

1 Use of *in vitro* gas production technique for predicting *in vivo* apparent
2 digestibility and voluntary intake of feedstuffs for sheep

3
4 F. Cadario¹

5
6 ¹ Department of Agriculture, the University of Reading, Early Gate, Reading Berks RG6 2AT
7

8 **Abstract**

Twenty four Suffolk x Kent wether lambs were used to measure the dry matter (DM) digestibility and DM intake of twelve feeds, six being classified as "long forages" and the rest were milled "other forages". Animals and feeds were divided into three groups of 4x4 latin squares. A gas production assay was carried out using the Theodorou et al. (1994) technique and incubating the samples in either Theodorou (contains nitrogen) or Menke (contains no nitrogen) media. For gas production there were significant effects ($P < 0.001$) of "media", "feeds" and "feed x medium", for both groups of forages. In vivo DM digestibility and DM intake of "long forages" were highly correlated with in vitro DM digestibility of Menke medium (R^2 0.92 $P < 0.01$, and 0.96 $P < 0.001$), but for Theodorou medium only digestibility was correlated (R^2 0.69 $P < 0.05$). The in vivo digestibility of "other forages" was correlated with the Theodorou medium (R^2 0.77 $P < 0.05$). Prediction of in vivo DM digestibility and DM intake of "long forages" was possible from Menke cumulative gas production (R^2 0.97 $P < 0.001$ and 0.99 $P < 0.001$) or Theodorou gas production (R^2 0.90 $P < 0.01$ and 0.90 $P < 0.01$). For "other forages" only DM digestibility was correlated (R^2 0.87 $P < 0.01$) with the Theodorou medium and there was no correlation with the Menke medium.

9 **Keywords:** *forages in vivo digestibility, intake, in vitro gas production*

10
11 **Introduction**

12
13 The quantity of feed consumed is fundamental to nutrition: it determines the level of
14 nutrients ingested and therefore the animal's response and function. Digestibility and
15 utilisation of nutrients are qualitative descriptions of the net feed intake (Van Soest,
16 1994). *In vivo* measurement of intake and digestibility may be a good guide to the
17 potential performance of animals on particular diets, but their measurement is time
18 consuming, involves a high labour input and requires a large number of animals of the
19 same age, breed and sex to minimise experimental variation (Prasad *et al.* 1994). The
20 prediction of digestibility and intake of feeds has been the subject of wide research,

1 where several techniques have been proposed in an attempt to simulate the ruminant
2 digestive process. The Tilley and Terry (1963) technique has been the most widely used
3 *in vitro* technique, as has the enzymatic method of Jones and Hayward (1975). The *in*
4 *situ* technique (Ørskov *et al.* 1980), although inside the rumen, has the limitation of the
5 restricted number of samples that can be measured at a given time. According to Menke
6 *et al.* (1979), techniques such as the Tilley and Terry (1963), Jones and Hayward (1975),
7 and Ørskov *et al.* (1980) have made important contributions in feedstuff evaluation, but
8 the techniques are still time consuming and imprecise. Looking for some other alternative
9 method, Menke *et al.* (1979) developed a simple technique based on the Tilley and Terry
10 (1963) method which consists of measuring gas production during *in vitro* incubation of
11 feedstuffs in rumen liquor. Menke *et al.* (1979) achieved good results in predicting
12 digestibility and metabolizable energy of feeds. Following the same procedure, Khazaal
13 *et al.* (1993) had success in predicting voluntary intake and dry matter digestibility of
14 legume hay. Khazaal *et al.* (1995) also predicted voluntary intake of graminaceous hay
15 from the gas production technique. Similar results were obtained by Blummel and
16 Ørskov (1993) in predicting voluntary intake and dry matter digestibility of cereal
17 straws.

18
19 Theodorou *et al.* (1994) proposed some changes to the Menke *et al.* (1979) technique.
20 The changes involved the incubation of samples in glass bottles, the use of a pressure
21 transducer for gas readings and the addition of nitrogen to the inoculum. Following the
22 Theodorou *et al.* (1994) suggestions, Prasad *et al.* (1994) achieved good results in
23 predicting digestibility of straws. Adesogan *et al.* (1995) and Sileshi *et al.* (1995) also
24 investigated using the gas technique of Theodorou *et al.* (1994) to predict *in vivo* and *in*
25 *situ* digestibility of forages.

26
27 The prediction of digestibility and intake through the gas production technique, has
28 generated much research interest. Both the Menke *et al.* (1979) and the Theodorou *et al.*
29 (1994) procedures have been researched, however, the two techniques have not been
30 directly compared in any published paper. The present study involves assessing the
31 reliability of the Theodorou and Menke media in the Theodorou *et al.* (1994) technique
32 for predicting digestibility and intake of a variety of forages. Two contrasting groups of

1 forages were compared i.e. “long forages” and “other forages”. “Long forages” were
2 hays, whereas “other forages” involved milled forages. It was hypothesised that the
3 relationship between *in vivo* digestibility and intake would be different for the two
4 groups of forages as also would the correlation between *in vivo* digestibility or intake
5 and gas production.

6 7 **Material and Methods**

8 9 *In vivo trial design*

10 Twenty four Suffolk x Kent wether lambs, averaging 20 kg live weight, were placed in
11 individual pens, with no bedding material. The lambs were divided into three groups of
12 eight animals distributed in a 4 x 4 latin square. The diets were twelve commercial feeds
13 (Table 1) which were divided into two groups, “long forages” and “other forages”.
14 “Other forages” were milled forages whereas “long forages” were unmilled hays. All the
15 feeds were given as single diets *ad libitum* where four of them were randomly allocated
16 in each latin square. Each latin square period consisted of three days of changeover,
17 followed by fourteen days of adaptation to the new feed and the last seven days involved
18 faecal collection for digestibility determination.

19 20 *Chemical analysis and measurements*

21 Representative samples for each feed in each period were taken and milled through a 1.0
22 mm sieve for chemical analysis and for the gas production assay. The DM content was
23 determined by drying the samples to a constant weight at 65 °C. The concentration of
24 nitrogen was determined by using the Kjeldal method and acid detergent fibre was
25 determined according to the Goering and Van Soest (1970) method. At the end of each
26 period, voluntary daily intake and apparent *in vivo* dry matter digestibility were
27 determined

28 29 *In vitro gas production*

30 The method described by Theodorou *et al.* (1994) was used. This consisted of the
31 anaerobic fermentation of dried ground feed samples with rumen microbes. One gramme
32 samples (in triplicate) were transferred under anaerobic conditions into 125 ml capacity

glass serum bottles together with 90 ml of a buffered microbiological medium, prepared
2 according to Theodorou *et al.* (1994) or Menke *et al.* (1979) (Appendix A), and the
bottles were sealed, with rubber stoppers. The mixture was inoculated with microbes
4 prepared from fresh rumen liquor collected from two fistulated sheep fed hay and
5 concentrate (75 : 25) and was fermented anaerobically at 39 °C.

6
7 After inoculating, all the bottles were readjusted to atmospheric pressure, using the
8 pressure transducer. Bottles were then placed in the incubator and this time was
9 considered as the starting point of the experiment i.e. 0^{h-1}. Gas production readings were
10 made after 3, 6, 9, 12, 16, 20, 24, 28, 33, 39, 45, 52, 60 and 70 h of incubation.

11 Measurement was made with a pressure transducer (Bailey and Mackey Ltd,
12 Birmingham B42 1DE, UK) which measured the head space pressure in the bottles. At
13 the end of the gas production run, the content of each bottle was vacuum filtered
14 through pre-weighed filter crucibles (Sintaglass porosity 1, 70 ml capacity; Gallenkamp,
15 Loughborough, UK) which were then oven dried overnight, cooled and weighed again.

16 *Computation of data and statistical analysis*

18 A spreadsheet Excel 5.0 PC program was used, where the cumulative gas production
19 data were adjusted to that produced from 1g dry matter (DM) of sample fermented. *In*
20 *vitro* digestibility (g/kg DM) was calculated assuming that all the residual DM after 70 h
21 of fermentation was unfermented substrate. An analysis of variance, using the Statistical
22 Analysis System (SAS 1985), was made to test the cumulative gas production at each
23 time of reading, using feeds, medium (Theodorou and Menke media) and the interaction
24 between feed x medium.

25
26 The cumulative gas production data were fitted to the France *et al.* (1993) model (MLP
27 1987) with the equation $y = A - B Q^t Z \sqrt{t}$ where $Q = e^{-b}$, $Z = e^{-c}$ and $B = e^{bt + ct}$. Here y
28 denotes cumulative gas production (ml), t is incubation time (h), A is the asymptotic
29 value for gas pool size (ml), t is the lag time and b(h⁻¹) and c (h^{-0.5}) are rate constants. A
30 combined fractional rate (h⁻¹) of gas production (μ) was calculated as $\mu = b + c/2\sqrt{t}$,
31 where t is the incubation time. A t-test was also made to evaluate the means of each
32 parameter of the France *et al.* (1993) equation and for the *in vitro* DM digestibility of

1 both media. The *in vivo* parameters DM digestibility and DM intake were correlated with
2 cumulative gas production at each time of reading with *in vitro* DM digestibility, and
3 with acid detergent fibre (ADF) and crude protein (CP) content of the two groups of
4 feeds, using simple regressions.

5

6 **Results**

7

8 The *in vivo* DM digestibility, DM intake, and chemical analysis (CP and ADF) are
9 presented in Table 1. The feeds were divided into two categories, being the “long
10 forages” with high fibre and low protein content, whereas the “other forages” consisted
11 of a variety of feeds. Hi fi light and grass pellets, although of high fibre content, were
12 classified in the second group because of their small particle size making them more
13 similar to the rest of “other forages”.

14

15 **Table 1**

16

17 Table 2 presents the means of cumulative gas production at each time of reading, for
18 both the “long” and “other” forages. Gas production was higher for the Theodorou
19 medium than the Menke. For both groups of forages, the differences were highly
20 significant throughout, except for the first 3 h. The “feed” factor was highly significant
21 as expected. The “feed x medium” interaction of “long forages” was significant, in the
22 late stage of fermentation, but the F values were small compared with those of the “other
23 forages”.

24

24 **Table 2**

25

26 Table 3 shows the parameters of the France *et al.* (1993) model for the Theodorou and
27 Menke media. The means of the “long” and “other” forages were tested through a t-test
28 to look at the differences between media for each component of the equation. There was
29 a significant ($P<0.001$) difference for the rate of gas production (μ) for the “long
30 forages”, and also to a lesser extent ($P<0.01$) for the Z parameter of the same group.
31 Other parameters were not significantly different. Table 3 also shows that DM
32 digestibility values of both media were different ($P<0.001$).

1

2

Table 3

3

4 The relationships between *in vivo* DM digestibility and DM intake and cumulative gas
production at each of the incubation times were investigated (Appendix C). Similarly the
6 relationships between *in vivo* DM digestibility and DM intake and the other measured
7 parameters were investigated (Tables 4, 5 and Appendix B). For “long forages”,
8 cumulative gas production was best correlated with DM digestibility at 3 h for Theodorou
9 medium and at 28 h for Menke medium. For DM of “long forages” the best correlation
10 with cumulative gas production occurred at 3 h for Theodorou medium and at 70 h for
11 Menke, as is shown in Tables 4 and 5.

12

Table 4

13

14 Tables 4 and 5 also show that for “other forages” except for *in vivo* DM digestibility and
15 cumulative gas production at 28 h for Theodorou medium (R^2 0.87 $P < 0.01$) there were
16 no significant correlations ($P > 0.05$) between cumulative gas production and *in vivo* DM
17 digestibility and DM intake.

18

19

Table 5

20

21 Discussion

22

23 *In vivo* DM digestibility and DM intake

24 The relationship between digestibility and intake was highly significant for the “long
25 forages”, but not for the “other forages” (Tables 4 and 5), thus confirming the
26 hypothesis. In Table 1 it is seen that the two lowest DM digestibilities are from the grass
27 pellets and the hi fi, where pellets had the highest intake, despite its high CP and ADF
28 content. This is explained by Forbes, (1993) who commented “some feeds may be poorly
29 digested, but pass through the digestive tract relatively quickly, thereby occupying space
30 in the rumen for less time than a more digestible feed with a slower rate of passage such
31 as the case of ground low quality forages. Although digestibility is relatively easy to
32 measure, is probably not the most useful measurement for predicting intake”.

1 A very different feature was observed in the case of sugar beet pulp which had the lowest
2 intake rate of its group, but the highest DM digestibility; this could be explained by the
3 fact of its highly soluble material content and highly digestible fibre. In this case was
4 probably determined by the animal's metabolic regulation (Forbes, 1993).

5

6 *Theodorou and Menke media*

7 The differences between the two media in *in vitro* digestibility were large and highly
8 significant (Table 2). As expected, there were also differences between feeds and their
9 interactions. Both media were basically the same, with some exceptions. The Theodorou
10 medium is more dilute and in the solution B (Appendix A) it has NH_4CO_3 , whereas the
11 Menke has none; NH_4CO_3 is a source of nitrogen and also a buffer. In solution B,
12 Menke has four more grammes of NaHCO_3 . In solution C, Menke has 3.75 g/l less
13 Na_2HPO_4 than the Theodorou. Cysteine is a reducing agent in the Theodorou medium
14 but is not used in Menke. The Theodorou medium contains nitrogen whereas the Menke
15 does not. The nitrogen content is likely to be the main cause of differences between the
16 two media. The Theodorou medium produced more gas and higher DM digestibility
17 (Tables 2 and 3) especially in the case of "long forages" which generally had low CP
18 content. According to Preston and Leng (1987), "deficiency of a nutrient needed by the
19 rumen micro-organisms, will reduce the microbial activity and therefore reduce feed
20 digestibility", particularly fibrous feeds, and probably the primary limitation to this is the
21 supply of nitrogen to the micro-organisms in the bottles. However, in the present study,
22 for "other forages" which had more CP content, the differences in *in vitro* digestibility
23 between the two media, were less although the Theodorou medium gave a higher
24 digestibility, this was not significant. ($P>0.05$) (Table 3).

25

26 There were also differences between the two media in cumulative gas production (Table
27 2) with the Theodorou medium producing more gas. The differences were probably due
28 to presence of extra nitrogen and cysteine for microbial activity and causing it higher
29 rates of fermentation and consequently production of gas (Russell and Trobel, 1993).
30 This was particularly the case for "long forages". On the other hand, "other forages"
31 were unlikely to have nitrogen deficiency (Table 1), even in the Menke medium, but it is
32 possible that pH fell because the Menke medium had a lower buffering capacity, and

consequently microbial activity and gas production may have been impaired. A question which arises is how much nitrogen has to be added to the Menke medium or how much nitrogen is in excess in the Theodorou medium?. A very recent investigation at the Natural Resources Institute, Chatham, by Rosales (1996) has addressed this question in the fermentation of tropical tree forages.

In vivo parameters and cumulative gas production

For “long forages” and the Menke medium, *in vivo* DM digestibility was highly correlated with gas production, reaching the highest correlation at 28 h of incubation. For the Theodorou medium the highest correlation occurred at 3 h, but was also high at 16 h (R^2 0.88), but after 16 h the correlation declined. “Long forages” DM intake was highly correlated with gas production with the Menke medium, being the highest correlation at 70 h (R^2 0.99), with the Theodorou medium the highest correlation being at 3 h (R^2 0.90) and thereafter declined (at 70 h; R^2 0.50) (Appendix C). These results show that DM digestibility and DM intake of “long forages” can be predicted at early stages of gas production using either Theodorou or Menke media, This confirms the results of Blummel and Ørskov (1993), Khazaal *et al.* (1995) and Prasad *et al.* (1994).

For “other forages”, the gas production gave poor prediction of *in vivo* digestibility and intake, with the only good correlation (R^2 0.87) being between *in vivo* DM digestibility and the gas at 28 h for the Theodorou medium. Kibon and Ørskov (1993), using a Menke medium, also found a poor correlation between gas production and *in vivo* DM digestibility of browse species. In the present study with “other forages”, the poor correlation between gas production and intake is understandable because of the fact that “other forages” were milled and intake would not be limited by physical capacity of the reticulo-rumen. The study by Kibon and Ørskov (1993) involved browse species; it is likely that the tannin content of the browse reduced the availability of nitrogen in the medium.

In the present study there were large and significant differences between gas production between the Theodorou and Menke media. For “long forages” the results suggest that *in vivo* DM digestibility and DM intake may be predicted from the *in vitro* gas production

1 technique using either the Theodorou or Menke media. Furthermore, *in vivo* digestibility
2 and intake of “long forages” can also be predicted from the *in vitro* digestibility of “long
3 forages” using the Menke medium and digestibility from the Theodorou medium. The
4 results are less clear for predicting *in vivo* digestibility and intake of milled “other
5 forages” using the gas production technique. More research is required concerning milled
6 forages.

7

8 **Acknowledgements**

9

10 The author would like to thank the assistance of Dr. E. Owen (supervisor) and Dr. D.
11 Romney during the conduct of the experiment and the preparation of the paper. Also Dr.
12 Maguie Gill, as well as the staff members of Natural Resources Institute (NRI)
13 laboratory at Wye, Ashford. The assistance and advice of Dr. M.J. Bryant (MSc course
14 tutor) is acknowledged. The author is grateful for a scholarship form Overseas
15 Development Administration (ODA) and a special acknowledgement to John Wilkins for
16 all his co-operation.

17

18 **References**

19

20 **Adesogan, A.T., Givens, D.C. and Owen, E.** 1995. A comparison between *in vitro*
21 digestibility, *in situ* degradability and a gas production technique for predicting the *in vivo*
22 digestibility of whole crop wheat. *Journal of Animal Science* 62 (3): 631

23

24 **Blummel, M. and Ørskov, E.R.** 1993. Comparison of *in vitro* gas production and
25 nylon bag degradability of roughages in predicting feed intake in cattle. *Animal Feed*
26 *Science and Technology* 40: 109 - 119

27

28 **Cadario, F.** 1996. Use of *in vitro* gas production technique for predicting *in vivo*
29 apparent digestibility and voluntary intake of feedstuffs for sheep. *Literature Review*
30 *Submitted in Partial Fulfilment of the Requirements for the MSc Degree in Animal &*
31 *Forage Science.* University of Reading.

32

33 **Forbes, J.M.** 1993. Voluntary Feed Intake in *Quantitative Aspects of Ruminant*
34 *Digestion and Metabolism.* (Eds. J.M. Forbes and J.D. Cole) C.A.B. International, UK.

35

36 **France, J., Dhanoa, M.S., Theodorou, M.K., Lister, S.J., Davies, D.R. and Isaac,**
37 **D.C.** 1993. A model to interpret gas accumulation profiles associated with *in vitro*
38 degradability of ruminant feeds. *Journal of Theoretical Biology* 163: 99 - 111

39

- 1 **Goering, H.K. and Van Soest, P.J.** 1970. Forage fibre analysis. Agriculture Handbook.
2 # 379. USDA, *Agricultural Research Service*. Washington, DC.
3
- 4 **Jones, D.I.H. and Hayward, M.V.** 1975. The effect of pretreatment of herbage on the
5 prediction of dry matter digestibility from solubility in fungal cellulase solutions. *Journal*
6 *of Science Food and Agriculture* 26: 711 - 718
7
- 8 **Khazaal, K., Dentinho, M.T., Ribeiro, J.M. and Ørskov, E.R.** 1993. A comparison
9 of gas production during incubation with rumen contents *in vitro* and nylon bag
10 degradability as predictors of the apparent digestibility *in vivo* and the voluntary intake of
11 hays. *Journal of Animal Production* 57: 105 - 112
12
- 13 **Khazaal, K., Dentinho, M.T., Ribeiro, J.M. and Ørskov, E.R.** 1995. Prediction of
14 apparent digestibility and voluntary intake of hays fed to sheep: comparison between
15 using fibre components, *in vitro* digestibility or characteristics of gas production or nylon
16 bag degradability. *Journal of Animal Science* 61: 527 - 538
17
- 18 **Kibon, A. and Ørskov, E.R.** 1993. The use of degradation characteristics of browse
19 plants to predict intake and digestibility by goats. *Journal of Animal Production* 57:
20 247 - 251
21
- 22 **Menke, K.M., Raab, L., Salewski, A., Steingass, H., Fritz, D. and Scheneider, W.**
23 1979. The estimation of the digestibility and metabolizable energy content of ruminant
24 feedingstuffs from the gas production when they are incubated with rumen liquor *in*
25 *vitro*. *Journal of Agricultural Science* 93: 217 - 222
26
- 27 **Ørskov, E.R., Hovell, F.D. and Mould, F.** 1980. The use of the nylon bag technique
28 for the evaluation of feedstuffs. *Tropical Animal Production* 5: 195 - 213
29
- 30 **Prasad, C.S., Wood, C.D. and Sampath, K.T.** 1994. Use of *in vitro* gas production to
31 evaluate rumen fermentation of untreated and urea treated finger millet straw (*Eleusine*
32 *coracana*) supplemented with different levels of concentrate. *Journal of Food Science*
33 *and Agriculture* 65: 457 - 464
34
- 35 **Preston, T.R. and Leng, R.A.** 1987. *Matching Ruminant Production Systems with*
36 *Available Resources in the Tropics and sub Tropics*. Penambul Books: Armidale.
37 Australia.
38
- 39 **Rosales, M.** 1996. *In vitro* assessment of the nutritive value of mixtures of leaves form
40 tropical fodder trees. *PhD thesis. University of Oxford*.
41
- 42 **Ross, G.J.S.** 1987. MLP. Maximum Likelihood Program (a manual). Rothamsted
43 Experimental Station. Harpenden, UK.
44
- 45 **Russell, J.B. and Strobel, H.J.** 1993. Microbial Energetics in *Quantitative Aspects of*
46 *Ruminant Digestion and Metabolism*. (Eds. J.M. Forbes and J.D. Cole) C.A.B.
47 International, UK.
48

1 **Sileshi, Z., Owen, E., Dhanoa, M.S. and Theodorou, M.K.** 1996. Prediction of *in situ*
2 rumen dry matter disappearance of Ethiopian forages from an *in vitro* gas production
3 technique using a pressure transducer, chemical analysis or *in vitro* digestibility. *Animal*
4 *Feed Science and Technology* (Inpress)
5
6 **Statistical Analysis Systems Institute Inc.,** 1985. *User's Guide: Statistics SAS*
7 *Institute Inc., Cary. NC.*
8
9 **Theodorou, M.K., Williams, B.A., Dhanoa, M.S., Mc Allan, A.B. and France, J.**
10 1994. A simple gas production method using a pressure transducer to determine the
11 fermentation kinetics of ruminant feeds. *Animal Feed Science and Technology* **48**: 185 -
12 197
13
14 **Tilley, J.M.A. and Terry, R.A.** 1963. A two stage technique for the *in vitro* digestion
15 of forage crops. *Journal of British Grassland Society* **18**: 104 - 111
16
17 **Van Soest, P.J.** 1994. *Nutritional Ecology of the Ruminant.* Cornell University Press.
18 Ithaca and London.
19
20
21
22

1 **Table 1** *DM intake and in vivo DM digestibility of the feedstuffs used*

2

Feedstuffs	Crude protein (g/kgDM)	Acid detergent fibre(g/kg DM)	DM intake (g/kg M. ^{0.75} day)	s.d.	DM digestibility (g/kg)	s.d.
<i>Long forages</i>						
Dried rye grass	59	421	58	6.1	584	24
Timothy hay	95	409	55	7.2	561	39
Meadow hay	57	503	50	5.9	536	55
Rye grass hay	63	493	48	4.4	518	76
Straw 2	37	555	26	6.0	475	42
Straw 1	41	598	27	7.1	457	52
<i>Other feeds</i>						
Sugar beet pulp	99	258	53	23.3	817	65
Maize gluten feed	213	200	86	25.4	702	49
Wheat feed	171	170	58	10.2	672	51
Lucerne	192	367	120	9.4	636	24
Hi fi light†	108	515	67	22.1	570	58
Grass pellets	194	459	157	29.8	495	23

† A mixture of lucerne and oat straw

Table 2 Cumulative gas production for different incubation times for Theodorou and Menke media; F value and significance of the effect of medium, feed and medium*feed interaction

Time (h)	Cumulative gas production Theodorou (ml)	Cumulative gas production Menke (ml)	Medium F value	Feed F value	Feed*medium F value
<i>Long forages</i>					
3	14.6	12.5	3.3 NS	6.68 ***	0.19 NS
6	35.1	29.4	18.77 ***	61.82 ***	0.41 NS
9	53.0	43.0	43.57 ***	91.04 ***	0.52 NS
12	72.7	54.5	105.61 ***	90.25 ***	0.61 NS
16	99.4	65.9	273.54 ***	84.55 ***	1.02 NS
20	122.7	76.4	444.88 ***	84.41 ***	1.68 NS
24	142.3	85.5	557.84 ***	82.29 ***	2.46 NS
28	160.7	94.3	629.2 ***	77.61 ***	2.94 *
33	181.1	104.3	648.0 ***	67.65 ***	3.39 *
39	203.5	115.6	632.42 ***	54.08 ***	4.34 **
45	223.3	126.9	643.38 ***	49.01 ***	5.26 **
52	241.2	139.7	603.23 ***	44.42 ***	5.90 **
60	257.1	154.0	491.23 ***	38.41 ***	6.33 ***
70	270.2	168.4	369.49 ***	32.86 ***	6.34 ***
5	<i>Other forages</i>				
3	14.2	13.1	1.52 NS	0.52 NS	0.56 NS
6	46.3	36.1	40.33 ***	40.09 ***	8.6 ***
9	77.1	61.0	76.51 ***	97.93 ***	12.39 ***
12	104.2	83.4	90.56 ***	103.29 ***	14.42 ***
16	133.8	105.7	123.32 ***	89.92 ***	22.73 ***
20	158.8	123.9	176.54 ***	87.17 ***	31.75 ***
24	178.6	138.5	215.25 ***	81.66 ***	40.27 ***
28	197.0	151.6	258.90 ***	78.92 ***	52.13 ***
33	214.3	165.0	262.26 ***	65.98 ***	54.14 ***
39	230.1	178.5	233.41 ***	45.88 ***	47.95 ***
45	242.8	191.9	205.20 ***	35.30 ***	44.21 ***
52	255.0	205.5	194.31 ***	32.06 ***	42.81 ***
60	266.8	219.6	173.45 ***	31.96 ***	36.33 ***
70	277.0	233.6	140.48 ***	38.59 ***	25.85 ***

Table 3 Estimated values for kinetics parameters from the fermentation and digestibility means of “1

2 Menke media

3

Feedstuffs	Lag time (T)		Fractional rates				Gas pool size	
	(h)		Q (h ⁻¹)		Z (h ^{-0.5})		(A) (ml)	
	Th ¹	Mk ²	Th ¹	Mk ²	Th ¹	Mk ²	Th ¹	Mk ²
<i>Long forages</i>								
Dried rye grass	1.072	0.711	0.96757	0.99992	1.00737	0.98285	370.56	417.16
Timothy hay	0.92	1.273	0.96288	0.99651	1.03585	0.96149	284.4	497.51
Meadow hay	1.394	1.222	0.95982	0.99218	1.10171	0.96203	312.52	340.44
Rye grass hay	1.0	0.77	0.97192	0.99909	1.00532	0.97307	313.22	738.92
Straw 2	2.14	1.427	0.96499	0.98965	1.10149	0.96847	290.09	174.48
Straw 1	1.84	1.36	0.96366	0.99843	1.11448	0.96237	306.13	346.69
Mean	1.394 ^a	1.127 ^a	0.96514 ^a	0.99596 ^a	1.061 ^w	0.968 ^x	312.82 ^a	446.42 ^a
<i>Other forages</i>								
Sugar beet	1.01	1.71	0.94952	0.9995	1.02545	0.96304	379.07	886.28
Maize gluten	2.44	2.24	0.96838	0.95625	0.896	0.95323	287.64	274.33
Wheat feed	2.49	2.29	0.97646	0.94821	0.76876	0.92104	262.6	230.35
Lucerne	1.192	1.66	0.95048	0.97998	1.0746	0.98191	262.9	293.94
Hi fi light	1.295	1.02	0.96204	0.99675	1.09209	0.98418	321.3	749.33
Grass pellets	1.295	0.78	0.9528	0.95211	1.05527	1.0906	242.78	232.58
Mean	1.787 ^a	1.616 ^a	0.9599 ^a	0.9721 ^a	0.985 ^a	0.982 ^a	292.15 ^a	447.47 ^a

¹ Th = Theodorou medium

² Mk = Menke medium

^{aa} = Non significant

^{wx} = **

^{yz} = ***

1 **Table 4 Relationship between *in vivo* DM digestibility and various measured parameters**

2

Parameter y	Parameter x	Equation
<i>Long forages</i>		
<i>In vivo</i> DMD (g/kg)	DM intake (g/kg)	$y = 3.3655x + 374.19$
	Acid detergent fibre (g/kg DM)	$y = -0.6397x + 839.31$
	Crude protein (g/kg DM)	$y = 1.6795x + 423.18$
	DM digestibility Theodorou (g/kg)	$y = 0.5828x + 140.59$
	DM digestibility Menke (g/kg)	$y = 0.377x + 359.79$
	Cumulative gas production at 3 h Theodorou (ml) †	$y = 13.909x + 318.86$
	Cumulative gas production at 28 h Menke (ml) †	$y = 1.7417x + 357.41$
<i>Other forages</i>		
<i>In vivo</i> DMD (g/kg)	DM intake (g/kg)	$y = -1.9242x + 822.29$
	Acid detergent fibre (g/kg DM)	$y = -0.5765x + 837.88$
	Crude protein (g/kg DM)	$y = -0.7581x + 722.11$
	DM digestibility Theodorou (g/kg)	$y = 0.787x + 60.676$
	DM digestibility Menke (g/kg)	$y = 0.1263x + 567.14$
	Cumulative gas production at 28 h Theodorou (ml) †	$y = 2.4191x + 172.15$
	Cumulative gas production at 70 h Menke (ml) †	$y = 3.8487x - 250.52$

† For data in Appendix C

1 **Table 5** Relationship between DM intake and various measured parameters

2

Parameter y	Parameter x	Equation
<i>Long forages</i>		
Dry matter intake (g/kg)	<i>In vivo</i> DM digestibility (g/kg)	$y = 0.2775x - 100.94$
	Acid detergent fibre (g/kg DM)	$y = -0.1766x + 131.54$
	Crude protein (g/kg DM)	$y = 0.5165x + 13.528$
	DM digestibility Theodorou (g/kg)	$y = 0.1534x - 56.521$
	DM digestibility Menke (g/kg)	$y = 0.111x - 3.8308$
	Cumulative gas production at 3 h Theodorou (ml) †	$y = 4.0294x - 14.932$
	Cumulative gas production at 70 h Menke (ml) †	$y = 0.2904x - 5.0544$
<i>Other forages</i>		
DM intake (g/kg)	<i>In vivo</i> DM digestibility (g/kg)	$y = -0.2609x + 259.49$
	Acid detergent fibre (g/kg DM)	$y = 0.1376x + 45.06$
	Crude protein (g/kg DM)	$y = 0.5362x + 291.97$
	DM digestibility Theodorou (g/kg)	$y = -0.2442x + 272.7$
	DM digestibility Menke (g/kg DM)	$y = -0.0478x + 121.11$
	Cumulative gas production at 3 h Theodorou (ml) †	$y = -35.644x + 596.68$
	Cumulative gas production at 6 h Menke (ml) †	$y = -1.762x + 153.76$

† For data in Appendix C

1 **Appendix A** Chemical composition of Theodorou and Menke media

2

Theodorou medium

To give 900 ml of medium

Microminerals (A)	0.1 ml
Buffer (B)	200 ml
Macrominerals (C)	200 ml
Resarzurin	1.0 ml
Distilled water	500 ml

Buffer (g/l)

NH_4HCO_3

NaCHO 35.0 g

Macrominerals (g/l)

$\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$ 9.45 g

KH_2PO_4 6.2 g

$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.6 g

Reducing agent

Cysteine HCl. $1\text{H}_2\text{O}$ 0.625 g

Distilled water 95.0 ml

1M NaOH 4.0 ml

Sodium sulphide 0.625 g

Menke medium

To give 900 ml of medium

Solution A	0.1 ml
Solution B	200 ml
Solution C	200 ml
Resarzurin	1.0 ml
Distilled water	400 ml

Solution B (g/l)

Na HCO_3 39.0 g

Solution C (g/l)

Na_2HPO_4 (anhydrous) 5.7 g

KH_2PO_4 6.2 g

$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.6 g

Reducing agent

Distilled water 95.0 ml

1M NaOH 4.0 ml

Sodium sulphide 0.625 ml

1 **Appendix B** *Correlation among all measured parameters*

2

Long forages

	<i>in vivo</i> DMD	DM intake	CG70 Th	CG 70 Mk	DMD Th	DMD Mk
<i>in vivo</i> DMD		0.93				
DM intake						
CG 70 Th	0.63	0.50				
CG 70 Mk	0.96	0.99				
DMD Th	0.69	0.58	0.98			
DMD Mk	0.92	0.96		0.95		
CP	0.49	0.57	0.01	0.55	0.04	0.57
ADF	0.92	0.85	0.38	0.86	0.47	0.90

Other forages

	<i>in vivo</i> DMD	DM intake	CG70 Th	CG 70 Mk	DMD Th	DMD Mk
<i>in vivo</i> DMD		0.5				
DM intake						
CG 70 Th	0.66	0.45				
CG 70 Mk	0.4	0.09				
DMD Th	0.77	0.54	0.72			
DMD Mk	0.02	0.02		0.05		
CP	0.1	0.39	0.56	0.02	0.17	0.24
ADF	0.54	0.22	0.06	0.11	0.37	0.36
