UNITED KINGDOM NATIONAL LIST TRIALS: TRIAL PROCEDURES FOR OFFICIAL EXAMINATION OF VALUE FOR CULTIVATION AND USE (VCU) HARVEST 2019

Spring Oilseed Rape

- this document is no longer in use, see GOV. W. for the latest proceeding

Spring Oilseed Rape VCU Procedures 2019

GROWING TRIALS, TESTS AND ASSESSMENT PROCEDURES FOR SPRING OILSEED RAPE

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SECTION A - GENERAL INFORMATION

A.1. PURPOSE

A.1.1 This document sets out the approved procedures to be used for growing trials, tests and assessments as required by the current Protocol for Official Examination of Value for Cultivation and Use for Spring Oilseed Rape.

A.3.1 Procedures Development Group The Procedures Development Group is responsibile for reviewing these procedures around and making amendments for which it has responsibility, in accordance with the provisions of the VOP Protocol. A.3.2 Organisers and Operators A.3.21 Trials Organiser British Society of Plant Breeders Ltd (BSPB) BSPB House 114 Lancaster Way Business Park Ely Tel No: 01352 Cambs. Fax M CB 3NX 3.2.2 T

0 The Trials Organiser is responsible for organized and Procedures requirements A.3.2.2 are followed and liaison with all Operators carrying out trials for National List purposes, including supply of seed and data handling.

A.3.2.3 Data Handling Operator

The Data Handling Operator identified by the Trials Organiser is responsible for trial design and data validation in accordance with the **VEU Protocol** and associated **Procedures**.

Growing Trial Operators, Seed Handling Operators and Quality Testing Operators. A.3.2.4

The Trials Organiser, responsible for identifying potential Growing Trial Operators and Quality Testing Operators to carry out trials and tests as determined by the Procedures Development annual review in accordance with the VCU Protocol, and these Procedures.

of all approved Organisers and Operators is shown in Appendix 1. A.3.2.5

CU Protocol and Procedures non-compliance

Where these procedures use the words "must or will" for any action then failure to carry out this action will result in non-compliance. Where the word "should" is used for any action then this is the method to be followed unless there are clear reasons not to, which can be justified by the operator as technically sound.

A.3.3.2 The Trials Organiser will forward any reports on VCU Protocol or Procedures non-compliance to APHA within 1 week of receipt. The Trials Organiser will obtain authorisation from APHA for any actions, including those necessary to remedy non-compliances, which are not within the requirements of the VCU Protocol. Such actions must be recorded as non-compliance. Where emergency action is required and APHA staff are not available (eg evenings/weekends) the Trials Organiser should act but report this to APHA at the earliest opportunity. Where GMOs are concerned the arrangements are as detailed in section 3.4.

A.3.4 **Procedures for GM varieties**

A.3.4.1 The National Authorities and Trials Organiser will develop procedures for GM varieties if an application for a GM candidate variety is received.

A.3.5 **Processing of Seed**

A.3.5.1 The Seed Handling Operator is responsible for organising the processing of seed of candidate varieties submitted by the applicant, and seed of control, or other reference varieties, in accordance with the requirements set out in these **Procedures** and the current **VCU Protocol**. The Trials Organiser with ensure that any seed treatments or additives are approved for the purpose. Approved products are listed by <u>Appendix</u> <u>2</u>.

A.3.6 Dispatch of Seed

A.3.6.1 The Trials Organiser will arrange for seed to arrive at the Seed Handling Operator by the relevant deadline - see <u>Appendix 3</u>. The Seed Handling Operator is responsible for processing and dispatch of seed to Growing Trial Operators and DUS testing centres (including, where appropriate, foreign testing authorities) within the relevant deadlines.

A.3.7 Monitoring of Growing Trial Operators

A.3.7.1 The Trials Organiser will take any necessary action to prove deadline dates and quality standards for required documentation.

A.3.7.2 The Trials Organiser will ensure Growing Trial Operators and Seed Handling Operators have access to all current protocols and procedures relevance them and that they are notified of any amendments.

A.3.8 Seed Quantities

A.3.8.1 The Trials Organiser will determine the quantity of seed required for all VCU tests and trials in each annual series, including authentication, and will notify the applicant of quantities and delivery addresses.

A.3 9 Labelling of seed

A.3.9.1 The Seed Handling operator is responsible for ensuring all seed is clearly labelled with variety name/breeders reference and AFP number.

A.3.10 Seed Quality

A.3.10.1 Seed submitted for VCU testing must meet the standards for the final generation of seed given in the appropriate seed regulations, in respect of germination, analytical purity and content of other seeds and any other inspirities. The seed must be free of adventitious genetically modified presence and accompanied by a declaration to this effect.

SUMMARY OF GROWING TRIALS, TESTS AND ASSESSMENT PROCEDURES A.4.

A.4.1 The number of trials and site locations are as detailed in Appendix 4.

A.4.2 Control varieties are listed in Appendix 5.

procedure A.4.3 The Trials Organiser is responsible for informing the Growing Trial Operators of any additional characters, which must be recorded as and when requested by applicants, and any samples that may be required for analysis. Any sensitivity to herbicides to be reported to the Trials Organiser.

A.4.4 VCU trial assessments required

Bold = Obligatory Italics = Additional only if requested by the applicant

Resistance to harmful organisms Section D Presence and severity of Light Leaf Spot, powdery milder and Stem Canker recorded the field. Other diseases should be recorded if they the infection levels specified in Section D. Other diseases should be recorded if they the infection levels specified in Section D. Quality characteristics (Laboratory Tests) Section E Glucosinolate content measured on each oil content JRTHER MEASUREMENTS Section E Section E	Moisture content Behaviour with respect to factors in the physical environment. Section C Maturity Standing ability Early vigour Plant height Earliness of flowering Resistance to harmful organisms Section D Presence and severity of Light Leaf Spot, powdery milder, and Stem Canker recorde the field. Quality characteristics (Laboratory Tests) Section E Glucosinolate content measured on each oil content JRTHER MEASUREMENTS he following must be measured or recorded in all trials, following procedures in Section C. Section C	Type of Character	Reference	Description of assessment
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Plot size Bird Damage Seed Loss	Combine losses 5			

SECTION B – SEED HANDLING PROCEDURES

B.1. RESPONSIBILITIES

B.1.1 The Seed Handling Operator is responsible for carrying out the following seed handling procedures.

B.2. SEED HANDLING PROCEDURES

B.2.1 The Seed Handling Operator will receive a sowing list from APHA. A list of chemicals approved by the Procedures Development Group is at <u>Appendix 2</u>.

B.2.2 The Seed Handling Operator must record receipt of seed from applicants by checking it off spainst the sowing list as it arrives. APHA should be notified of any damage to the packaging, loss of seed or identification problems within one working day of receipt.

B.2.3 Once seed has been treated, it must be kept safely until required for drilling, and entication and quality control. The Seed Handling Operator must retain 50 grams untreated sample of the seed of every variety in the trial for authentication by the DUS Test Centre.

B.2.4 The Seed Handling Operator must record use of treatment chemicals in accordance with best practice and in full observance of all manufacturers' recommendations and recevant statutory obligations.

B.2.5 Any seed treatment equipment used must be fit for the purpose, properly calibrated, set up and operated in accordance with the manufacturer's recommendation.

B.2.6 Cross contamination must be avoided by ensuring **comment** is cleaned between weighing and treatments.

B.2.7 A record must be kept of chemicals used and date of treatment.

B.2.8 Seed treatment should take place as near othe drilling date as possible.

B.2.9 Once seed has been treated, it must kept safely until required for drilling and quality control. The Seed Handling Operator must retain a 50 grant sample of treated seed until one month after harvest.

B.3. AUTHENTICATION OF VCN SEED

B.3.1 The Seed Handling Operator must forward 50 grams of untreated samples of the VCU seed submitted of every variety in the treal, for authentication by the DUS test centre according to procedures laid down and notified by APHAO

B.3.2 All samples **must** be kept under suitable conditions for the authentication procedures required and must be clearly labelled and sealed.

B.3.3 If the revel of off-types recorded in DUS tests or VCU authentication of a candidate variety exceeds 10%, the VCP data will be considered invalid.

SECTION C – GROWING TRIAL PROCEDURES

C.1. RESPONSIBILITIES

C.1.1 The Growing Trial Operators are responsible for conducting the trials according to these procedures.

C.2.

The Growing Trial Operator will be responsible for providing a suitable site, which meets the criteria. Previous cropping must be appropriate for a spring oilseed rape crop to be grown. C.2.1 following criteria.

C.2.2

C.2.3 Soil type should be typical of those on which spring oilseed rape is grown locally. texture should be uniform across the site. The soil should be sufficiently uniform with no submantial variations in previous cropping, ridges and furrows, etc.

The trial should be sited away from trees, hedges, headlands and other watures, which are likely C.2.4 to cause uneven growth or encourage grazing damage from birds, rabbits, hares mice etc.

Primary cultivation at the discretion of the growing trial operator and cultivations should follow best C.2.5 local practice.

C.3 SOWING THE TRIAL

C.3.1 Plot size

C.3.1.1 The harvested plot area per variety must be not less than 20 m² per replicate and four replicates must be used. Plots should be drilled to a greater legan than required and cut back to the required length prior to harvest. The plot width for calculating harvested area is measured centre gap to centre gap with an inter-plot gap in the range 0.5m to 0.8m. Sown plot width should reflect the blade width of the swathers where used.

C.3.2 Plant population

Seed rates may be adjusted to suit site conditions at the discretion of the trials operator with the C.3.2.1 f between 80 and 100 plants/m². Hybrids should be drilled at 80% seed aim of producing a plant populatie rate.

The following formula w used to calculate the seed rate for a given thousand seed weight:-

Parget population x Thousand seed weight) x 100) Seed rate (kg/ha) (Establishment% x Germination %)

using seed counters the following formula can be used to calculate required seed numbers For operators

er plot = ((Target population x Drilled plot area) x 10,000) (Establishment% x Germination %)

Establishment % can vary greatly between locations and drilling techniques and figures as low as 60% are not uncommon. A good assessment of this figure is important in establishing successful trials.

C.3.3 Trial layout

C.3.3.1 The Trials Organiser, following consultation with APHA, produces provisional sowing lists. The Trials Organiser will make final sowing lists available to Growing Trial Operators, along with the trial plans produced by the Data Handling Operator. There should be no correction for differences in germination percentage.

C.3.3.2 The trial should be sown according to the plan produced by the Data Handling Operator and may be an incomplete block design. In an incomplete block design each replicate is split into a number of subblocks. Any splitting of replicates must be between sub-blocks and not through sub-blocks. Varieties can be moved within a sub-block but must not be moved from their sub-block. Varieties must not be moved around within the plan e.g. if drilling errors occur. If plots are moved out of their original sub-block they will be to be treated as missing plots. If there are any queries please contact the Trials Organiser.

C.3.3.4 If there is a need to replace a planned variety e.g. if varieties are withdrawn, affected bots must be sown with any of the standard control varieties. Any such replacements must be agreed with the Trials Organiser. The control varieties are listed in <u>Appendix 5.</u>

C.3.4 Drilling

C.3.4.1 Trials should be drilled when soil and weather conditions are conducive to rapid establishment. Where possible, time the drilling of the trial to coincide with that of the surrounding term crop.

C.3.4.2 Care must be taken with drill settings and drilling speed to ensure satisfactory and uniform establishment and plant population from plot to plot. It is also important to ensure that there is no carry over of seed between plots.

C.3.4.3 At least **one** discard plot must be drilled on either side of the trial with the same drill and at the same time that the trial is drilled.

C.3.4.4 Precautions must be taken to avoid any missing rows. Any missing rows or parts of rows must be noted in the trial diary and reported to the Trials Organiser within one month of emergence.

C.3.4.5 Flea Beetle post emergence control should be applied.

C.3.5 Confirmation of trial layout

C.3.5.1 After full establishment and within one month of sowing the Growing Trial Operator must confirm that the trial has been sown to plan or give full details of any changes to plan. This should be done by clearly highlighting the changes in the electronic plan and returning it to the Data Handling Operator.

- Return a completed site data 1 sheet including the following information:
- Site location details including how to get to the field.
- Sketch showing the layout of the trial in the field, in relation to other trials and showing access road, gates, etc.
- Tria Sketch showing plot numbers and variety codes and/or names
 - A short post-establishment report of the condition of the trial.

HUSBANDRY

Agronomy

Where not specified in these procedures agronomy should follow best local practice, advisory and regulatory guidelines. Application of fertilisers and sprays should be uniform. It is normally best to apply these across the direction of the plots. Application wheelings should not run through the harvested plot area.

C.4.2 Fertiliser application

It should take into account inherent fertility, previous cropping, winter rainfall, the best local practice. All fertiliser applications should take account of the AHDB Nutrient Management Guide (RB209), the corresponding advisory publications in England, Wales, Scotland and Northern Ireland and past trialling experience.

the latest procedure Growing Trial Operators should be aware of the implications of other nutrient requirements (especially Sulphur) and should be prepared, if necessary, to apply appropriate treatments.

C.4.3 **Herbicides**

The Trials Organiser must be consulted. .

C.4.4 Growth Regulators

These should not be used on spring oilseed rape trials.

C.4.5 Pest and Disease Control

C.4.5.1 Pest Control

Seed dressings may include an insecticide element. Precautions should aken against attacks by slugs and insects such as cabbage stem flea beetle, cabbage root fly, pollen beetle, seed weevil and pod midge. Where there is a risk of significant flea beetle or pollen beetle attack proving Trial Operators must ensure that adequate control measures are taken. Birds can cause damage near harvest, especially when trials are near houses. Control is difficult but every effort should be made to minimise losses. Assessments should be made wherever damage occurs since decisions have to hade on the validity of each plot affected. Grazing, particularly by pigeons, may be selective and contro measures should be taken if necessary.

C.4.5.2 **Disease control**



Growing Trials Operators should be aware that severe outbreaks of Sclerotinia, powdery mildew and Alternaria could threaten the validity of the trial and should weather patterns favour the build up of these diseases, then an appropriate fungicide should be applied at mid-flower for Sclerotinia, or from mid-flower to pod senescence for Alternaria. Though the risks of Sclerotinia and Alternaria development are generally lower in SOSR than WOSR, damaging attacks could occur. Sclerotinia may develop if a flush of apothecia production coincides with flowering and periods of wet weather; Alternaria may develop rapidly if warm and wet conditions occur during late flowering and pod development. If control measures were ineffective for any reason, and these diseases difference on the period be recorded according to the instructions in <u>Appendix</u> 8. Other disease control sold only be undertaken after agreement by the Trials Organiser.

C.4.6 Irrigation

Irrigation will not be permitted without the specific agreement of the Trials Organiser.

C.4.7 Navs

SC.5.1

angways should be made after the risk of pigeon damage has passed.

HARVESTING

Timing of harvesting

Date of harvesting will be determined by the Growing Trial Operator based on crop maturity and C.5.1.1 local weather conditions.

C.5.1.2 Plots should be trimmed to their final length prior to harvesting. The plot dimensions must be measured prior to harvesting. If it is necessary to reduce the size of any plot at harvest give clear details on the yield file. Individual harvested plot lengths should be recorded.

C.5.2 Harvesting method:

Trials can be swathed or desiccated and direct combined, depending on the specific requirements of the trials as determined by the Growing Trials Operator. This should be recorded on the site information form. Side knives must not be used.

If the trial is to be cut direct, a header-extension should be fitted to the plot combine to minimise table losses -jedure

C.5.3 Samples

C.5.3.1 It is essential that all samples:

- Are representative of the variety/plot from which they are taken with minimal contamination sampling on-combine, it is essential to minimise the risk of contamination of set from the previous plot.
- Are taken from the same source.
- Contain the weight of seed requested.

C.5.3.2 Two samples must be taken from each plot at harvest. A 200 g sample Gust always be taken at the time of plot weighing and sealed in a polythene bag for dry matter and other other determination. In addition a 100 g sample is taken and sealed in a cloth bag for glucosinolate analysis. One label should be placed inside the bag and this sealed by rolling over the top and securing the bag and the second label with rubber bands. At sites where higher moisture levels are frequently experienced and dry matters are determined immediately in the trial operator's laboratory a single sample of 500g per plot and subsequently divided may be taken for dry matter, oil and glucosinolate content

All bagged samples must be kept in good condition at a moisture content and temperature C.5.3.3 appropriate for long term storage. They should be clearly waked both inside and outside the container/bag.

Samples may not be required from every variety - the Trials Organiser will provide details of which C.5.3.4 varieties require samples, the quantities required anothe tests to be carried out.

Sample drying should be undertaked using a cold/warm air drier to reduce moisture content to 9% C.5.3.5 or below according to the procedures in E

All plot samples must be labeled with trial identification number, variety name/breeders reference, C.5.3.6 AFP number, plot number and Growing Trial Operator identification number.

C.5.4 Submission of data and samples

Appendix 6 lists the records, with deadlines, to be sent to the Trials Organiser. Diary sheets and C.5.4.1 any other field recorde should be returned to the Trials Organiser within 5 working days of harvest.

C.5.4.2 All plot records should be transmitted to the Data Handling Operator following the deadlines set out in Appendix 6 The Growing Trial Operator should ensure that data are free from errors before transmission. After scruting copies of results will be returned to the Growing Trial Operator for action as agreed by the Trials Organiser.

All samples should be sent to the Quality Testing Operator following the deadlines set out in A spendix 6.

C.6 RECORDS

- C.6.1 There are four components:
 - Field notes of trial status. 1. Diarv
 - 2.* Site data part 1 Site details including site sketch, map and location, previous cropping, soil
 - 3.
 - 4
 - * Template available from Trials Organiser

C.6.1.1 variety performance should be recorded. If the trial is in good condition, with no problems, this should be recorded.

C.6.2

An entry in the Diary sheet should be made on every trials visit and any observations relevant to erformance should be recorded. If the trial is in good condition, with no protections, this should be **Plot records** C.6.2.1 Plot data may be recorded direct onto a data logger using stem approved by the Trials Organiser or recorded on paper then entered and validated onto a computer using an approved system. A system of ensuring that data are recoverable, in the event of loss of original data, must be implemented, e.g. copy and safe storage. Whichever method is used, individual pot data will only be accepted by the Data Handling Operator in an approved format using the variate names and units as listed in Sections C and D.

All observations should be checked at the time or the time of the C.6.2.2 limits for the character recorded. Observations that have been identified as exceptional by the recorder should be identified with a note on the approved data from hard copy medium describing the possible causes together with a recommendation for their exclusion in the trial analysis.

Plot numbers on record sheets must birrespond with the numbering on the field plan. C.6.2.3

C.6.2.4 If a character is not recorded or is missing the Growing Trial Operator should indicate in the diary or on the recording sheet the reason why it has been excluded.

Where a plot record is must be Growing Trial Operator should enter "*" in the approved data file C.6.2.5 or hard copy medium and, units the non-recording of the plot has already been agreed with the Trials Organiser, append a note Othe file explaining why a missing value has been entered for that plot. The Growing Trial Operator should not enter "0" for missing plots.

All records should be returned as soon as reasonably possible. Indicative deadlines are given in C.6.2.6 All records must be returned by the final deadlines. Appendix 6. this documer

C.6.3 **Procedures for recording Characters**

The following procedures must be followed for measuring all characters to be used in NL decision-making.

C.6.3.1 PLOT YIELD AND MOISTURE CONTENT

The following information must accompany the yield data:

The moisture content % of the harvested grain determined by oven method.

Plot width: the width of the harvested plot in metres. Plot width: the width of the harvested plot in metres from outer row to outer row plus half of the inter-on either side. The adjustment for the inter-plot gap should be no greater than 0.8m. If these are not the same for every plot, a separate record must be submitted

Growth stage: usually 9.9 at harvest. The Growth Stage Chart for oilseed rape is at A

Yield (in kilograms). Note clearly any tare weight to be subtracted. Yield, moisture content, plot length, plot width and harvest date should be sent to the data Handling Operator within 5 days of harvesting the trial.

C.6.3.2 MATURITY

from all plots

Maturity should be judged with a visual estimate of canopy senes

- all pods green 1
- all pods bleached and brittle 9

Unrepresentative areas of the plot should be avoided when making assessments, for example, localised diseased infections.

C.6.3.3 EARLY VIGOUR

- 1 very weak
- very vigorous 9

Record also, the weediness and predominant weeds present at the time of assessment.

C.6.3.4 PLANT HEIGH from all plots

(ADDITIONAL) (cm)

(1-9)

(ADDITIONAL)

(OBLIGATORY) (kg)

Record average plot height at the end of flowering before leaning or lodging takes place. If lodging has sentative area of the plot, lift a number of plants against the measuring pole and occurred, choose a rep record an average height.

ESS OF FLOWERING from all plots (ADDITIONAL) C.6.3.5 (1-9)1

ord when the earliest variety is in full flower and score all varieties relative to this. An assessment on one ecasion is normally sufficient. Estimate the date of full flowering for the earliest control variety.

C.6.3.6 SOWING DATE of each trial

(OBLIGATORY) (Day/month/year)

This is recorded in Part 1 of the Site Information Form.

C.6.3.7 STEM STIFFNESS

from all plots

SECTION C (OBLIGATORY) (1-9)

(Day/month/year)

(OBLIGATORY)

- 1 completely lodged
- 9 no lodging

The aim of this score is to describe the canopy structure at harvest. A score of 5 can describe half the plot completely flat or the whole plot leaning at 45 degrees.

C.6.3.8 HARVEST DATE

to minimise further

This must be recorded.

Base scores either on observation of pod shattering or courts of seed on the ground if shedding is thought to be serious. Seed loss is easier to assess before combining. Ensure that combines are set correctly to minimise losses at harvest. Estimate the number of seeds lost per m² for the plot(s) with the most losses so that the approximate yield loss can be estimated.

tom all plots

C.6.3.11 COMBINE LOSSES

(OBLIGATORY) (1-9)

This must be recorded.

Combine losses should be assessed if the losses are thought sufficient to exclude the yield data from results. ains lost per m² for the lowest score given on the 1 to 9 scale. Indicate the estimated number

9 = no combine losses.

C.6.3.12 SITE FAC

may have affected the yield of the trial or individual plots must be noted and accompany Any factors which the yield data

Where varietal differences are seen in pest or disease attack, records should be made in accordance with the procedure in Section D for disease.

drds for other scores should be taken as plants affected on a 1 to 9 scale. Include definitions for each rating on the 1 to 9 scales.

- To supply the inspector with information (for example pesticides applied etc) within seven days of a request.
- To co-operate with the inspector in making any non-routine assessments required to establishe
- <text><text><text><text><text>

SECTION D - DISEASE TESTING PROCEDURES

D.1 ASSESSMENT OF NATURAL INFECTION

D.1.1 The Growing Trial Operator is responsible for carrying out these procedures.

D.1.2 **Disease Observation Plots**

No disease observation plots are grown routinely.

D.1.3 Naturally occurring disease in VCU growing trials

redure Light leaf spot, powdery mildew and stem canker should be recorded when the level of inter-D.1.3.1 the most affected variety is over 5% or a score of 4 of the leaf or infected plants.

D.1.3.2 Other naturally occurring disease is not normally recorded in the growing trials. However, if disease levels increase to more than 5%/score 4 of the leaf area (or 5%/score 4 of infected plans, as appropriate for the diseases) on the most affected variety a score should be made on the whole trial and sent to the Data Handling Operator. Confirmation of the identity of a disease should be obtained tom an appropriate plant pathologist if required.

D.1.3.3 **Recording methods**

All usease records to be sent to the Data D.1.3.4 Appropriate assessment keys are given in Appendix 8. INOCULATED DISEASE TESTS No inoculated disease tests are carried out routine

routine this document is no longer in use

SECTION E - QUALITY TESTING PROCEDURES

E.1. RESPONSIBILITIES

E.1.1 The Quality Testing Operator appointed by the Trials Organiser is responsible for conducting approved quality tests according to these procedures.

E.2 QUALITY ASSESSMENT METHODOLOGY

E.2.1 Moisture content determination

The following procedure must be followed;

rocedure A 105 g sample of seed (± 5g) is placed in the drier which must be at a temperature of 100°C \pm air recirculator set in the range 80-100% recirculation in order to restore the temperature to C ± 4°C as rapidly as possible. When the temperature is restored to 100°C ± 4°C the air regulator is set at 80% recirculation i.e. 20% fresh hot air. The regulator is critical for rapid drying. The samples are dried at 100°C ± 4°C for such time as is necessary for complete drying. Each sample is identified with a label.

The dried sample is carefully removed from the drier as soon as the sample scool enough for accurate weighing. The dry weight is recorded to one decimal place.

When all samples from a given trial have been recorded, the fresh and dry weights are immediately reported to the Trials Organiser. When the moisture contents are reported as a percentage, the fresh weight should be reported as 100.

Moisture content determination by conductance moisture meter will not be acceptable.

E.2.2 **Glucosinolate Determination**

E.2.2.1 The standard method is based on X-ray flue scence spectroscopy. The standard procedure is given in modification of ISO 9167-2: 1994 (the modification being that the instrument is now a more up to date version which has silicon drift detectors (200)).

Because of the variable moisture content of trials material the following modification to ISO 9167-E2.2.2 2:1994 is permitted:

All analyses may be carried out material which has been oven dried at $100^{\circ}C + 4^{\circ}C$ for a minimum of 5 hrs. If this option is chosen the instrument must be calibrated against whole rapeseed reference materials which have been similarly dried. The sulphur reference values used for calibration should be the values for the un-dried reference material, this ensures the instrument gives analytical results at the correct moisture basis despite the drying step. A single determination is normally performed.

An alternative analytical procedure based on High Performance Liquid Chromatography (HPLC) is E.2.2.3 also permitted The standard procedure is given in ISO 10633-1:1995. HPLC is used when there is reason to believe somples contain exogenous sulphur - for example seed which is chemically dressed. The method is standardised daily (when in use) against reference samples to ensure the activity of the sulphatase preparation, the suitability of the sinigrin standard and the reproducibility of the laboratory procedures. Three extracts are prepared from a test sample, one without internal standard and two with added internal standard.

Oil Content Determination

Analysis is performed using continuous emission NMR following ISO 5511:1992. Results are expressed as apparent oil as a percentage at 9% moisture.

The stability of the equipment is checked at two-hourly intervals through the working day by the use of weighed oil standards. A single determination is normally performed on each test sample.

 $\mathbf{P}_{2.3}$

SECTION F - TRIAL DESIGN AND DATA HANDLING PROCEDURES

F.1. PLAN VALIDATION AND STORAGE

- F.1.1 After the trial has been drilled, the Growing Trial Operator must:
 - a) Confirm that the trial has been drilled according to plan and provide the sowing date, by returning site data 1 and associated trial sketch to the appropriate Data Handling Operator.
 - b) If any amendments to the plan have been made, return a hard copy of the plan to the appropriate Data Handling Operator with any amendments clearly indicated. Alternatively, amendments be notified electronically with the agreement of the Data Handling Operator.

F.1.2 The Data Handling Operator will check these for statistical validity and, once this has been done, will load the plan on the database.

F.2 DATA RECORDING

F.2.1 Data are recorded using the methods and characters given in Sections 20 and E.

F.2.2. Site information is recorded for each trial including, for example, data on previous cropping, seed rates, soil details and fertiliser applications.

F.2.3 Details of any agrochemical applications are also recorded and retained by the Growing Trial Operator.

F.3 OTHER TESTS AND TRIALS

F.3.1 Any additional or alternative designs required for the assessment of additional VCU characters not detailed in <u>Appendix 3</u> of the VCU TRIAL PROTOCOL for spring oilseed rape will be added to these **Procedures** as and when approved by the NLSC.

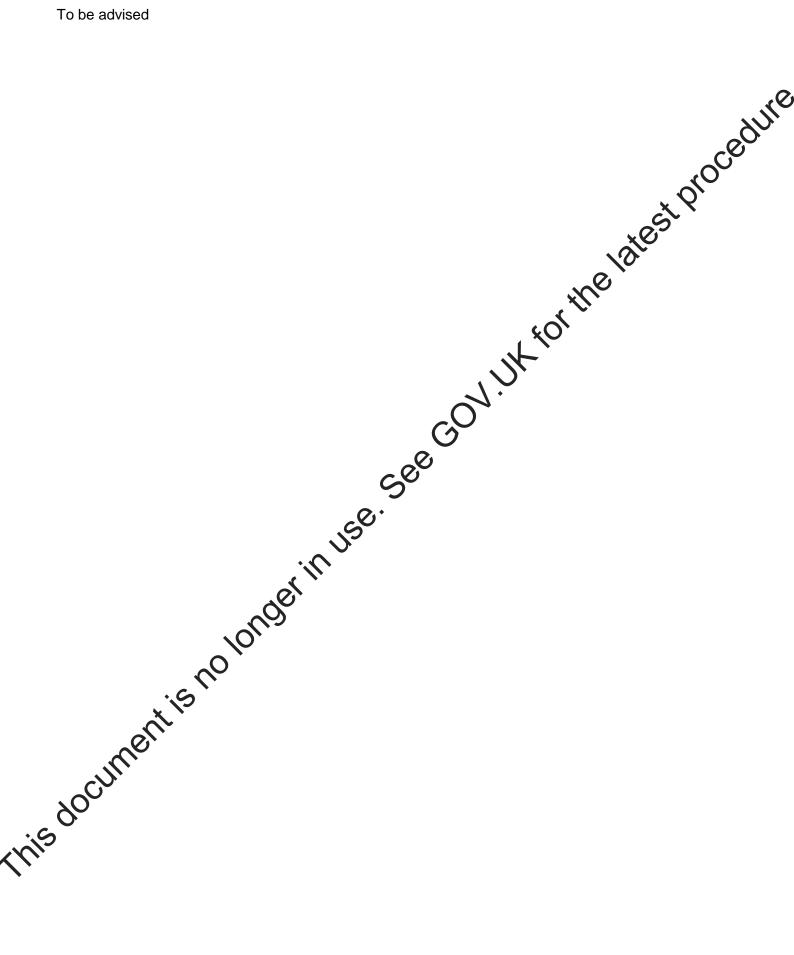
This document is no longer in use

APPROVED TRIAL ORGANISERS/OPERATORS FOR SPRING OILSEED RAPE

Trials Organiser	ORGANISERS/OPERATORS RESPONSIBLE
	BSPB
Seed Handling Operator	NIAB
Data Handling Operator	AHDB Cereals and Oilseeds
Pathology Trials Operator	None
rial inspection	AHDB Cereals and Oilseeds
echnical Validation Operator	AHDB Cereals and Oilseeds
Quality Testing Operator	NIAB
Data Review and Standards Setting Operator	NIAB
erinuse	AHDB Cereals and Oilseeds NIAB NIAB NIAB See

SEED TREATMENT PRODUCTS FOR USE ON NL TRIALS

To be advised



SEED DESPATCH DEADLINE DATES

VCU seed must be delivered to NIAB Seed Handling Unit by 15th January.

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GROWING TRIAL OPERATORS AND TRIAL LOCATIONS

1. Growing Trial Operators/Seed Handling Operators

Trial Operator Seed Handling Operator Location of Trial (If not trial operator) Shropshire Shropshire Plant Breeding NIAB Shropshire Plant Breeding NIAB Cambridgeshire pottish Agronomy NIAB Perthshire Pathology Trials Operator Location of Trial Operator thology Trials Operator Location of Trial Operator the applicable - - Operator connection - - - - Schoperator - - - - the applicable - - - - connection - - - - connection -	Growing Trial Operator	Seed Handling Operator	Location of Trial	
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CONTROL VARIETIES FOR VCU ASSESSMENTS

this document is no longer in use, see GOV. Whore the latest procedure The Control Varieties:

A. DATES BY WHICH RECORDS SHOULD BE SENT TO TRIALS ORGANISER

Record	Latest date of receipt by Trials Organiser
Site data part 1 (including site sketch)	Within 1 month of drilling trial
Site data part 2	By the time trials harvested
Plot records (in approved electronic format)	Growing Trial Operator should notify Trials Organiser that trial has been harvested within 2 days of harvest

B. DATES FOR SUBMISSION OF PLOT RECORDS TO DATA HANDLING OPERATOR

B. DATES FOR SUBMISSION OF PLOT RECORD	C	<u>ç</u>
Record	Date 🗙 🗙	
Plot records should be sent to the Data	Yield and moisture data within 3 days harvest	
Handling Operator	other data within 10 days of record Using taken	

C. DATES FOR SUBMISSION OF PLOT SAMPLES TO QUALITY TESTING OPERATOR

arvest

GROWTH STAGES OF OILSEED RAPE

• • • • • •	GrowthStage	
Germination and Emergence	0.0	Dry seed
Leaf Production	1.0	Both cotyledons unfolded and green
	1.1	First true leaf emerged
	1.2	Second true leaf emerged
	1.3 etc	Third true leaf emerged
Stem Extension	2.0	First true leaf emerged Second true leaf emerged Third true leaf emerged No internodes (rosette)
	2.5	About five internodes
		0/
Flowerbud Development	3.0	Only leaf buds present
	3.1	Flower buds present but enclosed by leaves
	3.3	Flower buds visible from above ('greed bud')
	3.5	Flower buds raised above leaves
	3.6	First flower stalks extending
	3.7	First flower buds yellow ('yellow bud')
Flowering	4.0	First flower opened
	4.1	10% all buds opened
	4.3	30% all buds opened
	4.5	50% all buds opened
Pod Development	5.3	30% potential pods
	5.5	50% potential pods
	5.7	70% potential pods
	5.9	Appotential pods
		D ⁻
Seed Development	6.1	Seeds expanding
	6.2	Most seeds translucent but full size
	6.2 6.3	Most seed green
	• • • • • • • • • • • • • • • • • • •	Most seed green-brown mottled
	6.5	Most seeds brown
	6.6	Most seed dark brown
0	6.7	Most seed black but soft
	6.8	Most seed black and hard
long	6.9	All seeds black and hard
0		
Leaf Senescence	7.0	
Stem Senescence	0.4	Most store gross
Stem Senescence	8.1	Most stem green
	8.5	Half stem green
	8.9	Little stem green
Porsenescence	0.1	Moot pada graap
ron senescence	9.1	Most pods green
	9.5	Half pods green
V	9.9	Few pods green

ASSESSMENT OF OILSEED RAPE DISEASES

Use for assessing light leaf spot, Alternaria, downy mildew, powdery mildew, Phoma and white leaf spot on leaves and pods:

- 1) Examine all leaves and pods in 3 areas of each plot.
- 2) Ignore all naturally senescent tissue.
- 3) Include all necrosis and chlorosis attributable to disease.
- 4) Estimate % infection using the descriptions below. Record the average % infection from the 3 areas. Interpolate values if necessary. Disease may be recorded on a 1-9 scale but the data must be submitted as a percentage score. Both scales are given in the assessment key.

1-9 score	%	LEAVES	PODS	
	Infection		, Q.	
1	0	No infection observable		
2	0.1	Trace of infection		
3	1	Diseased leaves with 1 small lesion;	Terminal raceme with the scattered	
		plants with a few scattered lesions	lesions	
4		Leaves appear 1/10 infected;	Terminal raceme appears 1/10 infected;	
		diseased leaves with 2 lesions	diseased pods with 1 or 2 lesions	
5	10	Leaves appear ¼ infected; diseased	Terminal receme appears 1/4 infected;	
		leaves with few large or many small	diseased bods with 2 or more lesions	
		lesions		
6	25	Area appears 1/2 infected 1/2 green		
7	50	Area appears more infected than green		
8	75	Very little green tissue left		
9	100	Leaves/pods cead - no green tissue left		

These descriptions are guides for specific levels; interprete between these points as necessary e.g.15%, 27%, 60% etc.

Other disease assessments:

Club root

Any suspected club root in trials should be confirmed by sampling between 10 and 30 plants within the suspected area, and its presence notified to the co-ordinators.

Sclerotinia %

Should be assessed as the stems with complete girdling leading to 'whiteheads' within a plot.

Botrytis%

Should be assessed as the % of stems infected within a plot.

Stem canker

Stem canker hay be assessed by pulling up 30 stems per plot before harvest. Stems should be pulled at random throughout the plot, but since access is likely to be very difficult, aim to take 15 stems from the second drill row on each side of the plot, using the first 3-5m of the plot length. Appropriate sampling times are usually from the middle of June onwards. If sampling is not carried out prior to swathing, it must be done **as possible afterwards, within a maximum of 2 days.**