UK Standards for Microbiology Investigations

Investigation of swabs from skin and superficial soft tissue infections
Acknowledgments

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The contributions of many individuals in clinical, specialist and reference laboratories who have provided information and comments during the development of this document are acknowledged. We are grateful to the medical editors for editing the medical content.

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Logos correct at time of publishing.
### Amendment table

Each SMI method has an individual record of amendments. The current amendments are listed on this page. The amendment history is available from standards@phe.gov.uk.

New or revised documents should be controlled within the laboratory in accordance with the local quality management system.

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<td><em>Haemophilus influenzae</em> was removed.</td>
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### Investigation of swabs from skin and superficial soft tissue infections

4.4.1 | Section regarding Gram stain has been clarified.

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<td>Title updated to indicate sample type.  References reviewed and updated throughout. Hyperlinks updated to gov.uk.</td>
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<td>Scope.</td>
<td>Inclusion of swabs of pus. Inclusion of links to relevant SMIs.</td>
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<td>Introduction.</td>
<td>Original text reorganised and streamlined. Additional text included from B14 – Investigation of pus and exudates and B17 – Investigation of tissues and biopsies from deep-seated sites and organs following reorganisation of these documents.</td>
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<td>Technical information/limitations.</td>
<td>Section of rapid methods included.</td>
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| Specimen processing/procedure. | 4.5.1 Culture media and organisms Specimen type added to table. All conditions – addition of Staph/Strep selective agar as an alternative to blood agar. Addition of CLED/MacConkey agar. Addition of swab of pus to supplementary media section. Removal of reference to swabs from dirty sites. Sabouraud agar incubation amended to 28-30°C for 14d. 4.6.1 Minimum level of identification Aeromonas, dermatophytes and mould added to the table. Additional information included in right hand column regarding exceptions and information for |

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specific situations.
Information regarding *C. diphtheria* included.

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UK SMI#: scope and purpose

Users of SMIs

Primarily, SMIs are intended as a general resource for practising professionals operating in the field of laboratory medicine and infection specialties in the UK. SMIs also provide clinicians with information about the available test repertoire and the standard of laboratory services they should expect for the investigation of infection in their patients, as well as providing information that aids the electronic ordering of appropriate tests. The documents also provide commissioners of healthcare services with the appropriateness and standard of microbiology investigations they should be seeking as part of the clinical and public health care package for their population.

Background to SMIs

SMIs comprise a collection of recommended algorithms and procedures covering all stages of the investigative process in microbiology from the pre-analytical (clinical syndrome) stage to the analytical (laboratory testing) and post analytical (result interpretation and reporting) stages. Syndromic algorithms are supported by more detailed documents containing advice on the investigation of specific diseases and infections. Guidance notes cover the clinical background, differential diagnosis, and appropriate investigation of particular clinical conditions. Quality guidance notes describe laboratory processes which underpin quality, for example assay validation.

Standardisation of the diagnostic process through the application of SMIs helps to assure the equivalence of investigation strategies in different laboratories across the UK and is essential for public health surveillance, research and development activities.

Equal partnership working

SMIs are developed in equal partnership with PHE, NHS, Royal College of Pathologists and professional societies. The list of participating societies may be found at [https://www.gov.uk/uk-standards-for-microbiology-investigations-smi-quality-and-consistency-in-clinical-laboratories](https://www.gov.uk/uk-standards-for-microbiology-investigations-smi-quality-and-consistency-in-clinical-laboratories). Inclusion of a logo in an SMI indicates participation of the society in equal partnership and support for the objectives and process of preparing SMIs. Nominees of professional societies are members of the Steering Committee and Working Groups which develop SMIs. The views of nominees cannot be rigorously representative of the members of their nominating organisations nor the corporate views of their organisations. Nominees act as a conduit for two way reporting and dialogue. Representative views are sought through the consultation process. SMIs are developed, reviewed and updated through a wide consultation process.

Quality assurance

NICE has accredited the process used by the SMI Working Groups to produce SMIs. The accreditation is applicable to all guidance produced since October 2009. The process for the development of SMIs is certified to ISO 9001:2008. SMIs represent a good standard of practice to which all clinical and public health microbiology

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Microbiology is used as a generic term to include the two GMC-recognised specialties of Medical Microbiology (which includes Bacteriology, Mycology and Parasitology) and Medical Virology.

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laboratories in the UK are expected to work. SMIs are NICE accredited and represent neither minimum standards of practice nor the highest level of complex laboratory investigation possible. In using SMIs, laboratories should take account of local requirements and undertake additional investigations where appropriate. SMIs help laboratories to meet accreditation requirements by promoting high quality practices which are auditable. SMIs also provide a reference point for method development. The performance of SMIs depends on competent staff and appropriate quality reagents and equipment. Laboratories should ensure that all commercial and in-house tests have been validated and shown to be fit for purpose. Laboratories should participate in external quality assessment schemes and undertake relevant internal quality control procedures.

Patient and public involvement
The SMI Working Groups are committed to patient and public involvement in the development of SMIs. By involving the public, health professionals, scientists and voluntary organisations the resulting SMI will be robust and meet the needs of the user. An opportunity is given to members of the public to contribute to consultations through our open access website.

Information governance and equality
PHE is a Caldicott compliant organisation. It seeks to take every possible precaution to prevent unauthorised disclosure of patient details and to ensure that patient-related records are kept under secure conditions. The development of SMIs are subject to PHE Equality objectives https://www.gov.uk/government/organisations/public-health-england/about/equality-and-diversity.

The SMI Working Groups are committed to achieving the equality objectives by effective consultation with members of the public, partners, stakeholders and specialist interest groups.

Legal statement
While every care has been taken in the preparation of SMIs, PHE and the partner organisations, shall, to the greatest extent possible under any applicable law, exclude liability for all losses, costs, claims, damages or expenses arising out of or connected with the use of an SMI or any information contained therein. If alterations are made by an end user to an SMI for local use, it must be made clear where in the document the alterations have been made and by whom such alterations have been made and also acknowledged that PHE and the partner organisations shall bear no liability for such alterations. For the further avoidance of doubt, as SMIs have been developed for application within the UK, any application outside the UK shall be at the user’s risk.

The evidence base and microbial taxonomy for the SMI is as complete as possible at the date of issue. Any omissions and new material will be considered at the next review. These standards can only be superseded by revisions of the standard, legislative action, or by NICE accredited guidance.

SMIs are Crown copyright which should be acknowledged where appropriate.

Suggested citation for this document
Investigation of swabs from skin and superficial soft tissue infections

http://www.hpa.org.uk/SMI/pdf
Scope of document

Type of specimen
Skin swab, swab from superficial, non-surgical and surgical wounds, and swab of pus

This SMI describes the processing of skin, superficial, non-surgical and surgical wound swabs, from sites accessible without intervention, for the microbiological investigation of skin and superficial soft tissue infections (SSTIs).

For pragmatic reasons the processing of swabs of pus has been included in this SMI. For further information regarding pus and exudate samples refer to B 14 – Investigation of pus and exudates.

It should be noted that many conditions are best diagnosed by submission of a skin biopsy for culture and histopathological examination (refer to B 17 - Investigation of tissues and biopsies from deep-seated sites and organs).

For information regarding dermatophyte infections see B 39 - Investigation of dermatological specimens for superficial mycoses.

Investigation of genital ulcers is dealt with in B 28 - Investigation of genital tract and associated specimens. Viruses such as herpes simplex and varicella-zoster, as well as parasites and non-microbial agents, may also cause skin lesions but are outside the scope of this SMI.

This SMI should be used in conjunction with other SMIs.

Key recommendations

Swabs are a diverse and heterogeneous group of specimens.

The specimen type and clinical details must therefore be taken into consideration when processing samples. For example, swabs of pus should be investigated in a similar way to pus samples. In addition to the standard media recommended, supplementary media (ie fastidious anaerobic, cooked meat broth or equivalent) is also required for these samples. Refer to table 4.5.1.

A mechanism for urgent reporting should be in place to communicate key, clinically significant results in a timely manner.

Introduction

The skin is colonised by normally non-harmful flora. When the skin is broken as a result of trauma, burns, bites or surgical procedures, colonisation with a range of bacteria may occur. Infections of the skin and subcutaneous tissues are caused by a wide range of organisms, however the majority are caused by Staphylococcus aureus and β haemolytic streptococci groups A, C and G.

Particular organisms are often typically associated with specific clinical conditions in skin and soft tissue infections, however overlaps in clinical presentation do occur. Diagnosis is normally based on clinical presentation. Guidelines for diagnosis and management have been published which focus on a wide range of SSTIs from minor superficial to life threatening infections. Microbiological cultures may be undertaken to
establish the causative organism enabling antibiotic sensitivity testing which is essential to ensure optimal treatment regimens.

**Skin infections**

**Cellulitis and erysipelas**

Cellulitis and erysipelas are diffuse spreading infections of the skin and subcutaneous tissue excluding cutaneous abscesses and necrotizing fasciitis. Cellulitis involves the deeper layers of the skin and subcutaneous tissues, whereas erysipelas involves the upper dermis and superficial lymphatic system.

Cellulitis is commonly caused by:
- β-haemolytic streptococci (including *Streptococcus pyogenes*)
- *S. aureus*

Wound infections may be caused by a broader range of organisms which, in addition to above, may include:
- *Bacteroides* species
- anaerobic cocci
- *Bacillus cereus* (especially after trauma or orthopaedic surgery)
- enterobacteriaceae

Superficial swabs in the absence of a skin break are often unrewarding; skin biopsies may produce better results but they are not frequently done. Recurrent cellulitis can occur following damage to local venous or lymphatic drainage systems.

**Ecthyma gangrenosum**

Ecthyma gangrenosum is a focal skin lesion characterised by haemorrhage, necrosis and surrounding erythema. It is usually caused by:
- *Pseudomonas aeruginosa*
- haematogenous dissemination of fungal infection (eg *Candida* species and mucoraceae fungi)

Ecthyma gangrenosum may also rarely be caused by *Stenotrophomonas maltophilia*. Similar lesions found in patients who are neutropenic may be due to infection with *Aspergillus* species or *Fusarium* species. Diagnosis is usually based on clinical history and physical examination.

**Impetigo**

Impetigo is a superficial, intra-epidermal infection producing erythematous lesions that may be bullous or nonbullous. Bullous impetigo is caused by *S. aureus*. Nonbullous impetigo is most frequently caused by Lancefield Group A streptococci or *S. aureus*, and has occasionally been caused by streptococci of Lancefield Groups C and G.

**Erysipelas**

Erysipelas is a rare superficial infection of the skin. It primarily involves the dermis and the most superficial parts of the subcutaneous tissues, with prominent...
involvement of the superficial lymphatics. It presents as a painful, fiery red, oedematous area of skin, occasionally with small vesicles on the surface. The margins have sharply demarcated, raised borders and the skin surface can appear orange peel like.

**Erythrasma**
Erythrasma is a common, chronic, superficial skin infection of the stratum corneum caused by *Corynebacterium minutissimum*. It presents with fine, scaly, reddish-brown plaques usually in the axillae and is often misdiagnosed as mycotic infection. Diagnosis is most often made on clinical grounds rather than by culture.

**Superficial mycoses**
Superficial mycoses are cutaneous fungal infections that involve the hair or nails or the keratinized layer of the stratum corneum. A number of fungi can cause infection and are diagnosed through biopsy or aspirate. Normally skin scrapings are the specimens of choice (see [B 39 - Investigation of dermatological specimens for superficial mycoses](#)).

Causative organisms include:
- dermatophytes
- *Candida* species
- Lipophilic yeasts

**Paronychia**
Paronychia is a superficial infection of the nail fold occurring as either an acute or chronic condition. Common isolates include:
- *S. aureus*
- Lancefield Group A streptococci
- yeasts
- anaerobic bacteria

**Folliculitis**
Folliculitis is infection and inflammation of a hair follicle. Dome-shaped papules or pustules form. These are each pierced by a hair and surrounded by a rim of erythema. The condition is usually caused by *S. aureus*.

Other possible causes include:
- *Pseudomonas aeruginosa* (can follow exposure in swimming pools or whirlpools)
- *Candida* species (in patients receiving prolonged antibiotic or corticosteroid treatment)
- *Malassezia furfur* (in patients with diabetes or granulocytopenia or receiving corticosteroid treatment)

**Necrotising skin and soft tissue infections**

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The terminology used for necrotising soft tissue infections is not consistent. Terms may relate to the kind of pathogen, the tissues involved, or the presence or absence of gas in the tissues\textsuperscript{32,33}.

It is clinically important to recognise these conditions as urgent surgical intervention, as well as antimicrobial therapy, is essential. Appropriate specimens are blood, fluid from bullae, and tissue biopsies. Growth from swabs taken from the surface of a lesion tends to be misleading, often yielding mixed cultures of colonising organisms. Mortality rates are high (30-60\%)\textsuperscript{33}.

**Gangrene**

There are 4 main types of gangrene:

Meleney's progressive synergistic gangrene presents as a burrowing lesion or chronic gangrene of the skin usually following abdominal operations and results from mixed infections by organisms such as:

- *S. aureus*
- streptococci
- enterobacteriaceae
- pseudomonads
- anaerobic Gram negative bacilli\textsuperscript{34}

Gas gangrene is a necrotising process associated with systemic signs of toxaemia and gas is present in the tissues. It often follows traumatic injuries such as penetrating wounds or crush injuries. Gas gangrene is caused by:

- *Clostridium perfringens*
- other *Clostridium* species

These organisms may however colonise a wound without causing disease. Alternatively, they may cause a spreading cellulitis, or extend into the muscle causing myonecrosis\textsuperscript{10}. Classical gas gangrene is associated with clinical shock, leakage of serosanguinous fluid, tissue necrosis and presence of gas in the tissues.

Fournier's gangrene applies to the non-sporing anaerobes. These are particularly important causes of infection in the pelvic and scrotal areas, and are common causes of gangrene in ischaemic and diabetic limbs. They often occur in infections mixed with:

- enterobacteriaceae
- streptococci
- *Clostridium* species\textsuperscript{35}

Spontaneous gangrene occurs either with no apparent relation to trauma or following mild, non-penetrating trauma. It is most commonly seen in patients with colonic carcinoma, leukaemia or neutropaenia. The main causative organisms are\textsuperscript{36}:

- *C. perfringens*
- *Clostridium septicum*
Actinomycosis

Actinomycosis is a chronic suppurative infection characterised by abscess formation with the production of sulphur granules which mainly consist of micro-colonies of *Actinomyces* species. Usual sites of infection are around the jaw, chest or abdomen. Material should be drained from these abscesses and biopsies taken.

Necrotising fasciitis

Necrotising fasciitis is a serious, infrequently occurring infection primarily affecting the subcutaneous fat and superficial fascia of muscles and often the overlying soft tissues. The infection is most commonly caused by Group A streptococci. Swabs are not the sample of choice for the investigation of this infection.

Myositis

Myositis is not strictly within the scope of this document. It is an inflammation of the muscle which may be caused by bacterial, fungal or parasitic infection as well as non-infective conditions such as autoimmune disease or genetic disorders. Localised infection is usually due to bacteria or fungi, whereas viral and parasitic infections tend to be more diffuse. Necrotising myositis rapidly involves the entire muscle bed and may spread to adjacent tissues. Both polymicrobial and unimicrobial forms may be seen.

Pyomyositis is a purulent infection of skeletal muscle and occurs more commonly in tropical countries. It usually presents as a single abscess but multiple abscesses do occur. Most patients have no underlying predisposing condition, previous trauma accounting for only 25% of cases. The majority of cases are due to *S. aureus*. More rarely, fungi and viruses may cause infection in patients who are immunocompromised.

Mycetoma

Mycetoma occurs in people living in tropical and sub-tropical climates, usually following a puncture wound. The condition results from a chronic destructive process involving the skin, subcutaneous tissue, muscle and bone. Granulation of tissue develops with chronic inflammation and fibrosis and is characterised by a draining sinus and the presence of granules. A mycetoma can form anywhere in the body, but is more common in the lower extremities. Formation in the foot is known as Madura foot.

Mycetomata are divided into two categories based on the aetiological agents involved; actinomycetoma caused by aerobic actinomycetes and eumycetoma caused by moulds. There are at least twenty moulds that may cause this condition; the species involved are often associated with distinct geographical areas.

Ninety five percent of the cases are caused by:

**Eumycetoma:**

- *Acremonium* species
- *Leptosphaeria senegalensis*
- *Madurella grisea*
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- *M. mycetomatis*
- *Scedosporium (Pseudallescheria) apiospermum*
- *Pyrenochaeta romeroi*
- *Curvularia* species
- *Exophiala jeanselmei*
- *Phialophora verrucosa*

**Actinomycetoma:**
- *Actinomadura* species
- *Nocardia* species
- *Streptomyces* species
- *Maduralla* species

Organisms are found in tissue sinuses as aggregates of filaments. These are called granules but differ from the sulphur granules of actinomycosis in that they do not have the characteristic clubbed peripheral fringe. Granules obtained directly from tissue will ensure the best cultural recovery of the causative organism because granules found in sinus discharge contain only dead organisms. Surgical biopsy to obtain material for culture is important for diagnosis, especially if sinus discharge is culture-negative for aerobic actinomycetes or is contaminated by other bacteria: the processing of tissue specimens in possible cases of mycetoma is described in **B 17 - Investigation of tissues and biopsies from deep-seated sites and organs**.

**Carbuncles, foruncles, cutaneous, soft tissue and other abscesses**

Carbuncles are deep and extensive subcutaneous abscesses involving several hair follicles and sebaceous glands. Foruncles are abscesses which begin in hair follicles as firm, tender, red nodules that become painful and fluctuant. Both carbuncles and foruncles are usually caused by *S. aureus*.

Cutaneous abscesses are usually painful, tender, fluctuant erythematous nodules often with a pustule on top. In some cases they are associated with extensive cellulitis, lymphangitis, lymphadenitis and fever. They are caused by a variety of organisms. The location of an abscess often determines the flora likely to be isolated. Thus *S. aureus* is most often isolated from cutaneous abscesses of the axillae, the extremities and the trunk, whereas cutaneous abscesses involving the vulva and buttocks may yield faecal or urogenital mucosal flora.

*Burkholderia pseudomallei* causes melioidosis, but is rare in the UK. The disease may present in a variety of forms with skin lesions and/or cellulitis. Diagnosis is made by blood culture, serology or culture of pus (refer to **B 37 – Investigation of blood culture (for organisms other than Mycobacterium species)**).
Abscesses in intravenous drug users

Cutaneous abscesses frequently occur as a complication of injecting drug use. They commonly result from the use of non-sterile solutions in which the drug is dissolved or from lubrication of the needle using saliva. Bacterial isolates include:

- oral streptococci
- \textit{Streptococcus anginosus} group
- \textit{Fusobacterium nucleatum}
- \textit{Prevotella} species
- \textit{Porphyromonas} species
- \textit{Porphyromonas} species
- \textit{Porphyromonas} species
- \textit{Porphyromonas} species
- \textit{S. aureus}
- \textit{Clostridium} species
- \textit{Bacillus anthracis}
  (this is a rare but severe infection that can occur by injecting heroin contaminated with anthrax)

Scalp abscess

Scalp abscesses are a recognised complication of electronic monitoring with fetal scalp electrodes during labour. A localised collection of pus surrounded by inflamed tissue forms where the electrodes are inserted. Anaerobes are most commonly isolated, probably as a result of contamination with vaginal organisms during delivery. Polymicrobial infections also occur, involving:

- anaerobes
- \(\beta\)-haemolytic streptococci
- \textit{S. aureus}
- enterobacteriaceae
- enterococci
- coagulase negative staphylococci

Kerion is a pustular folliculitis of adjacent hair follicles, creating dense inflamed areas of the scalp, and is caused by dermatophytes (refer to \textbf{B 39 – Investigation of dermatological specimens for superficial mycoses}). Secondary bacterial infection may occur.

Ulcers

A skin ulcer is a lesion of the skin with loss of the skin integrity, which can extend from the epidermis down to deeper layers. There are various types of ulcers with different aetiology: pressure sores, diabetic foot ulcers, venous leg ulcers, arterial ulcers, autoimmune conditions such as pemphigus/pemphgoid. All ulcers are invariably colonised by a polymicrobial flora and microbiology samples should be taken only if a clinical diagnosis of infection has been made. When swabs are taken from infected ulcers, they should be taken after cleansing and debridement: this aims at eliminating...
part of the superficial colonising flora. Sometimes chronic ulcer swabs are taken to identify the cause of underlying bone infections: in this scenario invasive bone biopsy specimens would be preferable, but ulcer swabs (after cleansing and debridement) are often taken in real practice but the results need careful interpretation.

Swabs from chronic non-healing ulcers or skin lesions with one of the following risk factors reported should be tested for Corynebacterium species:

- travel abroad to high risk area within the last 10 days
- contact with someone who has been to a high risk area within the last 10 days
- the patient works in a clinical microbiology laboratory, or similar occupation, where Corynebacterium species may be handled
- Corynebacterium diphtheriae, Corynebacterium ulcerans and Corynebacterium pseudotuberculosis can cause diphtheria and have been isolated from the skin of patients with chronic skin infections. For more information refer to ID 2 - Identification of Corynebacterium species.

Burns

Patients suffering from severe burns are at a higher risk of both local and systemic infection; sepsis is an important cause of mortality in this group of patients.

Organisms encountered include:

- S. aureus
- β-haemolytic streptococci
- pseudomonads, especially Pseudomonas aeruginosa
- Acinetobacter species
- Bacillus species
- enterobacteriaceae
- filamentous fungi, eg: Fusarium species and Aspergillus species
- Candida albicans, non-albicans Candida species and other yeasts
- coagulase negative staphylococci

Gram negative organisms cause the most severe infections; fungal infections on the other hand can spread quickly, but are more easily treated, although a definitive diagnosis is difficult to obtain.

Bite wounds and contact with animals

Bite wounds

Bite wounds can become contaminated by oral flora and normal human skin flora. Most bites are due to cats and dogs, but some are due to other pets (including reptiles, rodents and birds), domesticated animals (including horses, sheep etc) wild animals or other humans. Organisms most commonly isolated include Pasteurella multocida and S. aureus.
• α-haemolytic streptococci
• streptococcus anginosus group

Other organisms associated with bite wounds which are rarely isolated include:
• anaerobes (including *Bacteroides* species and Fusobacteria)
• *Capnocytophaga* species
• *Eikenella corrodens*
• *Haemophilus* species
• coagulase negative staphylococci
• *Streptobacillus moniliformis*
• *Staphylococcus intermedius*
• anaerobes (including Fusobacterium, Porphyromonas, Prerevotella etc)

*Capnocytophaga canimorsus* is associated with dog bites and causes sepsicaemia, particularly in patients with asplenia or underlying hepatic disease. This organism is usually isolated only from blood cultures.

*Streptobacillus moniliformis* is associated with rat bites and diagnosis is confirmed by culturing the organism from blood or joint fluid.

Other unusual organisms may be isolated including *Weeksella zoohelcum*, *Actinobacillus* species and *Neisseria canis*.

Insect bites are often associated with secondary Lancefield Group A streptococcus and *S. aureus* infection.

**Contact with animals or animal products**

**Erysipeloid**

Erysipeloid is an uncommon nonsuppurative cellulitis due to *Erysipelothrix rhusiopathiae*. It is an occupational disease of fishermen, fish handlers, butchers and abattoir workers. It affects the hands and fingers causing lesions which present as painful purplish areas of inflammation with erythematous advancing edges.

**Aeromonas and non-cholera Vibrio species**

*Aeromonas* and non-cholera *Vibrio* species are predominantly isolated from traumatic water-related wounds or lacerations received whilst swimming in fresh or salt water, from other environmentally contaminated wounds, or from fishing or shellfish inflicted injuries. Aeromonas infection may also follow the therapeutic use of leeches. Water-related injuries can be polymicrobial involving environmental Gram negative organisms such as *Edwardsiella tarda* and pseudomonads.

**Bacillus anthracis**

*Bacillus anthracis* is the causative agent of anthrax which appears clinically in one of several forms; cutaneous (skin) anthrax or inhalation anthrax, as well as, more recently, injective anthrax. Following the deliberate release of *B. anthracis* in the USA in 2001, there has been an increased awareness of the release of this and other organisms which may pose a biological threat. Cutaneous anthrax occurs through inoculation of spores to the skin or by contamination of abrasions. Skin lesions known
as malignant pustules develop, which are characteristic ulcers with a black centre. They are rarely painful, but if untreated the infection can spread to cause septicaemia. If untreated, the disease can be fatal in 5% of cases, but with antibiotic treatment recovery is usual. Cutaneous infection with *B. anthracis* can occur in industrial workers who use materials of animal origin such as wool, leather, bristles and fur, or in the agricultural workplace for example farmers, husbandmen, butchers and vets. In rare cases *B. anthracis* has been transmitted via insect bites.

**Other skin infections**

Skin infections may also be caused by the following:

- **MRSA** may colonise and/or infect wounds and soft tissue. Newly emerging community (mecIV) MRSA with virulence factors such as Panton-Valentine Leukocidin (PVL) or Scalded Skin Toxin (SST) cause highly contagious infections such as folliculitis in healthy children and young adults. Infections are often spread through poor hygiene. Panton-Valentine Leukocidin (PVL) is a toxin which is capable of destroying white blood cells. Scalded skin syndrome (Lyell's syndrome in older children; Ritter's syndrome in infants) is caused by *S. aureus* phage types group II and 7.

- *Mycobacterium* species can cause cutaneous infections. These may signify a disseminated systemic infection or may represent a local infection by non-tuberculcous mycobacteria (see B 40 - Investigation of specimens for *Mycobacterium* species).

- rapid growing mycobacterial strains such as *M. chelonae* and *M. fortuitum* have also been isolated from superficial skin infections. *M. chelonae* has been shown to be associated with tattoo related infections.

- *Sporothrix schenckii* causes sporotrichosis. Cutaneous sporotrichosis is acquired by contamination with soil, sphagnum moss or other vegetable matter and develops at the site of inoculation to form a primary lesion with lymphatic spread (see B 39 - Investigation of dermatological specimens for superficial mycoses). It is more common in warmer climates.

- cutaneous salmonellosis and listeriosis may also occur in veterinarians and farmers, typically on the arms, following assisted delivery of farm animals, usually cattle infected in utero. Cutaneous listeriosis in a patient with AIDS has also been reported.

- *Yersinia enterocolitica* can cause cutaneous infections.
Technical information/limitations

Limitations of UK SMIs

The recommendations made in UK SMIs are based on evidence (eg sensitivity and specificity) where available, expert opinion and pragmatism, with consideration also being given to available resources. Laboratories should take account of local requirements and undertake additional investigations where appropriate. Prior to use, laboratories should ensure that all commercial and in-house tests have been validated and are fit for purpose.

Selective media in screening procedure

Selective media which does not support the growth of all circulating strains of organisms may be recommended based on the evidence available. A balance therefore must be sought between available evidence, and available resources required if more than one media plate is used.

Specimen containers

SMIs use the term “CE marked leak proof container” to describe containers bearing the CE marking used for the collection and transport of clinical specimens. The requirements for specimen containers are given in the EU in vitro Diagnostic Medical Devices Directive (98/79/EC Annex 1 B 2.1) which states: “The design must allow easy handling and, where necessary, reduce as far as possible contamination of and leakage from, the device during use and, in the case of specimen receptacles, the risk of contamination of the specimen. The manufacturing processes must be appropriate for these purposes”.

Anaerobic plate incubation

The recommended incubation time for anaerobic plates is 48 hours. However some anaerobic bacteria such as certain species of Actinomyces require longer incubation (7 days) and will not be detected if plates are examined sooner.

Rapid methods

To reduce turnaround times, rapid identification and sensitivity tests may be performed in conjunction with routine methods where appropriate. A variety of rapid identification and sensitivity methods have been evaluated; these include molecular techniques and the Matrix Assisted Laser Desorption Ionisation Time-of-Flight (MALDI-TOF). It is important to ensure that fresh cultures of pure single isolates are tested to avoid reporting misleading results.

Laboratories should follow manufacturers’ instructions and all rapid tests must be validated and be shown to be fit for purpose prior to use.
1 Safety considerations

1.1 Specimen collection, transport and storage

Use aseptic technique. Collect swabs into appropriate transport medium and transport in sealed plastic bags. Compliance with postal, transport and storage regulations is essential.

1.2 Specimen processing

Containment Level 2.

If infection with a Hazard Group 3 organism, eg Bacillus anthracis (cutaneous anthrax is rare but needs to be recognised as a possibility in certain settings such as exposure to animal hides, injection of contaminated heroin in IVDUs and bioterrorist events such as the dissemination of spores in letters that took place in the USA in 2001), all specimens must be processed in a microbiological safety cabinet under full Containment Level 3 conditions.

Laboratory procedures that give rise to infectious aerosols must be conducted in a microbiological safety cabinet.

Refer to current guidance on the safe handling of all organisms documented in this SMI.

The above guidance should be supplemented with local COSHH and risk assessments.

2 Specimen collection

2.1 Type of specimens

Skin swab, swab from superficial, non-surgical and surgical wounds, swabs of pus

2.2 Optimal time and method of collection

For safety considerations refer to Section 1.1.

Collect specimens before starting antimicrobial therapy where possible.

Unless otherwise stated, swabs for bacterial and fungal culture should then be placed in appropriate transport medium.

Samples of pus/exudate, if present, are preferred to swabs (see B 14 – Investigation of deep-seated and organ, infections and abscesses). If only a minute amount of pus or exudate is available it is preferable to send a pus/exudate swab in transport medium to minimise the risk of desiccation during transport.

Sample a representative part of the lesion. Swabbing dry crusted areas is unlikely to yield the causative pathogen.

If specimens are taken from ulcers, the debris on the ulcer should be removed and the ulcer should be cleaned with saline. A biopsy or, preferably, a needle aspiration of the edge of the wound should be taken.

A less invasive irrigation-aspiration method may be preferred. Place the tip of a small needleless syringe under the ulcer margin and irrigate gently with at least 1mL sterile
0.85% NaCl without preservative. After massaging the ulcer margin, repeat the irrigation with a further 1mL sterile saline. Massage the ulcer margin again, aspirate approximately 0.25mL of the fluid and place in a CE marked leak proof container\textsuperscript{104}.

Fungal specimens for dermatophytes: See \textit{B 39 - Investigation of dermatological specimens for superficial mycoses}.

### 2.3 Adequate quantity and appropriate number of specimens\textsuperscript{96}

Numbers and frequency of specimens collected are dependent on clinical condition of patient.

### 3 Specimen transport and storage\textsuperscript{78,79}

#### 3.1 Optimal transport and storage conditions

For safety considerations refer to Section 1.1.

Specimens should be transported and processed as soon as possible\textsuperscript{96}.

If processing is delayed, refrigeration is preferable to storage at ambient temperature\textsuperscript{96}.

### 4 Specimen processing/procedure\textsuperscript{78,79}

#### 4.1 Test selection

N/A

#### 4.2 Appearance

N/A

#### 4.3 Sample preparation

For safety considerations refer to Section 1.2.

##### 4.3.1 Pre-treatment

N/A

##### 4.3.2 Specimen processing

See \textit{Q 5 - Inoculation of culture media for bacteriology}.

#### 4.4 Microscopy

##### 4.4.1 Standard

Gram stain is not normally required. However, Gram films should be considered from pus swabs if they originate from severe deep seated infections.

##### 4.4.2 Supplementary

See \textit{B 40 - Investigation of specimens for Mycobacterium species}, and \textit{TP 39 - Staining procedures}.
### 4.5 Culture and investigation

Inoculate each agar plate directly by rolling the swab on a part of the plate or by using a sterile pipette (Q 5 - Inoculation of culture media for bacteriology).

For the isolation of individual colonies, spread inoculum with a sterile loop.

#### 4.5.1 Culture media, conditions and organisms

<table>
<thead>
<tr>
<th>Clinical details/ conditions</th>
<th>Specimen</th>
<th>Standard media</th>
<th>Incubation</th>
<th>Cultures read</th>
<th>Target organism(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Temp °C</td>
<td>Atmos</td>
<td>Time</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>5 -10% CO₂</td>
<td>Air</td>
<td></td>
</tr>
<tr>
<td>All conditions</td>
<td>Swabs</td>
<td>Blood agar</td>
<td>35-37</td>
<td>40-48hr</td>
<td>Daily</td>
</tr>
<tr>
<td></td>
<td></td>
<td>And/or</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Staph/Strep selective agar</td>
<td>35-37</td>
<td>40-48hr</td>
<td>Daily</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Air</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>CLED/MacConkey agar</td>
<td>35-37</td>
<td>18-24hr</td>
<td>&gt;18hr</td>
</tr>
</tbody>
</table>

For these situations, add the following:

<table>
<thead>
<tr>
<th>Clinical details/ conditions</th>
<th>Specimen</th>
<th>Supplementary media</th>
<th>Incubation</th>
<th>Cultures read</th>
<th>Target organism(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Temp °C</td>
<td>Atmos</td>
<td>Time</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Air</td>
<td></td>
</tr>
<tr>
<td>Wound swabs eg traumatic wounds</td>
<td>Swabs</td>
<td>Selective anaerobe agar with metronidazole 5μg disc</td>
<td>35-37</td>
<td>Anaerobic</td>
<td>≥40hr* and at 5 d</td>
</tr>
<tr>
<td>Swabs of pus</td>
<td></td>
<td>Subculture to BA if evidence of growth (≥40hr), or at day 5</td>
<td>35-37</td>
<td>Air</td>
<td>40-48hr</td>
</tr>
</tbody>
</table>

For these situations, add the following:

<table>
<thead>
<tr>
<th>Clinical details/ conditions</th>
<th>Specimen</th>
<th>Supplementary media</th>
<th>Incubation</th>
<th>Cultures read</th>
<th>Target organism(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Temp °C</td>
<td>Atmos</td>
<td>Time</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>5 -10% CO₂</td>
<td>daily</td>
</tr>
</tbody>
</table>

For these situations, add the following:

<table>
<thead>
<tr>
<th>Clinical details/ conditions</th>
<th>Specimen</th>
<th>Supplementary media</th>
<th>Incubation</th>
<th>Cultures read</th>
<th>Target organism(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Temp °C</td>
<td>Atmos</td>
<td>Time</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>5 -10% CO₂</td>
<td>40-48hr</td>
<td></td>
</tr>
</tbody>
</table>

- **Top pathogens:**
  - Lancefield Groups A, C and G streptococci
  - *S. aureus*
  - In specific circumstances e.g. bites or exposure to animals and animal products or fresh/salt water (use blood agar):
    - *Pasteurella* species
    - *Vibrio* species
    - *Aeromonas* species
    - *Bacillus cereus/anthracis*
    - *Strep. pneumoniae*
    - *Eikenella corrudens*
    - Capnocytophaga
    - Erysipelothrix

- **Clinical circumstances determines significance of the following isolates**
  - Enterobacteriaceae
  - Pseudomonads

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### 4.5.2 Supplementary investigations

Toxigenicity testing of *C. diphtheriae*.

See [B 40 - Investigation of specimens for *Mycobacterium* species](bacteriology-206x795) for further details.

### 4.6 Identification

Refer to individual SMIs for organism identification.

#### 4.6.1 Minimum level of identification in the laboratory

<table>
<thead>
<tr>
<th>Organism</th>
<th>Level</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Aeromonas</strong></td>
<td>species level</td>
</tr>
<tr>
<td>Anaerobes</td>
<td>anaerobes level except in necrotising infections</td>
</tr>
<tr>
<td><strong>Bacillus species</strong></td>
<td>species level when appropriate to diagnose or exclude <em>B. anthracis</em> or <em>B. cereus</em> infections</td>
</tr>
<tr>
<td>β-haemolytic streptococci</td>
<td>Lancefield Group level</td>
</tr>
<tr>
<td>Coagulate negative staphylococci</td>
<td>coagulate negative level</td>
</tr>
<tr>
<td><strong>C. diphtheriae</strong></td>
<td>species level and urgent (same-day) toxigenicity test (when appropriate clinical details)</td>
</tr>
<tr>
<td><strong>C. minutissimum</strong></td>
<td>species level in erythrasma</td>
</tr>
<tr>
<td><strong>C. ulcerans</strong></td>
<td>species level (when appropriate clinical details)</td>
</tr>
<tr>
<td>Dermatophytes</td>
<td><a href="bacteriology-206x795">B 39 - Investigation of dermatological specimens for superficial mycoses</a></td>
</tr>
<tr>
<td><strong>E. corrodens</strong></td>
<td>species level</td>
</tr>
<tr>
<td>Enterobacteriaceae</td>
<td>coliforms level except in necrotising infections</td>
</tr>
</tbody>
</table>
### Investigation of swabs from skin and superficial soft tissue infections

<table>
<thead>
<tr>
<th>Organism</th>
<th>Level</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>E. rhusiopathiae</em></td>
<td>species level</td>
</tr>
<tr>
<td><em>Haemophilus</em></td>
<td>species level</td>
</tr>
<tr>
<td>Mould</td>
<td>genus level</td>
</tr>
<tr>
<td><em>Pasteurella</em></td>
<td>species level</td>
</tr>
<tr>
<td><strong>Pseudomonads</strong></td>
<td>Usually only at pseudomonads level except in echyma gangrenosum, recreational water folliculitis, necrotising infections, burns</td>
</tr>
<tr>
<td><em>S. aureus</em></td>
<td>species level</td>
</tr>
<tr>
<td>(consider Panton-Valentine leukocidin (PVL) and toxin testing if appropriate clinical details)</td>
<td></td>
</tr>
<tr>
<td><em>S. pneumoniae</em></td>
<td>species level</td>
</tr>
<tr>
<td>Yeasts</td>
<td>yeasts level</td>
</tr>
<tr>
<td><em>Vibrio</em></td>
<td>species level</td>
</tr>
</tbody>
</table>

Organisms may be further identified if this is clinically or epidemiologically indicated.

**Note:** All work on suspected isolates of *C. diphtheriae* which is likely to generate aerosols must be performed in a safety cabinet.

A medical microbiologist must be informed of all suspected isolates of *C. diphtheriae* as soon as possible (same-day toxigenicity testing is available from the reference laboratory).

### 4.7 Antimicrobial susceptibility testing

Refer to [British Society for Antimicrobial Chemotherapy (BSAC)](https://www.bsac.org.uk), [EUCAST](http://www.eucast.org) and/or [CSLI](http://www.clsi.org) guidelines or manufacturer’s validation for proprietary methods.

This SMI recommends selective and restrictive reporting of susceptibilities to antimicrobials. Any deviation must be subject to consultation that should include local antimicrobial stewardship groups.
### 4.7.1 Antimicrobial susceptibility testing and reporting table

It is recommended that the antimicrobials in bold in the table below are reported. Those antimicrobials not in bold may be reported based on local decisions.

For more information on Detection of bacteria with Carbapenem-Hydrolysing β-lactamases (Carbapenemases) refer to B 60.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>Examples of agents to be included within primary test panel (recommended agents to be reported are in bold depending on clinical presentation)</th>
<th>Examples of agents to be considered for supplementary testing (recommended agents to be reported are in bold depending on clinical presentation)</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Enterobacteriaceae from clean surgical sites</td>
<td>Ampicillin (or Amoxicillin) Cefpodoxime⁴ Co-amoxiclav⁵ Gentamicin</td>
<td>Amikacin Aztreonam Cefotaxime (or Ceftriaxone) Ceftazidime Cefuroxime Ciprofloxacin Co-trimoxazole Ertapenem Meropenem (or Imipenem) Piperacillin/Tazobactam Temocillin</td>
<td>3. Antibiotics should only be reported in the presence of clinical evidence of infection. 4. Cefpodoxime resistant organisms should be tested for the presence of ESBLs and screened for reduced susceptibility to carbapenems. 5. Co-amoxiclav resistant organisms should be tested at local level for sensitivity to carbapenems.</td>
</tr>
<tr>
<td>Enterobacteriaceae from sites prone to colonisation (e.g. ulcers)</td>
<td>Amikacin, Ampicillin (or Amoxicillin), Aztreonam, Cefpodoxime&lt;sup&gt;4, 6&lt;/sup&gt;, Cefuroxime, Ciprofloxacin, Ceftazidine, Cefotaxime (or Ceftriaxone), Co-amoxiclav&lt;sup&gt;5, 6&lt;/sup&gt;, Cotrimoxazole, Ertapenem, Gentamicin, Meropenem (or Imipenem), Piperacillin/Tazobactam, Temocillin</td>
<td>4. Cefpodoxime resistant organisms should be tested for the presence of ESBLs and screened for reduced susceptibility to carbapenems. 5. Co-amoxiclav resistant organisms should be tested at local level for sensitivity to carbapenems. 6. If susceptibility testing is being undertaken, include this agent.</td>
<td></td>
</tr>
<tr>
<td>Pseudomonads</td>
<td>Amikacin, Ceftazidine, Ciprofloxacin, Gentamicin, Meropenem (or Imipenem), Piperacillin/Tazobactam</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### 4.8 Referral for outbreak investigations

N/A

### 4.9 Referral to reference laboratories

For information on the tests offered, turnaround times, transport procedure and the other requirements of the reference laboratory, click here for user manuals and request forms.

Organisms with unusual or unexpected resistance, or associated with a laboratory or clinical problem, or anomaly that requires elucidation should be sent to the appropriate reference laboratory.

Contact appropriate devolved national reference laboratory for information on the tests available, turnaround times, transport procedure and any other requirements for sample submission:

England and Wales


Scotland

5  Reporting procedure

5.1  Microscopy

Standard
Gram stain (not usually required)
Report on WBCs and organisms detected.

Supplementary
For the reporting of microscopy for *Mycobacterium* species refer to B 40 – Investigation of specimens for *Mycobacterium* species.

5.1.1  Microscopy reporting time
All results should be issued to the requesting clinician as soon as they become available, unless specific alternative arrangements have been made with the requestors.

Urgent results should be telephoned or transmitted electronically in accordance with local policies.

5.2  Culture

Following results should be reported:

- clinically significant organisms isolated
- other growth
- absence of growth

5.2.1  Culture reporting time
Interim or preliminary results should be issued on detection of potentially clinically significant isolates as soon as growth is detected, unless specific alternative arrangements have been made with the requestors.

Urgent results should be telephoned or transmitted electronically in accordance with local policies.

Final written or computer generated reports should follow preliminary and verbal reports as soon as possible.

5.3  Antimicrobial susceptibility testing

Report susceptibilities as clinically indicated. Prudent use of antimicrobials according to local and national protocols is recommended.

Refer to table 4.7.1. The table includes guidance on the minimum range of agents that should be tested on the bacterial isolates listed. The table also includes additional agents that can be considered for inclusion in test panels in specific clinical scenarios.
Investigation of swabs from skin and superficial soft tissue infections

Any deviation from the guidance should be subject to local consultation and risk assessment.

Generally, all resistant results should be reported as this is good practice and informs the user.
6 Notification to PHE\textsuperscript{105,106}, or equivalent in the devolved administrations\textsuperscript{107-110}

The Health Protection (Notification) regulations 2010 require diagnostic laboratories to notify Public Health England (PHE) when they identify the causative agents that are listed in Schedule 2 of the Regulations. Notifications must be provided in writing, on paper or electronically, within seven days. Urgent cases should be notified orally and as soon as possible, recommended within 24 hours. These should be followed up by written notification within seven days.

For the purposes of the Notification Regulations, the recipient of laboratory notifications is the local PHE Health Protection Team. If a case has already been notified by a registered medical practitioner, the diagnostic laboratory is still required to notify the case if they identify any evidence of an infection caused by a notifiable causative agent.

Notification under the Health Protection (Notification) Regulations 2010 does not replace voluntary reporting to PHE. The vast majority of NHS laboratories voluntarily report a wide range of laboratory diagnoses of causative agents to PHE and many PHE Health protection Teams have agreements with local laboratories for urgent reporting of some infections. This should continue.

**Note:** The Health Protection Legislation Guidance (2010) includes reporting of Human Immunodeficiency Virus (HIV) & Sexually Transmitted Infections (STIs), Healthcare Associated Infections (HCAIs) and Creutzfeldt–Jakob disease (CJD) under ‘Notification Duties of Registered Medical Practitioners’: it is not noted under ‘Notification Duties of Diagnostic Laboratories’.


Other arrangements exist in Scotland\textsuperscript{107,108}, Wales\textsuperscript{109} and Northern Ireland\textsuperscript{110}. 
Appendix: Investigation of skin and superficial soft tissue infections

Prepared specimens

All specimens

Standard media

Supplementary media

Blood agar and/or
Staph/Strep agar

CLED/ MacConkey

Wound swabs
Traumatic wounds
Swabs of pus

Selective anaerobe
agar with
metronidazole 5μg
disc

Anaerobes
Refer to
ID 8, 14, 15, 25

Anaerobes
Refer to
ID 8, 10, 14, 25

Swabs of pus

Fastidious
anaerobic,
cooked meat
broth or
equivalent

Subculture to
BA

Incubate at 35-37°C
5-10% CO₂ and/or
Air
40-48hr
Read daily

Incubate at 35-37°C
Air
18-24hr
Read ≥18hr

Incubate at 35-37°C
Anaerobic
5 d
Read ≥40hr and
≥5 d

Incubate at 35-37°C
Anaerobic
5 d
Read >40hr and
≥5 d

Incubate at 35-37°C
Up to 5 d
Read daily

Incubate at 35-37°C
5-10% CO₂
40-48hr
Read daily

Incubate at 35-37°C
5-10% CO₂
40-48hr
Read daily

Incubate at 35-37°C
Air
40-48hr
Read daily

Chocolate agar †

Sabouraud agar

Haemophilus
species
Refer to
ID 12

Haemophilus
species
Refer to
ID 16

Yeast
Mould

C. diphtheriae
C. ulcerans
Refer to ID 2

Suspected
cutaneous
diphtheria

Burns
Patients who are
immunocompromised
Diabetic patients
Intertrigo
Panorhymia

Cellulitis in children
Human bites

Swabs of pus

Enterobacteriaceae
Pseudomonads
Refer to
ID 16, 17

Pasteurella species
Refer to ID 13
S. aureus
Refer to ID 7
Vibrio species
Refer to ID 19
Aeromonas species
Erysipelothrix rhusiopathiae

Enterobacteriaceae
Pasteurella species
Aeromonas species

† Either bacitracin 10 unit disc or bacitracin-containing agar may be used

† Either bacitracin 10 unit disc or bacitracin-containing agar may be used

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References


2. Dryden MS. Complicated skin and soft tissue infection. JAntimicrobChemother 2010;65 Suppl 3:iii35-iii44.


Investigation of swabs from skin and superficial soft tissue infections


Investigation of swabs from skin and superficial soft tissue infections


78. European Parliament. UK Standards for Microbiology Investigations (SMIs) use the term "CE marked leak proof container" to describe containers bearing the CE marking used for the collection and transport of clinical specimens. The requirements for specimen containers are given in the EU in vitro Diagnostic Medical Devices Directive (98/79/EC Annex 1 B 2.1) which states: "The design must allow easy handling and, where necessary, reduce as far as possible contamination of, and leakage from, the device during use and, in the case of specimen receptacles, the risk of contamination of the specimen. The manufacturing processes must be appropriate for these purposes". 1998.


Investigation of swabs from skin and superficial soft tissue infections


