

# Isolation and characterisation of an ACC oxidase gene in cassava

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## Introduction

Ethylene is involved in a wide range of physiological processes in plants including germination, abscission, senescence, flowering and fruit ripening. Ethylene production is also induced by stresses such as wounding, drought, chilling and pathogen attack. During cassava postharvest physiological deterioration (PPD), ethylene production in the roots was found to increase after a lag of 6 to 16 hours after harvesting (Plumbley *et al.*, 1981). 1-Aminocyclopropane-1-carboxylate (ACC oxidase or ACO) catalyses the last step of ethylene biosynthesis in plants. A number of ACC oxidase genes have been cloned, mainly from ripening fruits or flowers and some from vegetative tissues of different plant systems (Lasserre *et al.*, 1996). In most plant systems studied, ACC oxidase was encoded by a multigene family. An ACC oxidase cDNA clone, designated as MeACO1, has been isolated and characterized from a PPD-related cDNA library (Han, 2000). Here we report the isolation and characterization of an ACC oxidase genomic clone.

## Material and Methods

About  $2.3 \times 10^6$  plaques of a genomic library of cassava cultivar MBRA 534 DNA in  $\lambda$ EMBL3, a generous gift of Professor M A Hughes, University of Newcastle, were screened with the cDNA MeACO1 probe (Han, 2000) and one positive clone was isolated. PCR was carried out using the lysate of the positive plaque and primers designed from cMeACO1. The sequence of PCR product showed high homology to ACC oxidase genes from other plants. However, when compared with the cDNA clone, MeACO1, the similarity was 90% over a 300 bp length. Therefore, the isolated clone was an ACC oxidase clone but not the one corresponding to cDNA MeACO1. This genomic clone was designated gMeACO2.

The  $\lambda$  DNA of gMeACO2 clone was extracted, purified and digested respectively with *Hind*III and *Kpn*I, and the resulted two fragments (6 kb and 2 kb) were cloned into the pUC19 vector and sequenced. Putative exons, introns and promoter regions were identified by Genefeature at <http://vega.igh.cnrs.fr/bin/align-guess.cgi>, and by comparing the gene with other ACC oxidase genes.

## Results and Discussion

The determined sequence of the ACC oxidase gene, gMeACO2, was 6349 bp, including 4928 bp of 5' flanking region and four putative exons interrupted by three introns. The deduced amino acid sequence was 312 amino acids.

Motifs or putative regulatory elements similar to those in ACO promoters of other plant gene promoters were identified in the 5' flanking regions of the gMeACO2. The TCA motif (TCATCT-TCTT) and near matches, which were present in over 30 stress-induced genes (Goldsbrough *et al.*, 1993), was identified in gMeACO2 (at -723 to -733). Ethylene-inducible elements

ATT-TCAAA or AATTCAAA were initially identified in the E4 gene in tomato and glutathione-S-transferase (GST1) gene in carnation (Montgomery *et al.*, 1993; Itzaki *et al.*, 1994), and also present in ethylene-inducible ACO genes in tomato and mung bean (Blume *et al.*, 1997). There were two such motifs in the cassava gMeACO2 (-471 to -465, -2127 to -2121 and -2431 to -2424).

ACC oxidase genes from different plant species are of high similarity both in their nucleic acid and deduced amino acid sequences. Cassava gMeACO2 had 85% similarity to the ACC oxidase gene from *Prunus persica* in the coding region (Tang *et al.*, 1993). The isolated cassava ACO genes, cDNA MeACO1 and gMeACO2, shared very high amino acid sequence identity with a number of ACO genes from other plants such as *Carica papaya* (84% identity and 88% similarity, gene access number: U68215), *Prunus persica* (81% and 89%, AF129074), petunia (81% and 86%, Q08507) and *Nicotiana glutinosa* (79% and 88%, U54565). To demonstrate the high similarity, the deduced amino acid sequence of the two cassava genes were compared with four tomato ACO genes, a well-characterized ACO gene family (Figure 1). As can be seen, the deduced amino acid sequences of these ACO genes were highly conserved from the N terminal to the C terminal except for two short regions near the C terminal. The identities between gMeACO2 and tomato ACO genes, LeACO1, LeACO2, LeACO3 and LeACO3, were 83%, 79%, 82% and 81%, respectively. This means that gMeACO2 was much more similar to tomato LeACO1 and LeACO3 than to MeACO1 (81% identity with gMeACO2).

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## References

- Blume, B.; Barry, C.S.; Hamilton, A.J.; Bouzayen, M.; Grierson, D. (1997) Identification of transposon-like elements in non-coding regions of tomato ACC oxidase genes. *Molecular and General Genetics* 254, 297-303.
- Goldsbrough, A.P.; Albrecht, H.; Stratford, R. (1993) Salicylic acid-inducible binding of a tobacco nuclear protein to a 10 bp sequence which is highly conserved amongst stress-inducible genes. *Plant Journal* 3, 563-571.
- Han, Y. (2000) Molecular analysis of post-harvest physiological deterioration of cassava. University of Bath, UK; PhD thesis
- Itzaki, H.; Maxson, J.M.; Woodson, W.R. (1994) An ethylene-responsive enhancer element is involved in the senescence-related expression of the carnation glutathione-S-transferase (GST1) gene. *Proceedings of the National Academy of Sciences of the United States of America* 91, 8925-8929.

MeACO2	MET-FPVIDLSKLSGEERKPTMEMIQDACENWGGFFELVNHGISHELMDTVERLTKEHYKK
MeACO1	ME--FPVINLEKLNGEERAATMAKIKDACENWGGFFELLNHGIPEFLDRVESMTKGHYRK
LeACO1	MEN-FPIINLEKLNNGDERANTMEMIKDACENWGGFFELVNHGIPHEVMDTVEKMTKGHYKK
LeACO2	MEN-FPIINLEKLNGAERVATMEKINDACENWGGFFELVNHGIPHEVMDTVEKLTKGHYKK
LeACO3	MEN-FPIINLENLNGDERAKTMEMIKDACENWGGFFELVNHGIPHEVMDTVEKLTKGHYKK
LeACO4	MESNFPVVDMLLQTEKRPEAMDKIKDACENWGGFFELVNHGISHELLDAVENLTKGHYKK
	** **: : : * . : * : * * : *
MeACO2	CMEQRFKEMVASKGLEAVQSEISDLLDWESTFFLRHLPVSNMAEIPDLDEEYRKTMKFEFAE
MeACO1	CMEQRFKEMVANKGLDAVQTEIKDMDWESTFFIRHLPDSNLAQLPDLDEHRAVMKFEFAE
LeACO1	CMEQRFKELVASKGLEAVQAEVTDLDWESTFFLRHLPSTNISQVPDLDEEYREVMRDFAK
LeACO2	CMEQRFKELVAKKGLEGVEVEVTDMDWESTFFLRHLPSSNISQLPDLDDVYREVMRDFRK
LeACO3	CMEQRFKELVASKGLEAVQAEVTDLDWESTFFLRHLPSTNISQVPDLDEEYREVMRDFAK
LeACO4	CMEQRFKEMVASKGLEAVQTEIDDLLDWESTFFLRHLPVSNVYEVDPDLDEYRKMVMDKDFAL
	* *
MeACO2	ELEKLAEQLEVLNLCENLGLKGYLKKAFYGSKGPTFGTKVSNYPPCPKPDLIKGLRAHTD
MeACO1	KLEKLAEDLLDLCENLGLKGYLKKAFYGSRGPTFGTKVSNYPPCPKPDLIKGLRAHTD
LeACO1	RLEKLAEEELLDLCENLGLKGYLKNAFYGSKGNFPTGTVSNYPPCPKPDLIKGLRAHTD
LeACO2	RLEKLAEEELLDLCENLGLKGYLKNFYPYGSKGNFPTGTVSNYPPCPKPDLIKGLRAHTD
LeACO3	RLEKLAEEELLDLCENLGLKGYLKNAFYGSKGNFPTGTVSNYPPCPKPDLIKGLRAHTD
LeACO4	KLEKLAENLLDLCENLGLKGYLKKAFYGSKGPTFGTKVSNYPPCPKPDLIKGLRAHTD
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MeACO2	AGGI ILLFQDDKVSGLQLLKDQWFDVPPMKHSIVINIGDQLEVITNGKYKSVHRVIAQ
MeACO1	AGGI ILLFQDDRVSGQLLKDQWIDVPPMRHSIVVNLGDQLEVITNGKYKSVHRVIAQ
LeACO1	AGGI ILLFQDDKVSGLQLLKDQWIDVPPMRHSIVVNLGDQLEVITNGKYKSVLHRVIAQ
LeACO2	AGGI ILLFQDDKVSGLQLLKDGRWIDVPPMRHSIVVNLGDQLEVITNGKYKSVHRVIAQ
LeACO3	AGGI ILLFQDDKVSGLQLLKDQWIDVPPMRHSIVVNLGDQLEVITNGKYKSVHRVIAQ
LeACO4	AGGI ILLFQDDKVSGLQLLKDGNWIDVPPMKHSIVINLGDQLEVITNGRYKSVIEHRVIAQ
	* *
MeACO2	TDGTRMSLASFYNPGSDAVIYPAPALVEK--EAEKSQVYPKFVFDYMKLYAGLKFOAKE
MeACO1	TDGTRMSLASFYNPGSDAVIYPAPALVEKE--AEEKQVYPKFVFDYMKLYVGLKFOAKE
LeACO1	TDGTRMSLASFYNPGSDAVIYPAKTLVEKEAEEES--TQVYPKFVFDYMKLYAGLKFOAKE
LeACO2	KDGTRMSLASFYNPGNDALIYPAPALVDKEAEEHNKQVYPKFMFDDYMKLYANLKFQAKE
LeACO3	TDGTRMSLASFYNPGNDAVIYPAPSLIE---ES--KQVYPKFVFDYMKLYAGLKFOAKE
LeACO4	QDGTRMSIASFYNPGSDAVIFPAPELIEKT--EEDIKLYPKFVFDYMKLYAGLKFOAKE
	* *
MeACO2	PRFEAMKA-----MGP-----IATA-
MeACO1	PRFEAMKAVENNVN--LGPNCYCLINYY
LeACO1	PRFEAMKAMES-----DP-----IASA-
LeACO2	PRFEAMKAMES-----DP-----IAIA-
LeACO3	PRFEAMKAMEANVELVDQ-----IASA-
LeACO4	PRFEAMKAVETTVN--LGP-----IETV-
	* *

Figure 1. Comparison of the deduced amino acid sequences of MeACO1 and MeACO2 with those of four tomato ACC oxidase genes. Access number for tomato ACO genes in GeneBank, LeACO1: X58273, LeACO2: Y00478, LeACO3: Z54199 and LeACO4: AB013101. \* – single, fully conserved residue; : – conservation of strong groups; - – conservation of weak groups; - – no consensus

Lasserre, E.; Bouquin, T.; Hernandez, J.A.; Bull, J.; Pech, J.C.; Balague, C. (1996) Structure and expression of three genes encoding ACC oxidase homologs from melon (*Cucumis melo* L.). *Molecular and General Genetics* 251, 81–90.

Montgomery, J.; Goldman, S.; Deikman, J.; Margossian, L.; Fischer, R.L. (1993) Identification of an ethylene-responsive region in the promoter of a fruit ripening gene. *Proceedings of the National Academy of Sciences of the United States of America* 90, 5939–5943.

Plumbley, R.A.; Hughes, P.A.; Marriott, J. (1981) Studies on peroxidases and vascular discoloration in cassava root tissue. *Journal of Scientific Food and Agriculture* 32, 723–731.

Tang, X.; Wang, H.; Brandt, A.S.; Woodson, W.R. (1993) Organization and structure of the 1-aminocyclopropane-1-carboxylate oxidase gene family from *Petunia hybrida*. *Plant Molecular Biology* 23, 1151–1164.