

SARS-CoV-2 inactivation testing: interim report

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Report date	15 July 2020		
Undertaken by High Containment Microbiology, NIS Laboratories, National Infection			
Service, Public Health England			
N.B. This is an interim report and may be updated as further results are obtained			

Product/treatment details	
Product/treatment	Polyhexamethylene biguanide (PHMB)
Manufacturer	Blueberry Therapeutics

Sample details	^o
Sample type tested	Tissue culture fluid containing 5% (v/v) foetal calf serum
Virus strain tested	SARS-CoV-2 England 2
Ratio of spiked virus stock to sample matrix	Not applicable; tissue culture fluid used undiluted

Experimental conditions			
Ratio of sample to product tested	1 volume sample to 10 volumes product		
	0.1% (0.09% final concentration)		
Product concentrations tested	1% (0.91% final concentration)		
	2% (1.8% final concentration)		
Contact time/s	30 minutes		
Temperature of incubation	Room temperature		

	Triplicate samples were treated with test buffer for indicated contact time/s or mock-treated in triplicate with an equivalent volume of PBS. All samples were then subjected to a purification step to remove cytotoxic buffer components. PBS- treated samples were subjected to the same purification procedure in parallel.
Brief description of tests performed	Test 1: Purified samples were immediately titrated on Vero E6 cells to establish virus titre. This test is quantitative and reports the titre of virus in each treatment condition in TCID50 per ml. Reduction in virus titre following treatment is given as the difference between the mean log ₁₀ TCID50/ml for
	treated conditions and the PBS control. Test 2: In parallel, purified samples were seeded onto Vero E6 monolayers to amplify any remaining virus over the course of up to four serial passages.
	Virus amplification over each passage was detected by visual (microscopic) examination of monolayers for cytopathic effect, and confirmed by SARS-CoV-2-specific real-time PCR. This test is qualitative and reports either the presence or
report	absence of virus amplification. This test may detect levels of virus that are below the detection limit of the titration assay (test 1) due to a greater sample plating volume and the opportunity for any virus present to amplify over serial passages.
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Table of results (0.1% PHMB test)				
Maximum detectable vir	6.2			
	Test 1: Virus titration post-treatment		Test 2: Passage of samples in cell culture	
	Mean virus titre (log ₁₀ TCID50/ml)	Titre reduction (log ₁₀ TCID50/ml)	Virus detected/ Virus not detected	
PBS-treated	6.9	-	Virus detected (all replicates)	
0.1% PHMB-treated	5.5	1.4	Virus detected (all replicates)	
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Table of results (1% and 2% PHMB test)				
Maximum detectable viru	5.8			
	Test 1: Virus titration post-treatment		Test 2: Passage of samples in cell culture	
	Mean virus titre (log10 TCID50/ml)	Titre reduction (log10 TCID50/ml)	Virus detected/ Virus not detected	
PBS-treated	6.5	<u> </u>	Virus detected (all replicates)	
1% PHMB-treated	5.0	1.5	Virus detected (all replicates)	
2% PHMB-treated	4.9	1.6	Virus detected (all replicates)	
Interim				

Interpretation

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Test 1: Treatment with 0.1%, 1% or 2% PHMB for 30 minutes gave a 1.4-1.6 log₁₀ reduction in infectious virus titre, with a large amount of virus detectable in all treated sample replicates.

Test 2: Infectious virus was recoverable from all treated sample replicates.

This test has been performed on tissue culture fluid containing 5% (v/v) foetal calf serum. The effectiveness of this treatment against SARS-CoV-2 may vary when used to inactivate clinical samples or other types of sample matrix. Any results of inactivation testing using other sample matrices will be released as they become available.

Inactivation reagents should not be assumed to be 100% effective against SARS-CoV-2.

Suitability of products and treatments for inactivation of other pathogens has not been evaluated in this study.

All COVID-19 laboratory testing workflows must be subjected to suitable and sufficient risk assessment, with consideration given to any inactivation step. Risk assessments should be reviewed regularly as new information on the inactivation of SARS-CoV-2 becomes available.

The impact of chosen inactivation method on the sensitivity of subsequent SARS-CoV-2 detection should also be assessed locally.

Disclaimer

PHE's evaluations of commercial products and treatments for inactivating SARS-CoV-2 have been carried out primarily for PHE's own internal use and the reports of such evaluations are shared solely for readers information; PHE does not in any way recommend any particular product for virus inactivation; and PHE shall not be responsible for the choice of product or treatment for virus inactivation, and it is the responsibility of the testing laboratory to ensure that any such product or treatment implemented has undergone the necessary verification and validation; and PHE shall not be liable, to the greatest extent possible under any applicable law, for any claim, loss or damage arising out of or connected with use of this and related reports and choice of virus inactivation products or treatments.

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Summary of revisions

Version 1: New document

Queries regarding this report or HCM inactivation testing should be directed to <u>HCMgroup@phe.gov.uk</u>

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