

# Xpert MTB/RIF

Automated molecular detection of TB and MDR  
screening in peripheral laboratories

Introduction to the technology

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CSO FIND



GeneXpert

Xpert  
 MTB/RIF



5      20      80      Samples per shift      500-1000

## Multi-disease technology platform

**Table 34: Existing test cartridges on the GeneXpert platform**

*Staphylococcus aureus* colonization

Vancomycin resistance

*Clostridium difficile*

MRSA from tissue or blood

Group B *Streptococcus*

Enteroviral meningitis

Coagulation disorders

Anthrax

*Bordetella pertussis*

*Bordetella parapertussis*

HSV Type 1

HSV Type 2

RSV Type A

RSV Type B

Norovirus GI

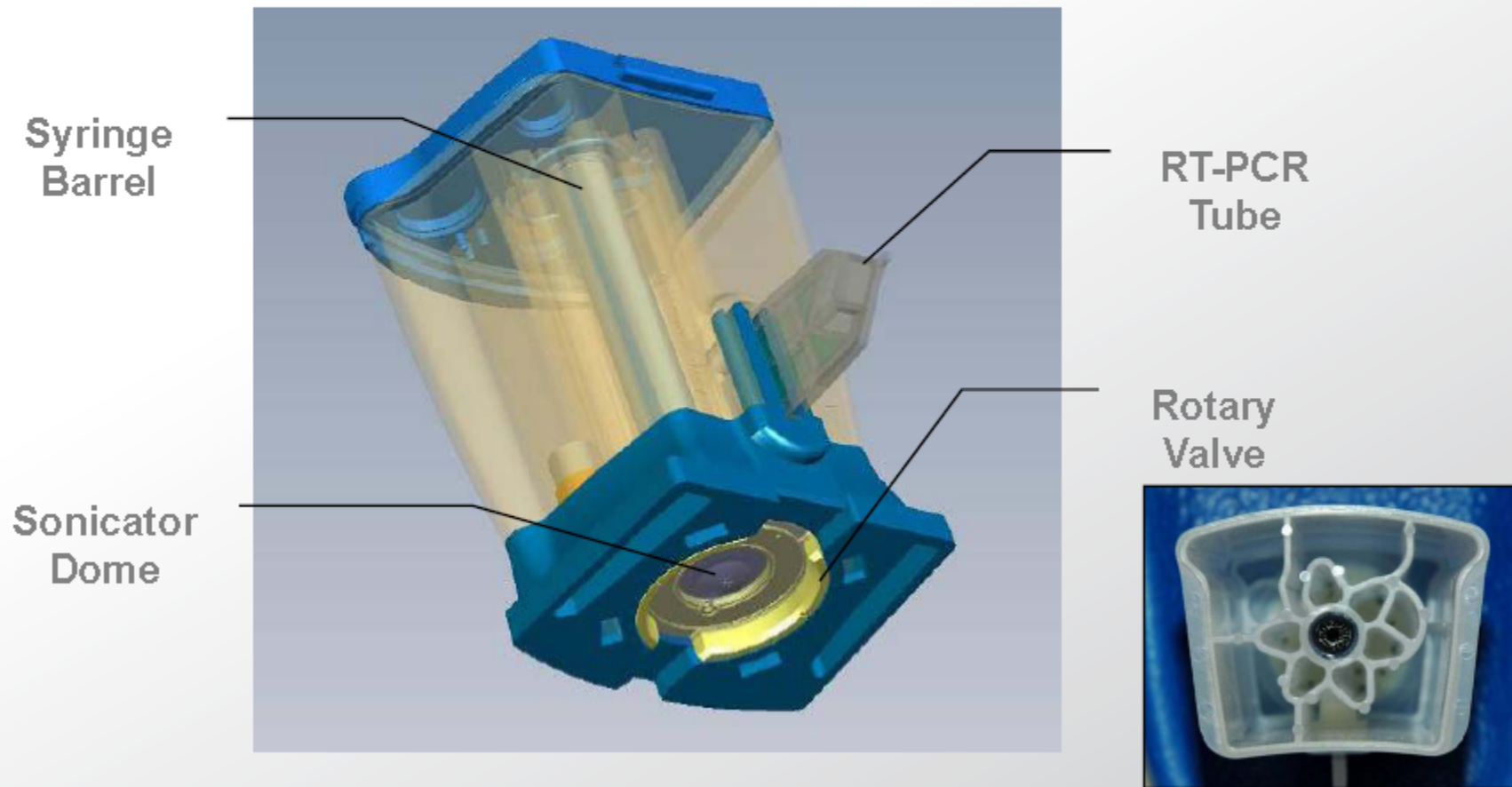
Norovirus GII

Flu A

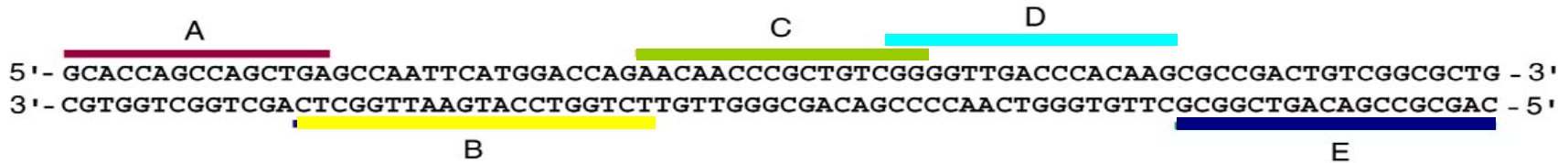
Flu B

Leukemia (BCR-ABL)

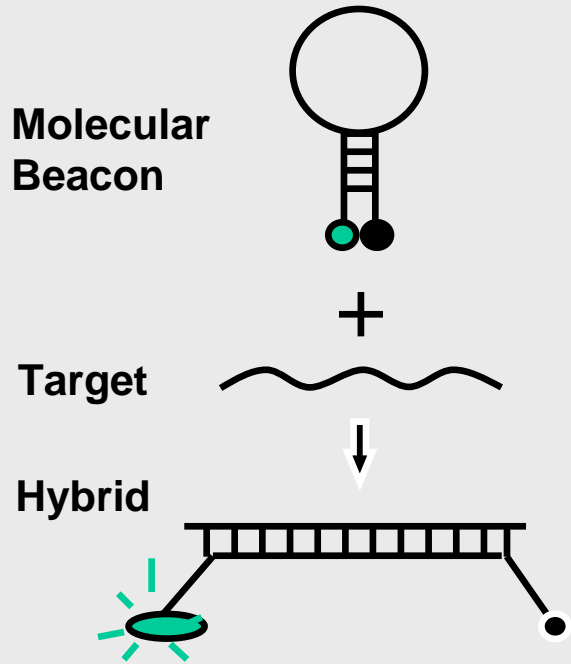
# Cartridge Design and Operating Principle



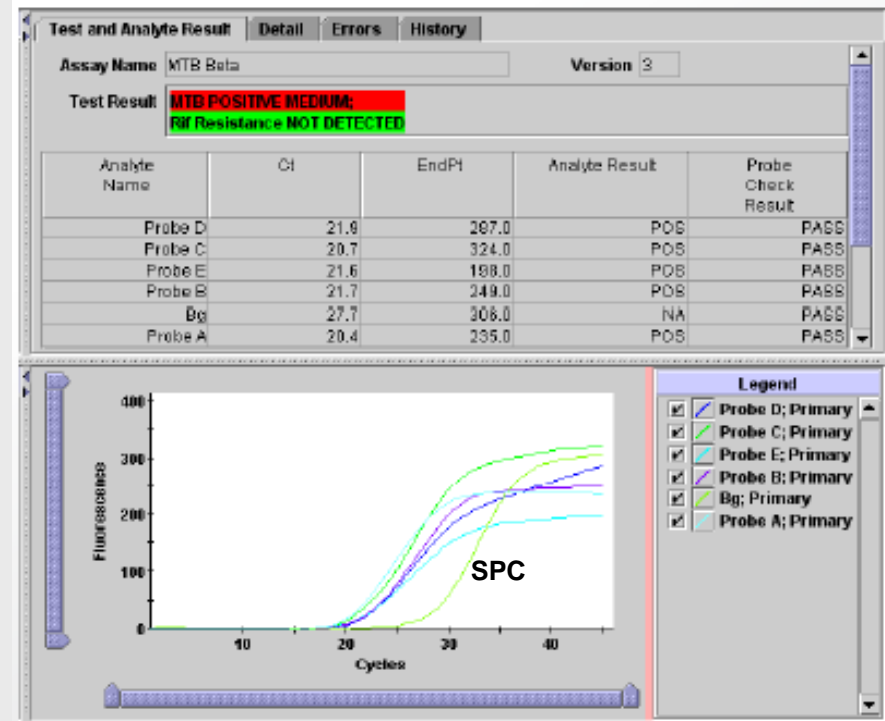
# Xpert MTB/Rif molecular beacon assay



The PCR target is the 81 bp region of the *rpoB* gene: 5 probes bind to wildtype, but not mutant target

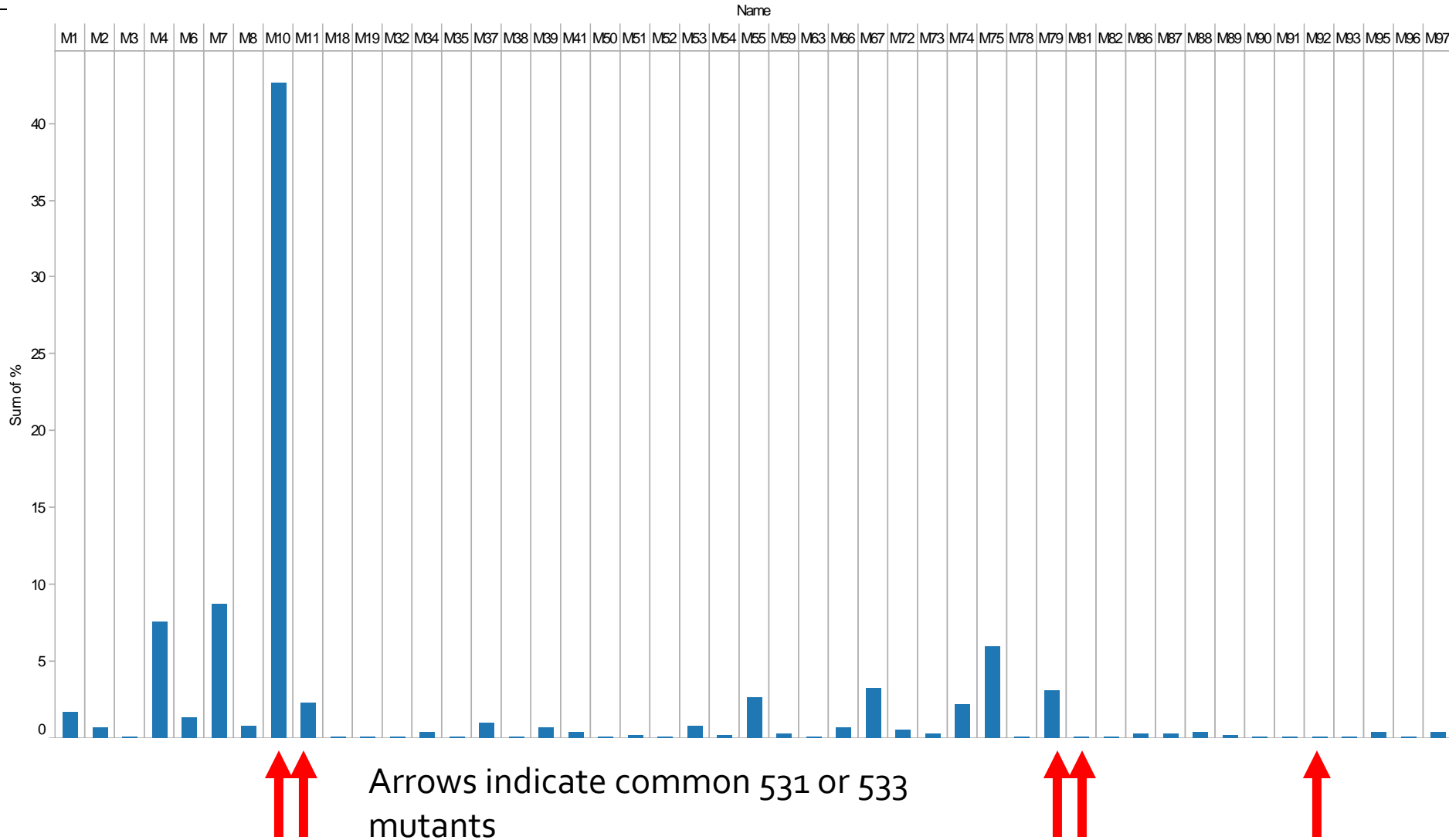


Each probe is labeled with a different fluorescent dye, permitting simultaneous detection



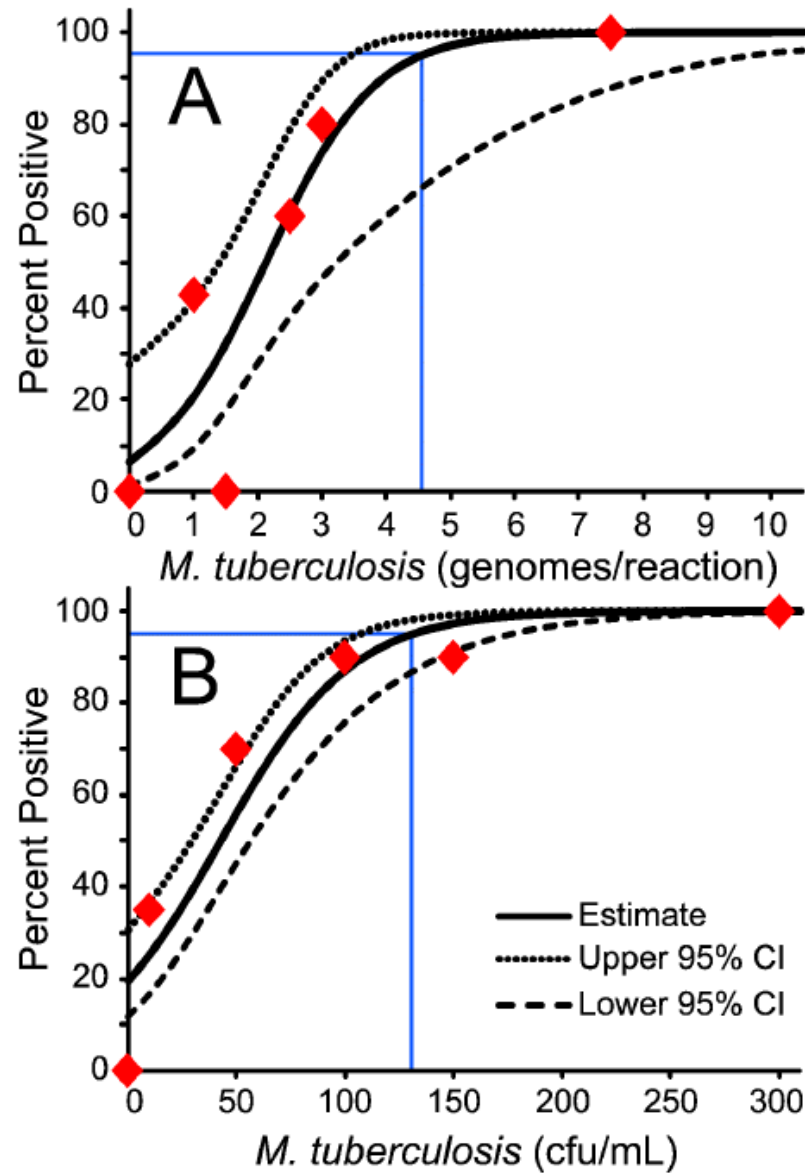
Example of Rif-Sensitive Profile – 5 probes & SPC show fluorescence

# Mutation Frequency



Frequency of mutation based on review of 4000+ strains

# Limits of detection



# Linearity of dynamic range

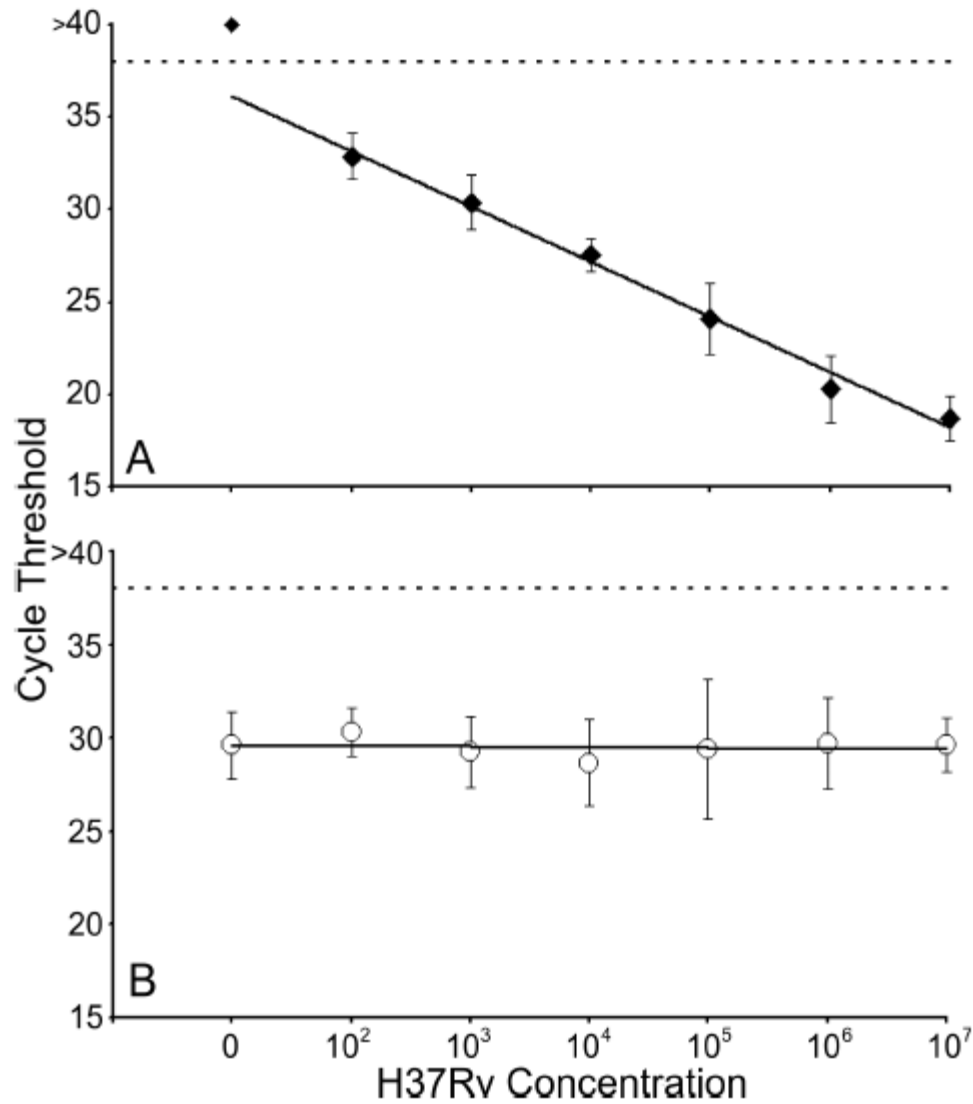




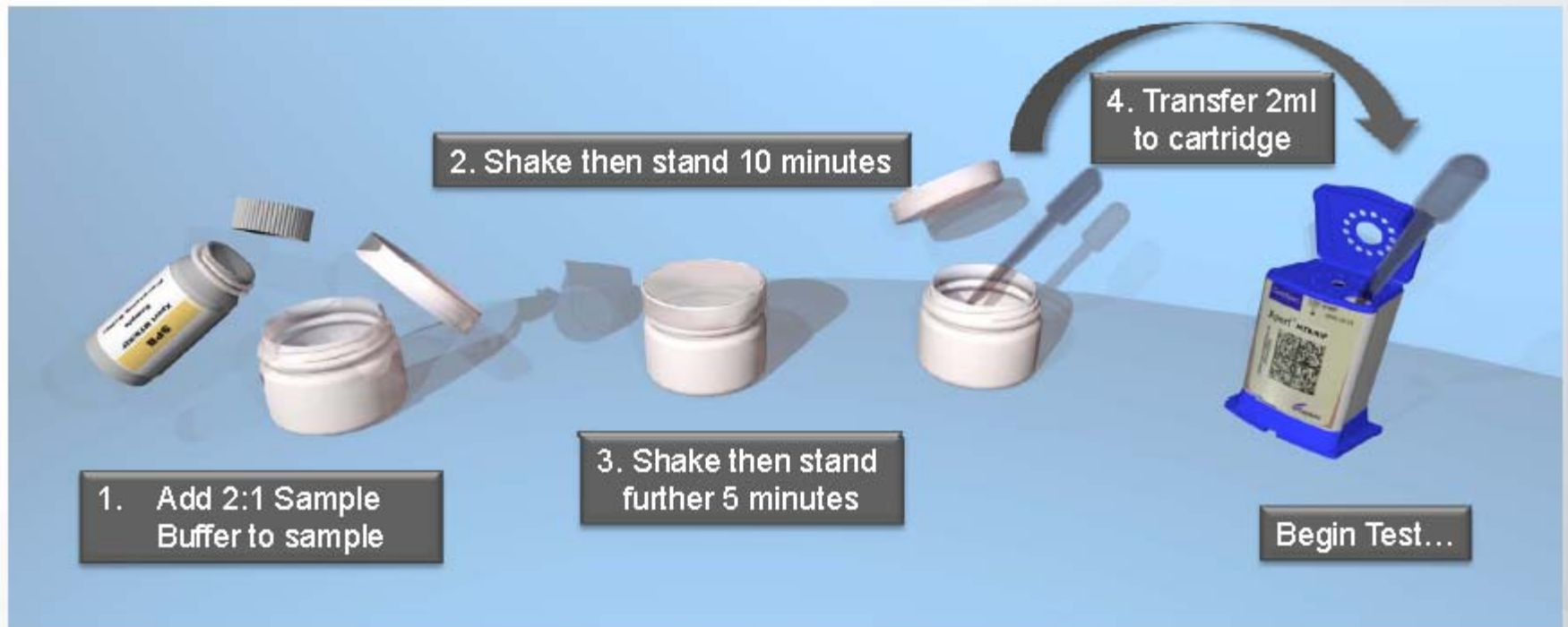
TABLE 1: Specificity Panel

BACTERIA <sup>c</sup>			
<i>Acinetobacter baumannii</i>	BEI NR-10146	<i>Propionibacterium acnes</i>	Clinical Strain
<i>Acinetobacter calcoaceticus</i>	Clinical Strain	<i>Proteus mirabilis</i>	Clinical Strain
<i>Actinomyces israelii</i>	ATCC 12102	<i>Proteus vulgaris</i>	BEI DD-460
<i>Actinomyces mayeri</i>	Clinical Strain	<i>Providencia alcalifaciens</i>	PI #368
<i>Bacillus cereus</i>	BEI NR-4198	<i>Pseudomonas aeruginosa</i>	ATCC 27853
<i>Bacillus subtilis</i>	Clinical Strain	<i>Rhodococcus equi</i>	ATCC 14187
<i>Bordetella parapertussis</i>	ATCC 151311D	<i>Salmonella enterica</i>	BEI NR-615
<i>Bordetella pertussis</i>	Cepheid 500-0780	<i>Salmonella typhi</i>	Clinical Strain
<i>Campylobacter jejuni</i>	BEI NR-3057	<i>Serratia marcescens</i>	Clinical Strain
<i>Chlamydia pneumoniae</i>	Cepheid VR 1360 CM-1	<i>Shigella boydii</i>	Clinical Strain
<i>Citrobacter freundii</i>	Clinical Strain	<i>Shigella flexneri</i>	Clinical Strain
<i>Corynebacterium diphtheriae</i>	PI #581	<i>Staphylococcus aureus</i>	ATCC 25953
<i>Corynebacterium pseudodiphtheriticum</i>	ATCC 10700	<i>Staphylococcus capitis</i>	Clinical Strain
<i>Corynebacterium xerosis</i>	Clinical Strain	<i>Staphylococcus epidermidis</i>	ATCC 12228
<i>Enterobacter aerogenes</i>	Clinical Strain	<i>Staphylococcus haemolyticus</i>	Clinical Strain
<i>Enterobacter cloacae</i>	Clinical Strain	<i>Staphylococcus hominis</i>	Clinical Strain
<i>Enterococcus avium</i>	Clinical Strain	<i>Staphylococcus lugdunensis</i>	Clinical Strain
<i>Enterococcus faecalis</i>	Clinical Strain	<i>Stenotrophomonas maltophilia</i>	Clinical Strain
<i>Enterococcus faecium</i>	Clinical Strain	<i>Streptococcus equi</i>	Clinical Strain
<i>Escherichia coli</i>	Clinical Strain	<i>Streptococcus pyogenes</i>	ATCC 19615
<i>Escherichia coli O157H7</i>	ATCC 35150	<i>Streptococcus agalactiae</i>	ATCC 12386
<i>Fusobacterium nucleatum</i>	ATCC 25586D	<i>Streptococcus constellatus</i>	Clinical Strain
<i>Haemophilus influenzae</i>	ATCC 49247	<i>Streptococcus mitis</i>	Clinical Strain
<i>Haemophilus parahaemolyticus</i>	ATCC 10014	<i>Streptococcus mutans</i>	Clinical Strain
<i>Haemophilus parainfluenzae</i>	Clinical Strain	<i>Streptococcus pneumoniae</i>	Clinical Strain
<i>Klebsiella oxytoca</i>	Clinical Strain	<i>Streptococcus uberis</i>	Clinical Strain
<i>Klebsiella pneumoniae</i>	Clinical Strain	<i>Veillonella parvula</i>	ATCC 10790D-5
<i>Legionella pneumophila</i>	ATCC 33152D	<i>Stenotrophomonas maltophilia</i>	Clinical Strain
<i>Leuconostoc mesenteroides</i>	Clinical Strain	<i>Yersinia pestis</i>	BEI NR-2644
<i>Listeria grayi</i>	ATCC 25401		FUNGUS
<i>Listeria monocytogenes</i>	BEI NR-4211	<i>Candida albicans</i>	ATCC 14053D
<i>Moraxella catarrhalis</i>	ATCC 8176	<i>Cryptococcus neoformans</i>	Casadevall Lab
<i>Morganella morganii</i>	Clinical Strain	<i>Histoplasma capsulatum</i>	Nosanchuk Lab
<i>Mycoplasma pneumoniae</i>	ATCC 15531D	<i>Kingella kingae</i>	ATCC 23332D
<i>Neisseria gonorrhoeae</i>	ATCC 49226		VIRUS
<i>Neisseria lactamica</i>	ATCC 23971	<i>Adenovirus<sup>b</sup></i>	ZM NATADV1-ST
<i>Neisseria meningitidis</i>	Clinical Strain	<i>Herpes simplex virus 1</i>	Cepheid 500-1268
<i>Neisseria mucosa</i>	ATCC 69696	<i>Herpes simplex virus 2</i>	Cepheid 500-1269
<i>Nocardia asteroides</i>	ATCC 19247	<i>Influenzavirus A<sup>a</sup></i>	Cepheid 500-0763
<i>Nocardia cyriacgeorgica</i>	ATCC BAA-1516	<i>Influenzavirus B<sup>a</sup></i>	500-0764
<i>Nocardia farcinica</i>	ATCC 3318	<i>Parainfluenza 2<sup>b</sup></i>	ZM NATPARA2-ST
<i>Pasteurella multocida</i>	ATCC 3160	<i>Parainfluenza 3<sup>b</sup></i>	ZM NATPARA3-ST
<i>Peptostreptococcus anaerobius</i>	ATCC 49031D	<i>Respiratory Syncytial Virus A<sup>a</sup></i>	Cepheid 500-0765
<i>Porphyromonas gingivalis</i>	ATCC 33277D-5	<i>Respiratory Syncytial Virus B<sup>a</sup></i>	Cepheid 500-0765
<i>Prevotella melanogenica</i>	ATCC 25845D-5	<i>Rhinovirus 6<sup>a</sup></i>	Cepheid
		<i>Rhinovirus 16<sup>a</sup></i>	Cepheid

- Testing GX with  $10^6$  copies of genomic DNA from common oral microorganisms and those causing tuberculosis like symptoms.

- DNA is spiked into PCR reagent resuspension buffer, simulating efficient capture, lysis and elution.

## Simple Sample Processing – Direct Sputum

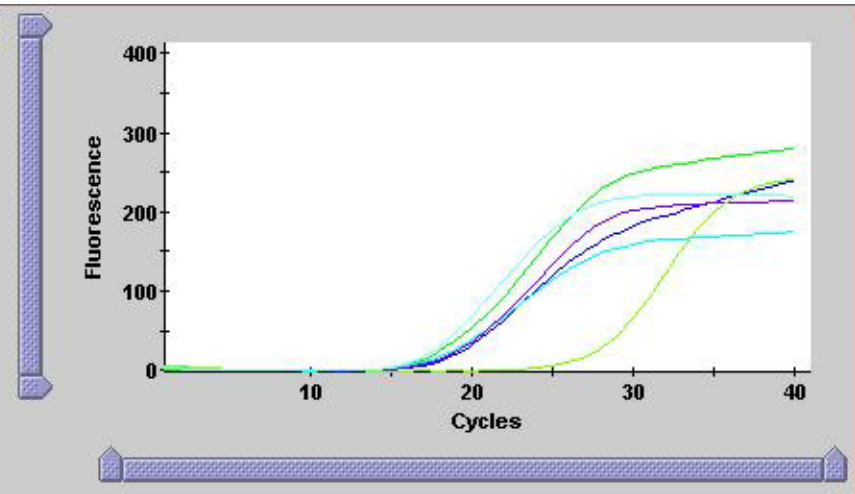


# Rifampin Susceptible Sample

**Module Name** B1  
**Sample ID** uganda sputum.6  
**Assay** MTB Beta  
**Assay Version** 2  
**Assay Type** Research Use Only  
**Reagent Lot ID** 00502  
**Cartridge S/N** 0  
**Expiration Date** <None>  
**Test Type** Specimen

**Notes**  
**Start Time** 2008/4/16 15:50:33  
**End Time** 2008/4/16 17:15:31  
**Status** Done  
**Error Status** OK  
**User** support  
**S/W Version** 2.1  
**Instrument/Module S/N** 702266/600013

- Views**
- Result View
  - MTB Beta Assay
  - Temperature-Sampl
  - Optic-All Options
  - Optic-Primary**
  - Pressure
  - Optic-Primary-Thres
  - 2nd Derivative



- Legend**
- Probe D; Primary
  - Probe C; Primary
  - Probe E; Primary
  - Probe B; Primary
  - Bg; Primary
  - Probe A; Primary

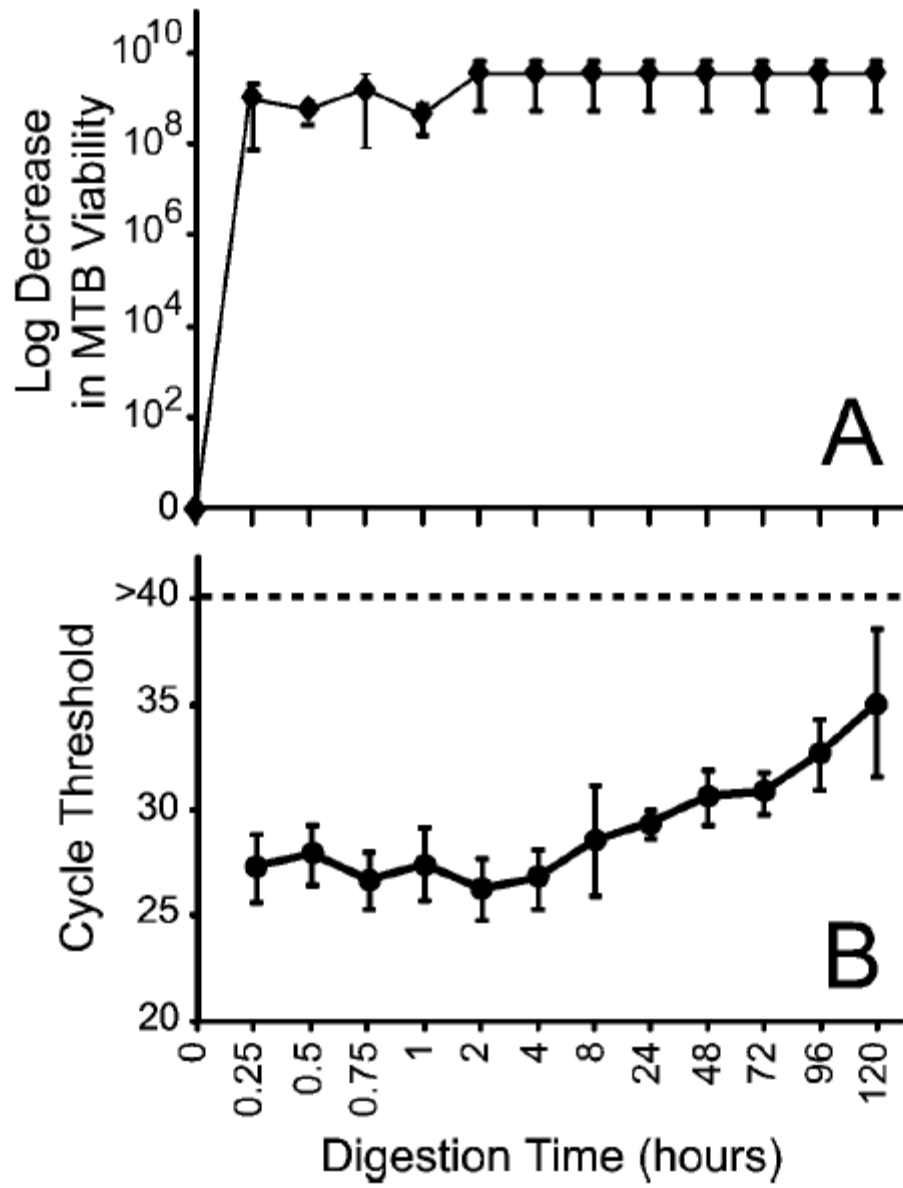
- Views**
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  - Optic-Primary
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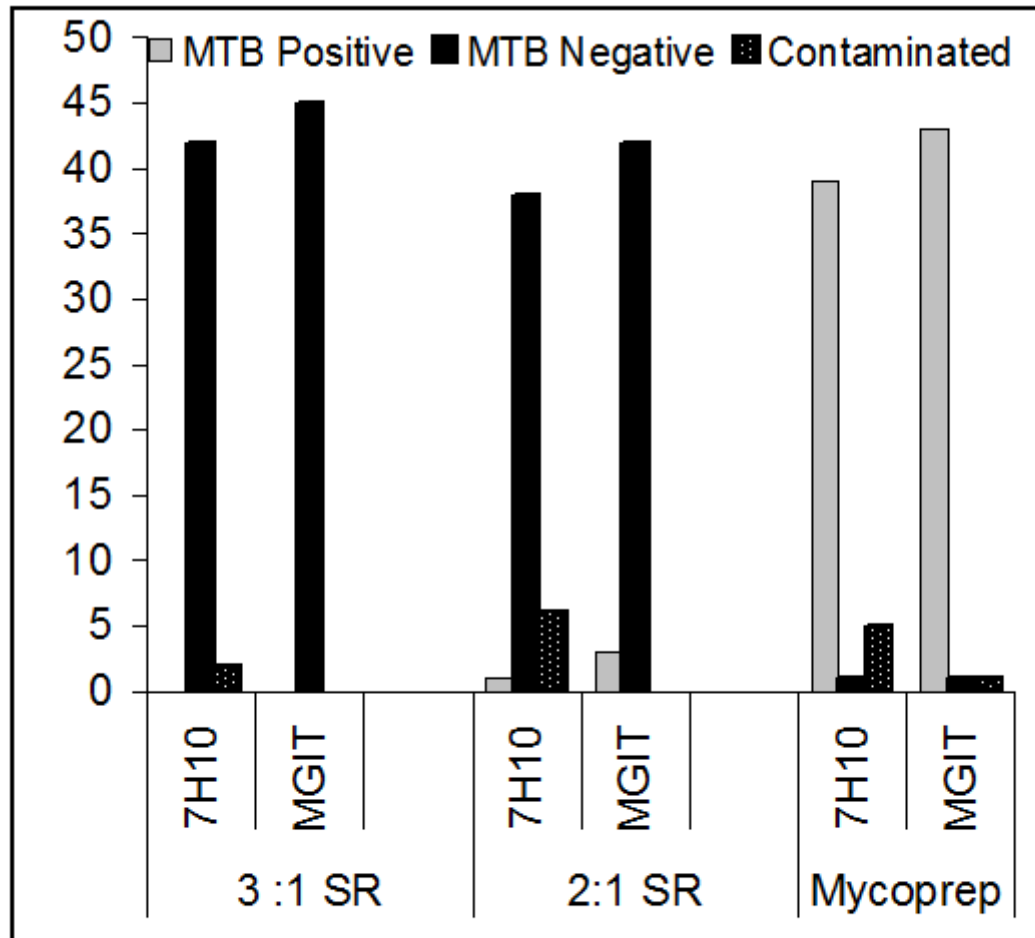
**Test and Analyte Result** | Detail | Errors | History | Messages  
**Assay Name** MTB Beta | **Version** 2  
**Test Result** **MTB POSITIVE MEDIUM;**  
**Rif Resistance NOT DETECTED**

Analyte Name	Ct	EndPt	Analyte Result	Probe Check Result
Probe D	19.1	240.0	POS	PASS
Probe C	17.7	279.0	POS	PASS
Probe E	18.4	174.0	POS	PASS
Probe B	18.8	214.0	POS	PASS
Bg	27.4	240.0	NA	PASS
Probe A	17.3	220.0	POS	PASS

# Inactivation procedure

	Starting cfu/mL	Diluent	Average cfu/ plate for each replicate	Average cfu/plate	Average cfu/mL	Average log Reduction	Percent Reduction
Study 1	$3.5 \times 10^7$ BCG	7H9 media			<10	$>3.5 \times 10^6$	>99.9
Study 2	$3.5 \times 10^7$ BCG	sputum			1.5	$2.3 \times 10^7$	>99.9
Study 3	$3.3 \times 10^7$ H37Rv	sputum	12, 13, 21	15.3	153	$2.15 \times 10^5$	99.9
Study 4	$3.63 \times 10^8$ H37Rv	sputum	4, 6, 2	4	40	$9.1 \times 10^6$	>99.9
Study 5	$4.0 \times 10^8$ H37Rv	sputum	2, 2, 3	2.33	23.3	$1.7 \times 10^7$	>99.9
Average log kill						$1.06 \times 10^7$	



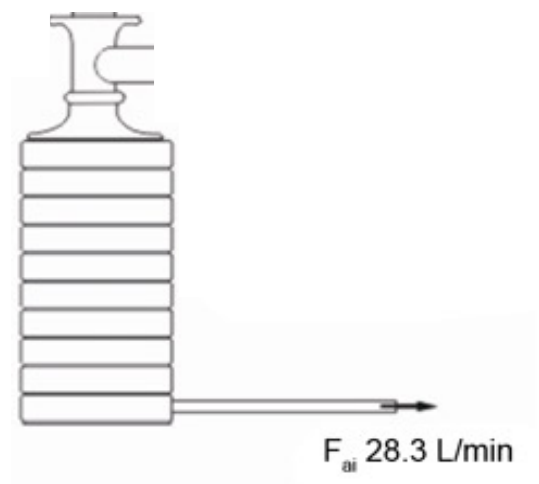


**Figure 12: Inactivation effectiveness of SR on smear-positive sputum**

Biosampler  
Liquid media



Anderson Impactor  
Solid media



# Aerosol Viability During Manual Steps

Mean cfu/m<sup>3</sup> air detected over 3 experiments

5 X 10<sup>8</sup> cfu BCG spiked into sputum.

Anderson impactor

BioSampler

SR added **15 min wait** then sample pipetted in and out of three Xpert TB cartridge over 15 min time period (equivalent to loading >30 cartridges)

0

0

Sputum smeared/layered on 10 microscope slides over 10 min period.

16

324



