DEPARTMENT FOR ENVIRONMENT, FOOD AND RURAL AFFAIRS
SCOTTISH GOVERNMENT
WELSH GOVERNMENT
DEPARTMENT OF AGRICULTURE, ENVIRONMENT AND RURAL AFFAIRS, NORTHERN IRELAND

EXPORT OF FROZEN OVINE EMBRYOS TO AUSTRALIA - PART A **No: .....**

HEALTH CERTIFICATE

EXPORTING COUNTRY: UNITED KINGDOM (GREAT BRITAIN)

FOR COMPLETION BY: TEAM VETRERINARIAN/OFFICIAL VETERINARIAN

I. Information concerning:

a) The donor animals, semen and embryos (attached Schedule (8552SUP) refers).
b) Total number of ampoules/straws certified:

II. Origin of the embryos

a) Name and address of exporter:

b) Address of premises of herd/flock of origin:

c) Name and Address of approved Embryo Collection Unit or premises at which embryos were collected:

III. Destination of the embryos

a) Name and address of consignee:

b) Address of destination:#

c) Means of transportation:

d) Import permit number(s):

IV. Information concerning the embryo collection team

a) Registration number of Embryo Collection Team:

b) Name and address of approved Embryo Collection Team Veterinarian

c) IETS No/Code:

V. Origin of the semen

a) Name and address of the Semen Collection Centre in which the semen of the donor sire originated, if different from that at Section II c):

Health Information

VI. I, the undersigned, certify that:

a) Foot and mouth disease (FMD). Immediately prior to the pre-collection period each donor was living in the United Kindgom or a zone of the UK recognised by the World Organisation for Animal Health (OIE) as being free from foot and mouth disease.

The embryos were not collected in the United Kingdom: between 1 January 2001 and 15 January 2002 and between 1 July 2007 and 18 February 2008 (inclusive of these dates).

b) The United Kingdom meets the Code Article definitions for country freedom from sheep and goat pox (capripox virus).

c) The embryos came from donors (both male and female) which are 5 years of age or older prior to export of the embryos, and which had lived only in the United Kingdom or a zone of the UK where:
i) scrapie has been compulsorily notifiable during the previous 6 years;
ii) an effective and continuous national surveillance system is practiced;
iii) brains from clinically suspect animals which are slaughtered or die are examined in a laboratory in accordance with the diagnostic techniques set out in the World Organisation for Animal Health (OIE) Manual of Standards for Diagnostic Tests and Vaccines;
iv) the feeding of ruminant derived meat-and-bone meal to sheep and goats is banned;
v) scrapie-affected sheep and goats are slaughtered and their carcasses disposed of in a manner that would reliably preclude the spread of scrapie infective agent (such as complete incineration);
vi) procedures are followed which allow tracing of each scrapie affected animal back to its flock of birth.

d) The embryos came from donors (both male and female) which are 5 years of age or older prior to export of the embryos, and which are of a homozygous PrP genotype known to be susceptible to scrapie, in relation to the particular breed of sheep (approved by Department of Agriculture), as verified in the attached certificate/s from a laboratory/laboratories officially approved by the Veterinary Administration to do PrP genotype testing.

[Note: Breeds and genotypes permitted without consultation with Department of Agriculture: Suffolk – QQ at Codon 171, and Cheviot, Texel, Charollais – VRQ/VRQ (at Codons 136/154/171].

Requests for the importation of embryos from other breeds will be considered after receiving details of breed specific PrP genotype and scrapie susceptibility through the veterinary administration of the exporting country. Suffolk semen donors may be of genotype QH but must undergo the same autopsy procedure as required of female donors]

e) The embryos came from donors (both male and female) which are 5 years of age or older prior to export of the embryos, and which originated from flocks:
i)in which no case of scrapie has been confirmed or suspected during the 5 years immediately prior to collection;
ii) in which all animals are identified and can be traced back to their flock of birth
iii) for which records of parentage, and movements of animals in and out of the flock, are maintained for a minimum period of 5 years;
iv) into which, during the previous 5 years, introductions were only permitted from flocks with equivalent scrapie status;
v) in which no animals have commingled with flocks of lower scrapie status during the previous 5 years;
OR:
vi) for which confirmed information is available which would provide equivalent security to the above.*
**[Note: Applications for this option must be made to Department of Agriculture through the Veterinary Administration of the exporting country.] [The veterinary certificate must indicate the option that applies.]*

f) Each donor was isolated from all ruminants, except other donors of equivalent health status, during the pre-collection and the collection periods.
Prior to entry into quarantine each donor was individually identified by microchip

implanted midline between the shoulder blades or behind the ear.

The microchip or electronic implant is any radio frequency identification device approved for use by the exporting country Veterinary Administration which is tamper resistant and readable by equipment available to the Veterinary Administration. The donor must be identified by scanning the microchip at the each process which must be certified.

g) Donors were:
i) not vaccinated against any diseases, except tetanus using a killed vaccine, during the pre-collection period nor during collection;
ii) clinically inspected at least each week during the pre-collection period and on each day blood samples were collected and, at each inspection, were found to be free from signs of contagious and infectious diseases *(by the team veterinarian, Official Veterinarian or a registered veterinarian appointed by the team veterinarian and acting under written instruction).*

h) All animal health testing, to meet these conditions, was performed at laboratories, and using tests, approved by the Veterinary Administration of the exporting country.

j) Bluetongue (BT). Prior to the export of this consignment each female embryo donor must be certified as follows for Bluetongue:

i) A competitive enzyme linked immunosorbent assay (cELISA) for antibody to the bluetongue virus (BTV) group on a blood sample, with negative results, between 28 and 60 days after the collection of embryos for this consignment;

OR:
ii) A bluetongue virus isolation test or an approved real time reverse- transcriptase polymerase chain reaction (RT- PCR) test* on a blood sample taken on the day of collection of embryos for this consignment, with negative results;

** Real time reverse transcriptase- polymerase chain reaction (RT-PCR) tests must be approved by the competent authority and be able to detect all known 24 BTV serotypes. These tests must use primer sequences directed against highly conserved segments of the BTV genome which code for BTV serogroup (not serotype). An example of an appropriate test is the TaqMan real time RT-PCR test according to the method of Shaw et al. (2007), which uses two primers directed against segment 1 of BTV ribonucleic acid (RNA).*

[All tests for BTV should be validated according to the current OIE Manual of diagnostic tests and vaccines for Terrestrial Animals, calibrated to a diagnostic sensitivity of at least 98.0% and carried out in a laboratory approved by the competent authority of the exporting country.] [Serological testing for BTV antibodies with agar gel immunodiffusion (AGID) tests should not be used.]

AND (iii) Donors vaccinated against BTV: Yes / No

If Yes, vaccines against BTV administered to embryo donors must be inactivated, and · approved by the competent authority in the exporting country, and administered more than 60 days before embryo collection for this consignment.

Name of BTV used:

Date of administration of BTV vaccine to embryo donor
[The veterinary certificate must indicate the option that applies. The attached table must include dates of sampling for test, type of tests used, test results.]

k) Brucella melitensis infection. Each donor:
i) has lived only in a country or zones which meets World Organisation for Animal Health (OIE) Code Article requirements for country freedom;
OR:
ii) immediately prior to the pre-collection period, was part of a flock officially free from B. melitensis infection according to World Organisation for Animal Health (OIE) Code Article requirements and gave a negative result to a complement fixation test (CFT) and a Rose Bengal plate agglutination test for B. melitensis infection on the same blood sample taken during the pre-collection period or at autopsy.

[The veterinary certificate must indicate which option applies. The attached table must include dates of sampling for test, type of test used, test results.]

l) Contagious agalactia (CA). Each donor has lived on premises in which contagious agalactia has not been diagnosed during the 6 months immediately prior to the pre-collection period.

m) Maedi-visna (MV) Each donor:
i) immediately prior to embryo collection was part of an accredited MV free flock recognised by the Veterinary Administration;

OR:
ii) immediately prior to embryo collection was part of a flock in which MV had not been diagnosed during the previous 3 years and during this 3 year period no commingling with goats occurred and no animals were introduced from flocks with a lesser disease status;

(Commingling is defined as animals grouped together having physical contact. This does not include incidental contact between animals off the flock's premises, such as occurs at shows and sales)

AND (iii) either gave a negative result to an approved ELISA for MV antibodies on two blood samples collected 30 days apart during the pre-collection period, at the time of collection or at autopsy or was sourced from a flock which tested negative within the 6 months immediately prior to export.

(A flock which "tested negative" is defined as a closed flock in which a sample, sufficient to provide 95% probability of detecting evidence of MVV at 10% prevalence, tested negative to an approved AGID or ELISA.) [The veterinary certificate must indicate the option that applies. The attached table must include dates of sampling for test, type of tests used, test results.]

n) Enzootic abortion of ewes. Each donor:
i) has lived on premises in which enzootic abortion of ewes (EAE) had not been diagnosed during the 2 years immediately prior to the pre-collection period;

AND ii) gave a negative result to a CFT test for EAE during the pre-collection period. [The attached table must include dates of sampling for test, type of tests used, test results.]

Stamp **Signed .....RCVS**

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Name in block letters
Team Veterinarian
Telephone number:
Email address:

Date ..... **Address .....**

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Stamp **Signed .....RCVS**

.....
Name and title in block letters

Official Veterinarian
Telephone number:
Email address:

Date ..... **Address .....**

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*** Delete as appropriate**

CONTINUED (Please note that this export health certificate consists of three parts, Part A, Part B and Schedule: 8552EHC, 8552CON and 8552SUP which must be complete in full to accompany the export).