Public Health England

SARS-CoV-2 Inactivation Testing: Interim Report

Report identifier	HCM/CoV2/064/v1
Report date	15 December 2020
Undertaken by High Containment Microbiology, NIS Laboratories, National Infection Service, Public Health England N.B. This is an interim report and may be updated as further results are obtained	

Product/treatment details	
Product/treatment	MicroLYSIS-RNA Direct to PCR Lysis Solution
Manufacturer	Clent Life Science
Product code	2MLR
Manufacturer's recommended ratio of sample to product	1 volume sample to 1 volume product

Sample details	
Sample type tested	Tissue culture fluid containing 5% (v/v) foetal calf serum
Virus strain tested	SARS-CoV-2 England 2
Ratio of spiked virus stock to sample matrix	Not applicable; tissue culture fluid used undiluted

Experimental conditions	
Ratio of sample to product tested	1 volume sample to 1 volume product
Contact time/s	5 minutes; 20 minutes
Temperature of incubation	Ambient temperature

Brief description of tests performed	Triplicate samples were treated with test buffer for indicated contact time/s, or mock-treated in triplicate with an equivalent volume of PBS. All samples were then subjected to a purification step to remove cytotoxic buffer components. PBS-treated samples were subjected to the same dilution and purification procedure in parallel. Purified samples were titrated on Vero E6 cells to establish virus titre. This test is quantitative and reports the titre of virus in each treatment condition in TCID50 per ml. Reduction in virus titre following treatment is given as the difference between the mean log_{10} TCID50/ml for treated conditions and the PBS control.

Table of results

	Mean virus titre in log_{10} TCID50/ml [95% confidence interval]	Mean titre reduction in log_{10} TCID50/ml [95% confidence interval]
PBS-treated	6.1 [5.8-6.4]	-
Test buffer-treated (5 minutes)	5.1 [4.8-5.4]	1.0 [0.6-1.4]
Test buffer-treated (20 minutes)	4.2 [3.9-4.4]	1.9 [1.6-2.3]

Interpretation

MicroLYSIS-RNA reduced mean SARS-CoV-2 titre by 1.0 log_{10} and 1.9 log_{10} TCID50/ml after 5 and 20 minutes treatment, respectively.

This product should not be relied upon to completely inactivate infectious samples.

Demonstrating complete inactivation is dependent on the starting titre of virus used for testing. Complete inactivation may occur if samples contained much lower levels of infectious virus than those tested here, and sample treatments that inactivate virus effectively in our testing may fail to inactivate samples containing higher levels of virus than those evaluated in this study.

This test has been performed using concentrated tissue culture fluid. The effectiveness of this treatment against SARS-CoV-2 may vary when used to inactivate clinical samples or other types of sample matrix. Any results of inactivation testing using other sample matrices will be released as they become available.

Inactivation reagents should not be assumed to be 100% effective against SARS-CoV-2.

Suitability of products and treatments for inactivation of other pathogens has not been evaluated in this study.

All COVID-19 laboratory testing workflows must be subjected to suitable and sufficient risk assessment, with consideration given to any inactivation step. Risk assessments should be reviewed regularly as new information on the inactivation of SARS-CoV-2 becomes available.

The impact of chosen inactivation method on the sensitivity of subsequent SARS-CoV-2 detection should also be assessed locally.

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Summary of revisions
Version 1: New document

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