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|  **Food and Environmental Proficiency Testing Unit** Store LENTICULE disc samples @ -20°C± 5ºC**Laboratory Identification no. Lab No.** Dispatch date: **XX XXXX 20XX** Final date for return of results: **XX XXXX 20XX**  |
| **Contact details:**The Organisers - FEPTUPublic Health England61 Colindale Avenue, London, NW9 5EQ, UK. Fax: +44 (0) 20 8200 8264 Tel: +44 (0) 20 8327 7119 e-mail: foodeqa@phe.gov.uk | **RESULTS TO BE RETURNED VIA ONLINE SURVEY AS PER EMAIL SENT TO YOU** |
|  **0006**  |
|  |
| **Shiga toxin producing *Escherichia coli* (STEC) Request/Report Form** |
| **Distribution No.: STXXX** | **Sample numbers**: **STX00X & STX00X**  |
| Download the sample Instruction sheet | [www.gov.uk/government/publications/shiga-toxin-escherichia-coli-scheme-sample-instruction-sheet](http://www.gov.uk/government/publications/shiga-toxin-escherichia-coli-scheme-sample-instruction-sheet) |
| Download the safety data sheet: | [www.gov.uk/government/publications/safety-data-sheet-for-lenticules](http://www.gov.uk/government/publications/safety-data-sheet-for-lenticules) |
| ***If you cannot examine any of these samples return your results as ‘Not examined’*** |
| **Request:** | 1. Examine samples for STEC genes
 |
|  |  |

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**SAMPLE: STX00X**

**STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-**

|  | **Target Genes** | **Positive (circle)** |  **CT value** |   |  | **Not examined** | **Serogroup****(e.g. O157)** | **Positive (circle)** | **CT value** | **Serotype****(e.g. H7)** | **Positive (circle)** | **CT value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  **STEC virulence gene detection**  | *stx* *(stx* 1/2 combined) | Yes / No / NE |  | **STEC identification gene detection** | 🞏 | O157 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 1 | Yes / No / NE |  | O26 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 2 | Yes / No / NE |  | O103 | Yes / No / NE |  |  | Yes / No / NE |  |
| *eae*  | Yes / No / NE |  | O104 | Yes / No / NE |  |  | Yes / No / NE |  |
| Other …….. | Yes / No / NE |  | O111 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | O145 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | Other………. | Yes / No / NE |  |  | Yes / No / NE |  |

 NE = Not Examined

**Interpretation of results:-**

|  |  |
| --- | --- |
| **Select interpretation of your results** | **Explanation** |
| 🞏 | Not examined / Not applicable  |  |
| 🞏 | STEC not detected in the test portion of x g or xml |  |
| 🞏 | Presumptive detection of STEC in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml |  |
| 🞏 | Other: |  |

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| **Laboratory Identification no. Lab No.**   |  |

**SAMPLE: STX00X**

**STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-**

|  | **Target Genes** | **Positive (circle)** |  **CT value** |   |  | **Not examined** | **Serogroup****(e.g. O157)** | **Positive (circle)** | **CT value** | **Serotype****(e.g. H7)** | **Positive (circle)** | **CT value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  **STEC virulence gene detection**  | *stx* *(stx* 1/2 combined) | Yes / No / NE |  | **STEC identification gene detection** | 🞏 | O157 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 1 | Yes / No / NE |  | O26 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 2 | Yes / No / NE |  | O103 | Yes / No / NE |  |  | Yes / No / NE |  |
| *eae*  | Yes / No / NE |  | O104 | Yes / No / NE |  |  | Yes / No / NE |  |
| Other …….. | Yes / No / NE |  | O111 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | O145 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | Other………. | Yes / No / NE |  |  | Yes / No / NE |  |

NE = Not Examined

**Interpretation of results:-**

|  |  |
| --- | --- |
| **Select interpretation of your results** | **Explanation** |
| 🞏 | Not examined / Not applicable  |  |
| 🞏 | STEC not detected in the test portion of x g or xml |  |
| 🞏 | Presumptive detection of STEC in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml |  |
| 🞏 | Other: |  |

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| **Laboratory Identification no. Lab No.**   |  |

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| **Any further comments:-** |

**Authorised by: Date reported:**

**Laboratory Identification no. Lab No**

**METHOD QUESTIONNAIRE – not all findings will be presented in the scheme report (include as much detail as possible or references):**

|  |  |  |
| --- | --- | --- |
| **1. Do you exactly follow ISO/TS 13136:2012** *Microbiology of food and animal feed -- Real-time polymerase chain reaction (PCR)-based method for the detection of food-borne pathogens - Horizontal method for the detection of Shiga toxin-producing* Escherichia coli *(STEC) and the determination of O157, O111, O26, O103 and O145 serogroups* | **YES** | **NO** |

|  |  |
| --- | --- |
| **2. If the answer to question 1 is NO please state deviations to ISO/TS 13136:2012**  |  |

|  |  |
| --- | --- |
| **3. For routine samples, briefly describe your enrichment process**e.g. 25 g into 225 ml mTSB+N/BPW 41.5 °C for 24hr  |  |

|  |  |
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| **4. Provide the details of your DNA extraction kit** Include company name, kit name, protocol(s) if applicable:e.g., Promega, Maxwell® 16 Cell DNA purification kit |  |

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| **5. If your DNA extraction process requires a platform please state platform used**   |  |

**Laboratory Identification no. (Check) Lab No**

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| **6. State if you use conventional** **RT PCR or real-time RT PCR** |  |

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| **7. If using a commercial kit for primer / probe / amplification mix**Please provide company name, kit name: e.g., Bio-Rad, IQ-Check STEC SerO; Bio-Rad IQ-Check® STEC VirX |  |

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| --- | --- |
| **8. What volume of extracted DNA is used?** |  |

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
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| **9. Please complete the Cycle information tables** | **Pre-incubation:**

|  |  |  |  |
| --- | --- | --- | --- |
| **Time (hh:mm:ss)** |  | **Temperature (°C)** |  |

**Cycle information:**

|  |  |  |  |
| --- | --- | --- | --- |
| **Parameter** | **Denaturisation** | **Cycling** | **Cooling** |
| **Cycles** | **x1** | **x**…….. | **x1** |
| **Temperature (°C)** |  | **Step 1:** | **Step 2:** | **Step 3:** | **Step 4:** |  |
|  |  |  |  |
| **Hold (hh:mm:ss)** |  |  |  |  |  |  |

**Other:**  |

**Laboratory Identification no. Lab No**

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| --- | --- |
| **10. Amplification platform used**e.g. Bio-Rad CFX96 Touch™ Deep Well RT-PCR Detection Systems |  |
| **11. Please use this space for any additional comments you feel are relevant or general comments about the scheme**  |  |