



SARS-CoV-2 Inactivation Testing: Interim Report

Report identifier	HCM/CoV2/041/v1
Report date	18 September 2020
Undertaken by High Containment Microbiology, NIS Laboratories, National Infection Service, Public Health England N.B. This is an interim report and may be updated as further results are obtained	

Product/treatment details	
Product/treatment	Buffer from Virus RNA Collection Kit
Manufacturer	Xiamen Zeesan Biotech
Product code	401120
Manufacturer's recommended ratio of sample to product	1 volume sample to 2 volumes product

Sample details	
Sample type tested	Tissue culture fluid containing 5% (v/v) foetal calf serum
Virus strain tested	SARS-CoV-2 England 2
Ratio of spiked virus stock to sample matrix	Not applicable; tissue culture fluid used undiluted

Experimental conditions	
Ratio of sample to product tested	1 volume sample to 2 volumes product
Contact time/s	10 minutes; 30 minutes
Temperature of incubation	Room temperature

<p>Brief description of tests performed</p>	<p>Triplicate samples were treated with test buffer for indicated contact time/s or mock-treated in triplicate with an equivalent volume of PBS. All samples were then subjected to a purification step to remove cytotoxic buffer components. PBS-treated samples were subjected to the same purification procedure in parallel.</p> <p>Test 1: Purified samples were immediately titrated on Vero E6 cells to establish virus titre. This test is quantitative and reports the titre of virus in each treatment condition in TCID50 per ml. Reduction in virus titre following treatment is given as the difference between the mean \log_{10} TCID50 for treated conditions and the PBS control.</p> <p>Test 2: In parallel, purified samples were seeded onto Vero E6 monolayers to amplify any remaining virus over the course of up to four serial passages. Virus amplification over each passage was detected by visual (microscopic) examination of monolayers for cytopathic effect, and confirmed by SARS-CoV-2-specific real-time PCR. This test is qualitative and reports either the presence or absence of virus amplification. This test may detect levels of virus that are below the detection limit of the titration assay (test 1) due to a greater sample plating volume and the opportunity for any virus present to amplify over serial passages.</p>
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Table of results			
Maximum detectable virus reduction in test 1 (log ₁₀ TCID ₅₀)			5.7 [†]
	Test 1: Virus titration post-treatment		Test 2: Passage of samples in cell culture
	Mean virus titre (log ₁₀ TCID ₅₀)	Titre reduction (log ₁₀ TCID ₅₀)	Virus detected/ Virus not detected
PBS-treated	7.4	-	Virus detected (all replicates)
Test buffer-treated (10 minutes)	4.1	3.3	Virus detected (all replicates)
Test buffer-treated (30 minutes)	2.9	4.5	Virus detected (all replicates)

[†]Limit of detection was 1.7 log₁₀ TCID₅₀ due to buffer cytotoxicity

Interpretation
<p>Test 1: Treatment with this product for 10 or 30 minutes reduced virus titre by 3.3 and 4.5 log₁₀, respectively. Virus was detectable after treatment for all conditions.</p> <p>Test 2: Virus was detectable after treatment for all conditions.</p> <p>This test has been performed on tissue culture fluid containing 5% (v/v) foetal calf serum. The effectiveness of this treatment against SARS-CoV-2 may vary when used to inactivate clinical samples or other types of sample matrix. Any results of inactivation testing using other sample matrices will be released as they become available.</p> <p>Inactivation reagents should not be assumed to be 100% effective against SARS-CoV-2.</p> <p>Suitability of products and treatments for inactivation of other pathogens has not been evaluated in this study.</p> <p>All COVID-19 laboratory testing workflows must be subjected to suitable and sufficient risk assessment, with consideration given to any inactivation step. Risk assessments should be reviewed regularly as new information on the inactivation of SARS-CoV-2 becomes available.</p>

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The impact of chosen inactivation method on the sensitivity of subsequent SARS-CoV-2 detection should also be assessed locally.

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Summary of revisions

Version 1: New document

Queries regarding this report or HCM inactivation testing should be directed to HCMgroup@phe.gov.uk