar must be filled and then sealed with a close-fitting lid.

### **E.2.2.2.3 Determination of Crude Protein or Total Nitrogen Content**

Determination of protein content by NIR spectroscopy is permissible provided that the calibration utilised is commercially available and specific for pulse protein content measurement. Quality assurance of the analytical procedure must include regular analysis of a suitable test material – for example a sample of grain maintained for that purpose. Records should be kept to demonstrate that the instrument is performing correctly.

Each season, approximately 10% of the samples analysed by NIRS, should be analysed by a chemical method (as described below – E.2.2.2.4) to check the precision of the NIRS protein prediction. If appropriate and when sufficient variety, site and year chemical data is available, the NIRS calibration should be biased appropriately to improve the precision of the prediction.

E.2.2.2.4 Determination of Crude Protein or Total Nitrogen Content must be by a chemical method, recognised by competent authorities (IBD, AOAC, ISO, etc) and which makes direct measurement of nitrogen content.

E.2.2.2.5 Methods acceptable to the National Authorities are currently total nitrogen determined by the Kjeldahl method and total nitrogen using the Dumas method. These methods are only acceptable where instrumentation used is capable of analysing sample sizes greater than 0.5g.

E.2.2.2.6 Quality assurance of the analytical procedures must include regular analysis of a suitable test material - for example, a sample of flour maintained for that purpose.

E.2.2.2.7 Systematic errors in Kjeldahl nitrogen analysis must be controlled by the inclusion of blank analyses and by the analysis of a suitable analytical standard (Ammonium Sulphate, Methionine in a suitable bulking agent) for which the nitrogen content is known.

E.2.2.2.8 Instrument drift in Dumas nitrogen must be controlled by standardisation against a suitable analytical standard (EDTA, Glycine), for which the nitrogen content is known.



# Section F – Trial design and data handling procedures

## F.1 Plan validation and storage

F.1.2 After the trial has been drilled, the Growing Trial Operator must

- a) Confirm that the trial has been drilled according to plan and provide the sowing date to the appropriate Data Handling Operator.
- b) If any amendments have been made, return a hard copy of the plan to the appropriate Data Handling Operator with any amendments clearly indicated. Alternatively, amendments may be notified electronically with the agreement of the Data Handling Operator.

F.1.3 The Data Handling Operator will check these for statistical validity and, once this has been done, will load the plan on the database.

## F.2 Data recording

F.2.1 Data are recorded for the characters and using the methods given in Sections C, D and E.

F.2.2 Site information is recorded for each trial including, for example, data on previous cropping, seed rates, soil details and fertiliser applications.

F.2.3 Details of any agrochemical applications are also recorded and retained by the Growing Trial Operator.

## F.3 Other tests and trials

F.3.1 Any additional or alternative designs required for the assessment of additional VCU characters not detailed in Appendix 3 of the **VCU TRIAL PROTOCOL** for field peas will be added to these **Procedures** as and when approved by the NLSC.

# Appendix 1 – Approved Trial Organisers/ Operators

## Spring sown peas

Activity	Organisers/operators responsible
Data Handling Operator	BioSS
Trials Organiser	BSPB
Pathology Trials Operator	NIAB
Trial Inspection Operator	PGRO
Technical Validation Operator	NIAB
Quality Testing Operators	NIAB/SASA
Data Review and Standards Setting Operator	NIAB

## Winter sown peas

Activity	Organisers/operators responsible
Data Handling Operator	NIAB
Trials Organiser	BSPB
Growing Trial Operator	NIAB
Seed Handling Operator	NIAB
Pathology Trials Operator	NIAB
Trial Inspection	PGRO
Technical Validation Operator	NIAB
Quality Testing Operators	NIAB/SASA
Data Review and Standards Setting Operator	NIAB

## **Appendix 2 – Seed treatment and fungicide products for use on NL trials**

To be advised

## Appendix 3 – Seed despatch deadline dates

VCU seed must be delivered to each Seed Handler by:

Spring sown peas – 31 January

Winter sown peas – 1 October

Authentication samples, if requested, must be delivered to the appropriate DUS Test Centre.

## Appendix 4 – Growing Trial Operators and Trial Locations

### Spring sown peas

Growing Trial Operator	Seed Handling Operator (If not Trial Operator)	Location of trial
NIAB		Kent
PGRO		Lincolnshire
Darlow Contract Services		Cambridgeshire
NIAB		Hampshire
NIAB		North Yorkshire

### Winter sown peas

Growing Trial Operator	Seed Handling Operator (If not Trial Operator)	Location of trial
NIAB		Cambridgeshire

### Pathology Trials Operator

Pathology Trials Operator	Location of Trial
NIAB	Cambridge

# Appendix 5 – Control varieties for VCU assessments

## Spring peas

### Yield controls

Bluetime  
Karpate

### Quality controls

Sakura (for comparison with marrowfat pea candidates)  
Rose (for comparison for pigeon feed market)  
Greenwood

### Pathology benchmark varieties

Downy Mildew: Kahuna  
Bibao  
Maro

## Winter peas

TBC following an application

## Appendix 6 – Dates by which records should be submitted

### To Trials Organiser

Record	Latest date of receipt by Trials Organiser
Site data part 1 (including site sketch)	Within 2 months of drilling trial (autumn sown trials). Within one month of drilling trial (spring sown trials)
Site data part 2	By the time trials are harvested
Plot records (in approved electronic format)	Growing Trial Operator should notify Trials Organiser that trial has been harvested within 2 days of harvest

### Plot records to Data Handling Operator

Record	Date
Plot records SHOULD be sent to Data Handling Operator	Within 10 days of record being taken

### Plot samples to Quality Testing Operator

Samples	Date
Plot samples for quality testing SHOULD be sent to the Quality Testing Operator	Within 2 days of harvest

# Appendix 7 – Moisture content

## Determination for yield

Moisture content % of harvested material enables yield at 15% moisture content to be calculated.

This can be determined by either of two methods.

1. The Oven Method. Here the sealed sample taken at harvest is dried in an oven until no more moisture can be removed. The dried weight is then recorded and by comparison with the pre-dried sample weight, moisture content can be calculated.
2. Harvesting and conditioning of each plot and then reweighing and measuring moisture content electronically.

## Oven method

The following procedure must be followed:

1. A fully representative sub-sample of approx 500 grams is weighed to 1 decimal place and then placed in the drier, which must be at a temperature of  $100^{\circ}\text{C} \pm 4^{\circ}\text{C}$  with the air recirculator set in the range 80-100% recirculation in order to restore the temperature to  $100^{\circ}\text{C} \pm 4^{\circ}\text{C}$  as rapidly as possible. When the temperature is restored to  $100^{\circ}\text{C} \pm 4^{\circ}\text{C}$  the air regulator is set at 80% recirculation i.e. 20% fresh hot air. The regulator is critical for rapid drying. The samples are dried at  $100^{\circ}\text{C} \pm 4^{\circ}\text{C}$  for such time as is necessary for complete drying.
2. The dried sample is carefully removed from the drier as soon as the sample is cool enough for accurate weighing. The dry weight is recorded to one decimal place.
3. When all samples from a given trial have been recorded, the moisture content % must be immediately reported to the Data Handling Operator electronically using the character names given in Section C 6.3

## Conditioning and electronic moisture meter method

### Conditioning

1. Each plot must be harvested, and the entire produce put into clean sacks or other suitable containers, labelled and sealed. The grain parcels should then be dried using a cold/warm air drier where the drying temperature is not in excess of  $60^{\circ}\text{C}$ .
2. The grain should be dried for such time as is necessary to reach equilibrium with their surroundings. The parcels should then be weighed, and the moisture content recorded using an appropriate electronic moisture meter as set out below. The moisture content after drying must not exceed 17%.



3. The Growing Trial Operator returns the weight and moisture content to the Data Handling Operator.

## Moisture meters

### Principles

Moisture meters may only be used for the measurement of grain moisture below 17%. There are no restrictions on the make or model of moisture analyser that may be used, provided the conditions described below are met.

The manufacturer's recommendations for use must be followed. On-combine analysis is not approved, as currently no model is sufficiently accurate over the likely range of moisture contents.

### Equipment

The analysing equipment must:

- Be calibrated at least once annually for each crop according to the manufacturer's instructions using check samples (see reference below) and have a moisture content accuracy of plus/minus 0.5%. The calibration data should be retained for a minimum of 1 year.
- Be serviced regularly, especially just prior to harvest, according to manufacturer recommendations. The action taken should be documented and the information held for a minimum of 1 year.
- Be fit for use in accordance with manufacturer instructions. It should have an adequate power supply throughout operation. Instructions should be held with the machine and all operators adequately trained in its operation.

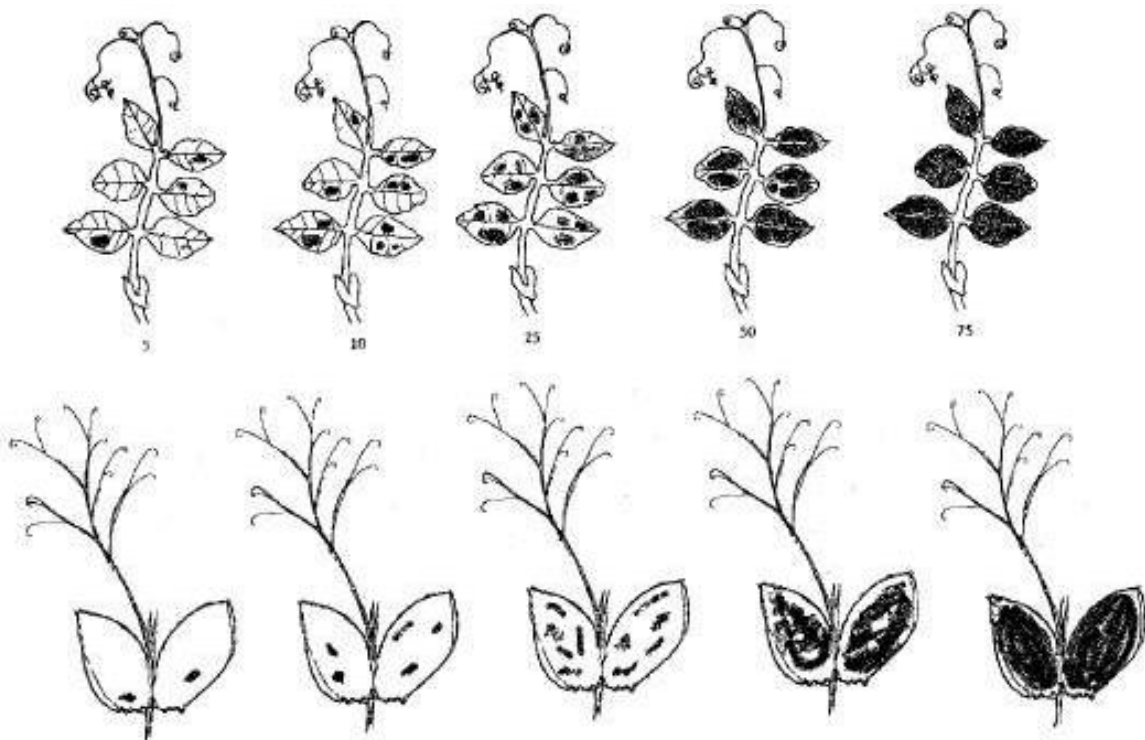
### Measuring moisture in conditioned grain

- The grain samples to be analysed must be between 12 to 17% moisture content.
- The grain to be analysed must be fully ripe. In other cases, the samples for the oven method should be used.
- The data must be in the form of moisture content %.

References: BS 4317-24:1990, BS 7700/1-2008 Methods of test for cereals and pulses. Method of checking the calibration of moisture meters for cereals.

## Appendix 8 – Disease assessment for pea

### Downy mildew (*Peronospora viciae*)



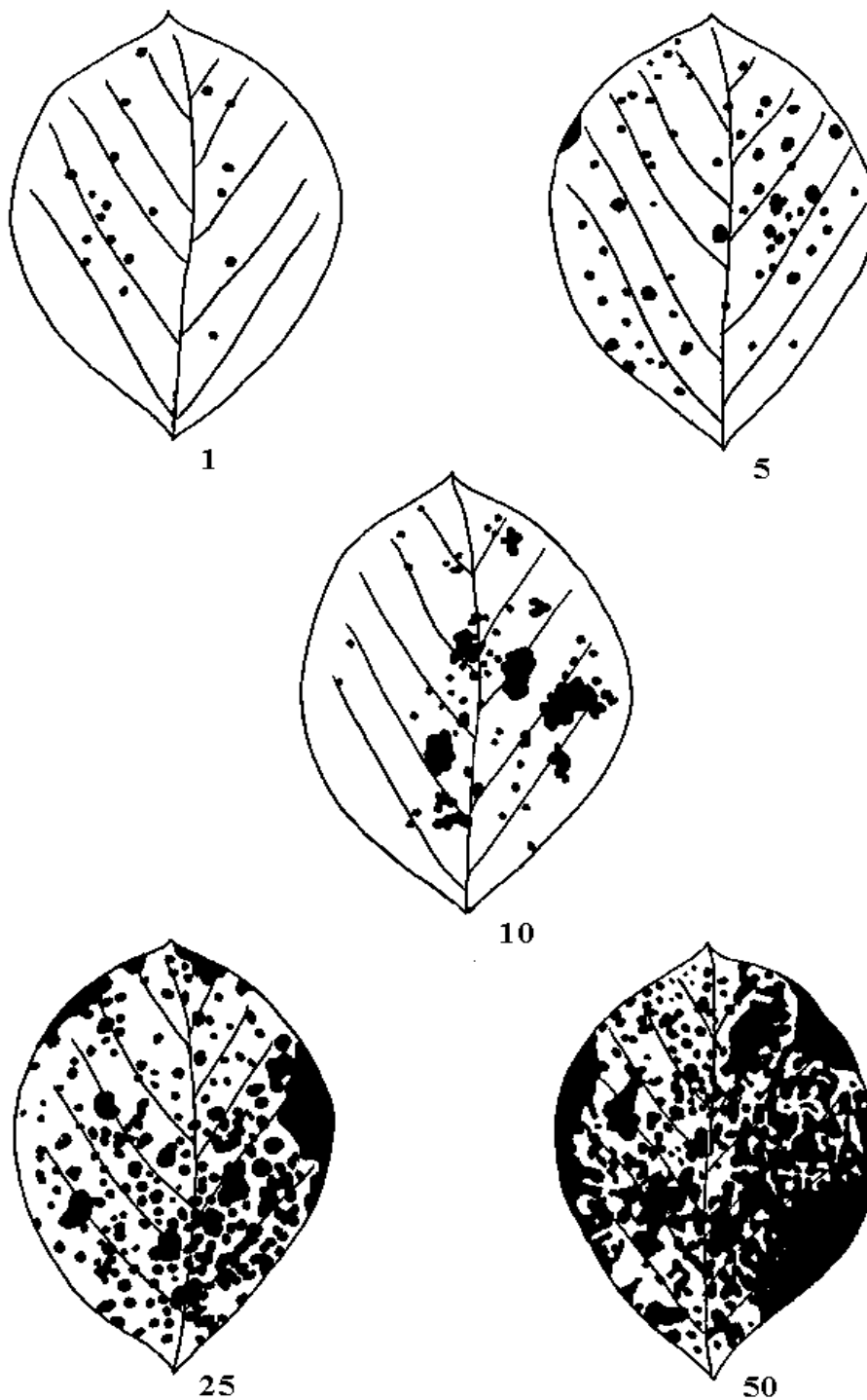
**Percentage area infected**

**(0 = Healthy)**

**Notes on assessment**

- 1. Examine all leaves in 3 areas of each plot.**
- 2. Include all chlorosis and necrosis attributable to downy mildew.**
- 3. Where isolated foci of high infection occur, they should be averaged over the area as a whole.**
- 4. Estimate % infection using the illustrations above as a guide. Interpolate values if necessary. Record the average % infection per plot from the 3 areas.**

## Bacterial blight and mycosphaerella



Examine all the leaves on plants within 4 to 6 areas of each plot, each area being approximately  $1\text{m}^2$ . Assess the mean % leaf area infected with disease.

## Appendix 9 – Phenological growth stages and BBCH-identification keys of pea (*Pisum sativum* L.)

Weber and Bleiholder, 1990; Feller et al., 1995 b

The extended BBCH-scale is a system for a uniform coding of phenologically similar growth stages of all mono- and dicotyledonous plant species. The decimal code, which is divided into principal and secondary growth stages, is based on the well-known cereal code developed by ZADOKS et al. (1974) in order to avoid major changes from this widely used phenological key.

- The general scale forms the framework within which the individual scales are developed. It can also be used for those plant species for which no special scale is currently available.
- Similar phenological stages of each plant species are given the same code
- For each code, a description is given, and for some important stages, drawings are included
- For the description of the phenological development stages, clear and easily recognised (external) morphological characteristics are used.
- Except where stated otherwise, only the development of the main stem is taken into consideration
- The growth stages refer to representative individual plants within the crop stand. Crop stand characteristics may also be considered.
- Relative values relating to species- and/or variety-specific ultimate sizes are used for the indication of size.
- The secondary growth stages 0 to 8 correspond to the respective ordinal numbers or percentage values. For example, stage 3 could represent: 3rd true leaf, 3rd tiller, 3rd node or 30% of the final length or size typical of the species or 30% of the flowers open.
- Post-harvest or storage treatment is coded 99.
- Seed treatment before planting is coded 00.

The entire developmental cycle of the plants is subdivided into ten clearly recognizable and distinguishable longer-lasting developmental phases. These principal growth stages are described using numbers from 0 to 9 in ascending order. Owing to the very many different plant species there may be shifts in the course of the development or certain stages may even be omitted. The principal growth stages need not proceed in the strict sequence defined by the ascending order of the figures, but can occasionally also proceed in parallel.

## Code Description

### Principal growth stage 0: germination

1. Dry seed
2. Beginning of seed imbibition
03. Seed imbibition complete
05. Radicle emerged from seed
7. Shoot breaking through seed coat
8. Shoot growing towards soil surface, hypocotyl arch visible
9. Emergence: shoot emerges through soil surface (“cracking stage”)

### Principle growth stage 1: leaf development

10. Pair of scale leaves visible
  11. First true leaf (with stipules) unfolded or first tendril developed
  12. 2 leaves (with stipules) unfolded or 2 tendrils developed
  13. 3 leaves (with stipules) unfolded or 3 tendrils developed
- Stages continuous till .....
19. 9 or more leaves unfolded (with stipules) unfolded or 9 or more tendrils developed

### Principal growth stage 3: Stem elongation (main shoot)

30. Beginning of stem elongation
  31. 1 visibly extended internode <sup>1</sup>
  32. 2 visibly extended internodes <sup>1</sup>
  33. 3 visibly extended internodes <sup>1</sup>
- Stages continuous till .....
39. 9 or more visibly extended internodes <sup>1</sup>

<sup>1</sup> The first internode extends from the scale leaf node to the first true leaf node

### Principal growth stage 5: Inflorescence emergence

51. First flower buds visible outside leaves
55. First separated flower buds visible outside leaves but still closed
59. First petals visible, flowers still closed

### Principal growth stage 6: flowering

60. First flowers open (sporadically within the population)
61. Beginning of flowering: 10% of flowers open
62. 20% of flowers open
63. 30% of flowers open
64. 40% of flowers open
65. Full flowering: 50% of flowers open
67. Flowering declining
69. End of flowering

## Principal growth stage 7: development of fruit

- 71. 10% of pods have reached typical length; juice exudes if pressed
- 72. 20% of pods have reached typical length; juice exudes if pressed
- 73. 30% of pods have reached typical length; juice exudes if pressed. Tenderometer value: 80 TE
- 74. 40% of pods have reached typical length; juice exudes if pressed. Tenderometer value: 95TE
- 75. 50% of pods have reached typical length; juice exudes if pressed. Tenderometer value: 105 TE
- 76. 60% of pods have reached typical length; juice exudes if pressed. Tenderometer value: 115 TE
- 77. 70% of pods have reached typical length. Tenderometer value: 130 TE
- 79. Pods have reached typical size (green ripe); peas fully formed

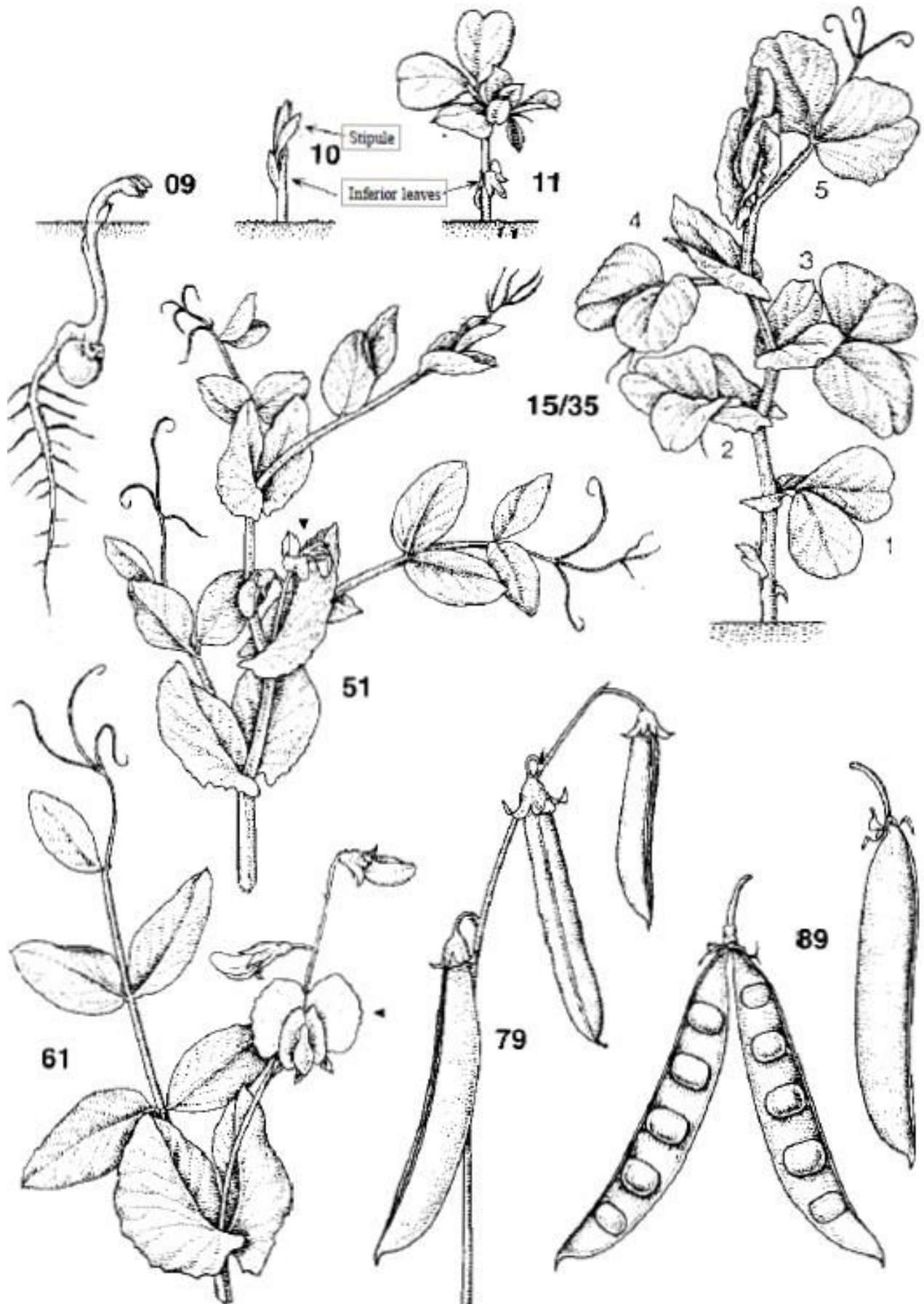
## Principal growth stage 8: ripening

- 81. 10% of pods ripe, seeds final colour, dry and hard
- 82. 20% of pods ripe, seeds final colour, dry and hard
- 83. 30% of pods ripe, seeds final colour, dry and hard
- 84. 40% of pods ripe seeds final colour, dry and hard
- 85. 0% of pods ripe seeds final colour, dry and hard
- 86. 60% of pods ripe seeds final colour, dry and hard
- 87. 70% of pods ripe seeds final colour, dry and hard
- 88. 80% of pods ripe seeds final colour, dry and hard
- 89. Fully ripe: all pods dry and brown. Seeds dry and hard (dry ripe)

## Principal growth stage 9: senescence

- 97 Plants dead and dry
- 99 Harvested product

# Pea growth stages





© Crown copyright 2022

You may re-use this information (excluding logos) free of charge in any format or medium, under the terms of the Open Government Licence v.3. To view this licence visit [www.nationalarchives.gov.uk/doc/open-government-licence/version/3/](http://www.nationalarchives.gov.uk/doc/open-government-licence/version/3/) or email [PSI@nationalarchives.gsi.gov.uk](mailto:PSI@nationalarchives.gsi.gov.uk)

This publication is available at [www.gov.uk/government/publications](http://www.gov.uk/government/publications)

Any enquiries regarding this publication should be sent to us at

[webmaster@apha.gov.uk](mailto:webmaster@apha.gov.uk)

[www.gov.uk/apha](http://www.gov.uk/apha)

The Animal and Plant Health Agency (APHA) is an executive agency of the Department for Environment, Food & Rural Affairs, and also works on behalf of the Scottish Government and Welsh Government.