

SARS-CoV-2 Inactivation Testing: Interim Report

Report identifier HCM/CoV2/075/v1				
Report issue date	sue date 17 February 2021			
Undertaken by High Containment Microbiology, NIS Laboratories, National Infection				
Service, Public Health England				
N.B. This is an interim report and may be updated as further results are obtained				

Product/treatment details	
Product/treatment	Hypochlorous acid 500ppm
Concentration	Product tested undiluted and diluted in 1 part product to 4 parts deionised water
Manufacturer	F & K Holdings
Product code	LOG6
Manufacturer's recommended ratio of sample to product	1 volume sample to 10 volumes product

Sample details		
Sample type tested	Tissue culture fluid containing 5% (v/v) foetal calf	
	serum	
Virus strain tested	SARS-CoV-2 England 2	
Ratio of spiked virus stock to sample matrix	Not applicable; tissue culture fluid used undiluted	

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Experimental conditions		
Ratio of sample to product tested	1 volume sample to 10 volumes product	
Contact time/s	1 minute; 5 minutes	
Temperature of incubation	Ambient temperature	
Brief description of tests performed	Triplicate samples were treated with undiluted or pre-diluted test buffer for indicated contact time/s, or mock-treated in triplicate with an equivalent volume of PBS. Samples were immediately titrated on Vero E6 cells to establish virus titre. This test is quantitative and reports the titre of virus in each treatment condition in TCID50 per ml. Reduction in virus titre following treatment is given as the difference between the mean log ₁₀ TCID50/ml for treated conditions and the PBS control.	

Table of results					
	Mean virus titre in	Titre reduction in	Virus detectable in		
	log ₁₀ TCID50/ml	log ₁₀ TCID50/ml	titration: Yes/No		
	[95% confidence	[95% confidence	(no. of replicates		
	interval]	interval]	positive)		
PBS-treated	5.8 [5.5-6.1]	-	Yes (3/3)		
Undiluted (1 minute)	≤0.7*†	≥5.1 [4.8-5.4]	No		
Undiluted (5 minutes)	≤0.7*†	≥5.1 [4.8-5.4]	No		
1:4 diluted (1 minute)	4.2 [4.0-4.5]	1.6 [1.2-2.0]	Yes (3/3)		
1:4 diluted (5 minutes)	4.4 [4.1-4.6]	1.4 [1.0-1.8]	Yes (3/3)		

^{*}Limit of detection for test was 0.7 log10 TCID50/ml. Mean titres are reported as ≤ when at least one replicate was below the limit of detection.

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^{†95%} confidence interval cannot be calculated

Interpretation

Treatment with undiluted product for 1 minute or 5 minutes reduced SARS-CoV-2 titre by ≥5.1 log₁₀ TCID50/ml, to below the limit of detection for the test.

Treatment with 1:4 diluted product for 1 minute or 5 minutes reduced SARS-CoV-2 titre by 1.6 log₁₀ TCID50/ml and 1.4 log₁₀ TCID50/ml, respectively. Considerable levels of virus were detectable in all sample replicates treated with diluted product.

Here, effectiveness of this product for SARS-CoV-2 inactivation has been assessed in virus suspension tests. Performance in other types of inactivation tests (e.g. in surface tests) may differ.

Demonstrating complete inactivation is dependent on the starting titre of virus used for testing. Complete inactivation may occur if samples contained lower levels of infectious virus than those tested here, but sample treatments that inactivate virus effectively in our testing may fail to inactivate samples containing higher levels of virus than those evaluated in this study.

This test has been performed using tissue culture fluid. The effectiveness of this treatment against SARS-CoV-2 may vary when used to inactivate clinical samples or other types of sample matrix. Any results of inactivation testing using other sample matrices will be released as they become available.

Inactivation reagents should not be assumed to be 100% effective against SARS-CoV-2.

Suitability of products and treatments for inactivation of other pathogens has not been evaluated in this study.

All COVID-19 laboratory testing workflows must be subjected to suitable and sufficient risk assessment, with consideration given to any inactivation step. Risk assessments should be reviewed regularly as new information on the inactivation of SARS-CoV-2 becomes available.

The impact of chosen inactivation method on the sensitivity of subsequent SARS-CoV-2 detection should also be assessed locally.

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Summary of revisions

Version 1: New document

Queries regarding this report or HCM inactivation testing should be directed to HCMgroup@phe.gov.uk

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