



Summary of Results

External Quality Assessment of Food Microbiology Standard Scheme

Distribution Number: 294

Sample Numbers: S0625, S0626

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Results Due:	14 July 2017
Report Date:	24 July 2017
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For further information on the scheme please refer to:

Scheme Guide: <https://www.gov.uk/government/publications/food-and-water-proficiency-testing-schemes-scheme-guide>

Guide to Scoring and Statistics:

<https://www.gov.uk/government/publications/food-and-water-proficiency-testing-schemes-scoring-systems-and-statistics>

General guidance for z-scores:

Participants' enumeration results are converted into z-scores using the following formula:

$$Z = \frac{(X_i - X_{pt})}{\sigma_{pt}}$$

X_i = participants' result (expressed as a log₁₀ value)
 X_{pt} = assigned value (participants' consensus median (expressed as a log₁₀ value))
 σ_{pt} = the fixed standard deviation for the examination (calculated by FEPTU)

The σ_{pt} -value expresses the acceptable difference between the individual participant's result and the participants' consensus median. The σ_{pt} -value used for calculating z-scores for all parameters in the Standard Scheme is 0.35. A guide to interpreting z-scores follows, although laboratories must interpret their scores in the context of their own laboratory situation:

z = -1.99 to +1.99 **satisfactory**
z = -2 to -2.99 or +2 to +2.99 **questionable**
z = < -3.00 or > + 3.00 **unsatisfactory**

It is usually recommended that z-scores exceeding +/-2 are investigated to establish the possible cause. As a general rule, PHE recommends that all questionable and unsatisfactory results are investigated.

FEPTU Quality control: To demonstrate homogeneity of the sample, a minimum of 10 freeze-dried vials, selected randomly from a batch, are tested in duplicate for parameters requiring enumeration and 10 freeze-dried vials are examined for pathogen detection.

To demonstrate stability of the sample, a minimum of nine vials, selected randomly from a batch, are examined throughout the distribution period, either for enumeration or pathogen detection.

FEPTU results are determined using methods based on ISO methods and are included in the 'intended results' letters which provide guidance for participants regarding the assigned values.

The FEPTU results are used for guidance in the preliminary intended results notification, letters are posted on the website immediately after every distribution; electronic notification of their availability is sent to all participants

Refer to section 17.0 of the Scheme Guide if you have experienced difficulties with any of the examinations .

<https://www.gov.uk/government/publications/food-and-water-proficiency-testing-schemes-scheme-guide>

All participants are reminded that reporting an incorrect or incomplete identification of pathogens from food samples could have serious public health implications. Similarly, the levels of micro-organisms reported in the sample may affect the subsequent outcome for the product.

Participants are reminded that the purpose of scoring is to draw attention to incorrect or outlying results. Participants who report outlying results for enumerations on single occasions only should not be alarmed, although they should still assess the reason(s) for the outlying result. The results, as summarised in the performance assessment sheet included in this report, provide a more effective indication of on-going problems with food microbiology examinations.

The bar charts in this report are compiled using the processes outlined in the Guide to Scoring Systems and Statistics for the allocation of PHE scores. Z-scores are included on the sample-specific pages only; the relevant sections will be left blank if a z-score does not apply.

Please contact FEPTU staff for advice and information:

Repeat samples	Carmen Gomes or Kermin Daruwalla	Tel: +44 (0)20 8327 7119
Data Analysis	Nita Patel or Manchari Rajkumar	Fax: +44 (0)20 8200 8264
Microbiological advice	Nita Patel or Zak Prior	Email: foodeqa@phe.gov.uk
General comments and complaints	Nita Patel or Zak Prior	FEPTU's website
Scheme consultants	Melody Greenwood and Julie E. Russell	
Scheme Co-ordinator	Nita Patel	

Accreditation: PHE Food EQA Scheme for Standard is accredited by the United Kingdom Accreditation Service (UKAS) to ISO/IEC 17043:2010.



0006

Sample: S0625

Contents: *Campylobacter jejuni* 1.4x10⁴ (wild strain), *Salmonella* Typhimurium 1,4 ,[5],12:i :1.2 39 (wild strain), *Klebsiella oxytoca* 2.1x10³ (wild strain), *Enterococcus faecalis* 1.6x10⁴ (wild strain)

All levels are presented as colony forming units (cfu) per ml reconstituted sample

All levels are presented as colony forming units (cfu) per ml reconstituted sample

Expected Results:

Examination	Expected Result	Your Result	Score	Z-score
<i>Campylobacter</i> spp.	Detected in 25g			
<i>Campylobacter</i> spp. enumeration	1.2x10 ³ - 1.7x10 ⁴ cfu g ⁻¹			
<i>E. coli</i> O157	Not Detected in 25g			
<i>Salmonella</i> spp.	Detected in 25g			
Aerobic colony count	5.1x10 ³ - 5.1x10 ⁴ cfu g ⁻¹			
<i>Enterobacteriaceae</i>	3.2x10 ² - 3.2x10 ³ cfu g ⁻¹			

Campylobacter spp.

Total participants reporting for <i>Campylobacter</i> spp.	98
Participants reporting correctly	94 (96%)

Campylobacter spp. enumeration

Total participants reporting for <i>Campylobacter</i> spp. enumeration	53
Total participants enumerating <i>Campylobacter</i> spp. enumeration	53
Participants reporting a low censored value	1
Assigned value (participants' median)	5.4x10 ³ cfu g ⁻¹ (3.73 log ₁₀)
Uncertainty of assigned value ($U(X_{pt}) = \log_{10} \text{cfu g}^{-1}$)	0.07
No. of outlying counts	8 (6 low / 2 high)
Participants mean	4.9x10 ³ cfu g ⁻¹ (3.69 log ₁₀)
Standard deviation of participants results *	0.39 log ₁₀ cfu g ⁻¹
FEPTU QC medians	
▪ ISO/TS 10272-2	1.4x10 ⁴ cfu g ⁻¹ (4.15 log ₁₀)

<i>E. coli</i> O157	
Total participants reporting for <i>E. coli</i> O157	70
Participants reporting correctly	70 (100%)

<i>Salmonella</i> spp.	
Total participants reporting for <i>Salmonella</i> spp.	150
Participants reporting correctly	148 (99%)

Aerobic colony count	
Total participants reporting for Aerobic colony count	146
Assigned value (participants' median)	1.6x10 ⁴ cfu g ⁻¹ (4.2 log ₁₀)
Uncertainty of assigned value ($U(X_{pt}) = \log_{10}$ cfu g ⁻¹)	0.01
No. of outlying counts	5 (1 low / 4 high)
Participants mean	1.7x10 ⁴ cfu g ⁻¹ (4.22 log ₁₀)
Standard deviation of participants results *	0.08 log ₁₀ cfu g ⁻¹
FEPTU QC median	1.6x10 ⁴ cfu g ⁻¹ (4.2 log ₁₀)

<i>Enterobacteriaceae</i>	
Total participants reporting for <i>Enterobacteriaceae</i>	140
Assigned value (participants' median)	1.0x10 ³ cfu g ⁻¹ (3 log ₁₀)
Uncertainty of assigned value ($U(X_{pt}) = \log_{10}$ cfu g ⁻¹)	0.02
No. of outlying counts	12 (10 low / 2 high)
Participants mean	9.5x10 ² cfu g ⁻¹ (2.98 log ₁₀)
Standard deviation of participants results *	0.17 log ₁₀ cfu g ⁻¹
FEPTU QC median	1.4x10 ³ cfu g ⁻¹ (3.15 log ₁₀)

Total sent samples	167
Non-returns	3
Not examined	2

The fixed standard deviation value (σ_{pt} value) used for calculation of the z-scores is **0.35** for all parameters.

* Robust S^* based on median absolute deviation about the participants' median (*MAD_e*).

Sample: S0626

Contents: *Campylobacter coli* 3.0x10³ (wild strain), *Escherichia coli* O157 2.2x10² (wild strain), *Enterobacter aerogenes* 2.1x10⁴ (wild strain), *Staphylococcus epidermidis* 2.5x10⁶ (wild strain)

All levels are presented as colony forming units (cfu) per ml reconstituted sample

All levels are presented as colony forming units (cfu) per ml reconstituted sample

Expected Results:

Examination	Expected Result	Your Result	Score	Z-score
<i>Campylobacter</i> spp.	Detected in 25g			
<i>Campylobacter</i> spp. enumeration	3.0x10 ² - 3.2x10 ³ cfu g ⁻¹			
<i>E. coli</i> O157	Detected in 25g			
<i>Salmonella</i> spp.	Not Detected in 25g			
Aerobic colony count	7.6x10 ⁴ - 7.6x10 ⁵ cfu g ⁻¹			
<i>Enterobacteriaceae</i>	3.8x10 ³ - 3.8x10 ⁴ cfu g ⁻¹			

Campylobacter spp.

Total participants reporting for <i>Campylobacter</i> spp.	80
Participants reporting correctly	74 (93%)

Campylobacter spp. enumeration

Total participants reporting for <i>Campylobacter</i> spp. enumeration	40
Total participants enumerating <i>Campylobacter</i> spp. enumeration	40
Participants reporting a low censored value	1
Assigned value (participants' median)	9.6x10 ² cfu g ⁻¹ (2.98 log ₁₀)
Uncertainty of assigned value ($U(\chi_{pt}) = \log_{10}$ cfu g ⁻¹)	0.08
No. of outlying counts	8 (4 low / 4 high)
Participants mean	1.0x10 ³ cfu g ⁻¹ (3.01 log ₁₀)
Standard deviation of participants results *	0.41 log ₁₀ cfu g ⁻¹
FEPTU QC medians <ul style="list-style-type: none"> ▪ ISO/TS 10272-2 	3.0x10 ³ cfu g ⁻¹ (3.48 log ₁₀)

<i>E.coli</i> O157	
Total participants reporting for <i>E.coli</i> O157	69
Participants reporting correctly	65 (94%)

<i>Salmonella</i> spp.	
Total participants reporting for <i>Salmonella</i> spp.	126
Participants reporting correctly	121 (96%)

Aerobic colony count	
Total participants reporting for Aerobic colony count	119
Assigned value (participants' median)	2.4x10 ⁵ cfu g ⁻¹ (5.38 log ₁₀)
Uncertainty of assigned value ($U(X_{pt}) = \log_{10}$ cfu g ⁻¹)	0.02
No. of outlying counts	5 (2 low / 3 high)
Participants mean	2.5x10 ⁵ cfu g ⁻¹ (5.39 log ₁₀)
Standard deviation of participants results *	0.14 log ₁₀ cfu g ⁻¹
FEPTU QC median	2.7x10 ⁵ cfu g ⁻¹ (5.43 log ₁₀)

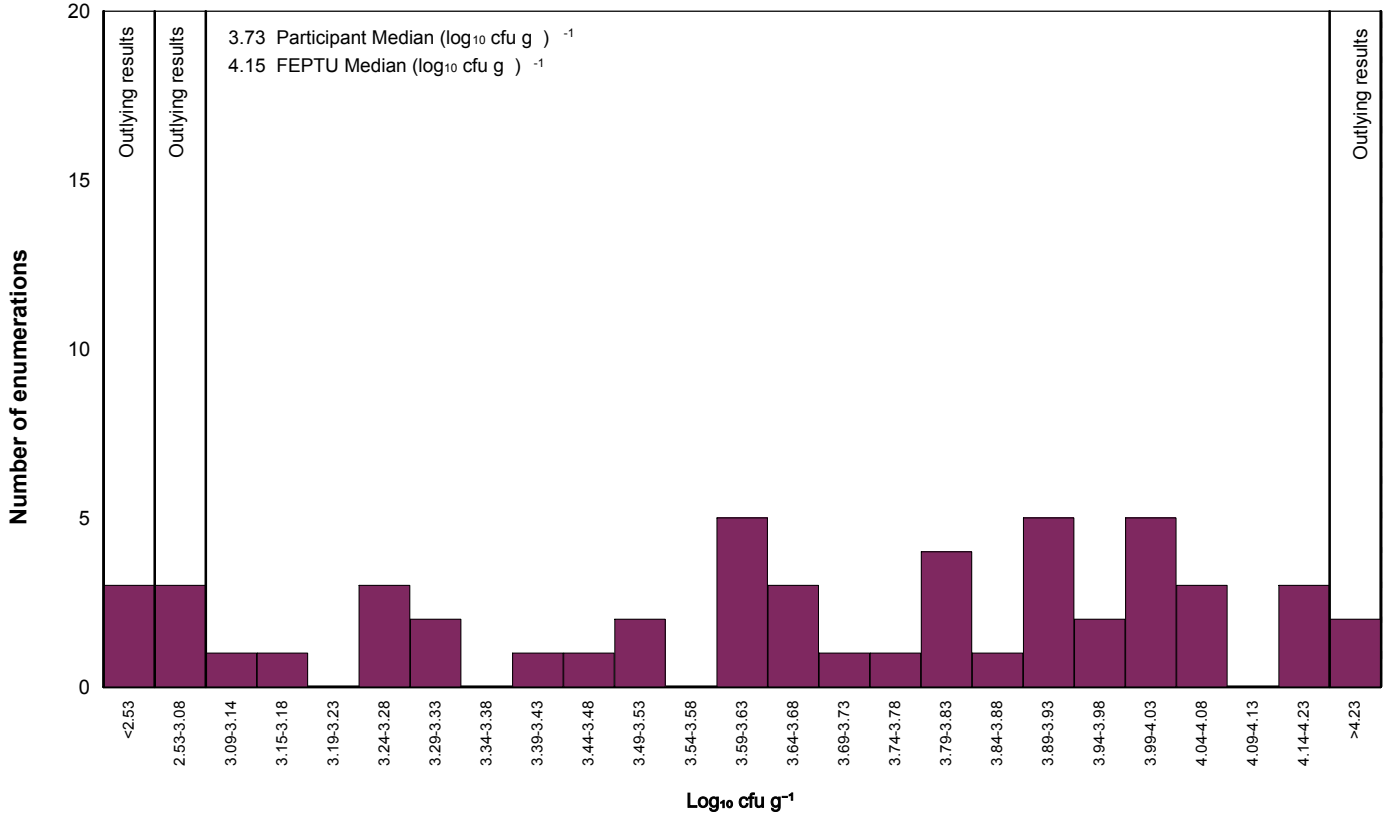
<i>Enterobacteriaceae</i>	
Total participants reporting for <i>Enterobacteriaceae</i>	115
Participants reporting a low censored value	1
Assigned value (participants' median)	1.2x10 ⁴ cfu g ⁻¹ (4.08 log ₁₀)
Uncertainty of assigned value ($U(X_{pt}) = \log_{10}$ cfu g ⁻¹)	0.02
No. of outlying counts	10 (9 low / 1 high)
Participants mean	1.1x10 ⁴ cfu g ⁻¹ (4.05 log ₁₀)
Standard deviation of participants results *	0.19 log ₁₀ cfu g ⁻¹
FEPTU QC median	1.7x10 ⁴ cfu g ⁻¹ (4.23 log ₁₀)

Total sent samples	167
Non-returns	3
Not examined	27

The fixed standard deviation value (σ_{pt} value) used for calculation of the z-scores is **0.35** for all parameters.

* Robust S^* based on median absolute deviation about the participants' median (MAD_e).

Campylobacter spp. enumeration reported by participants - Sample S0625



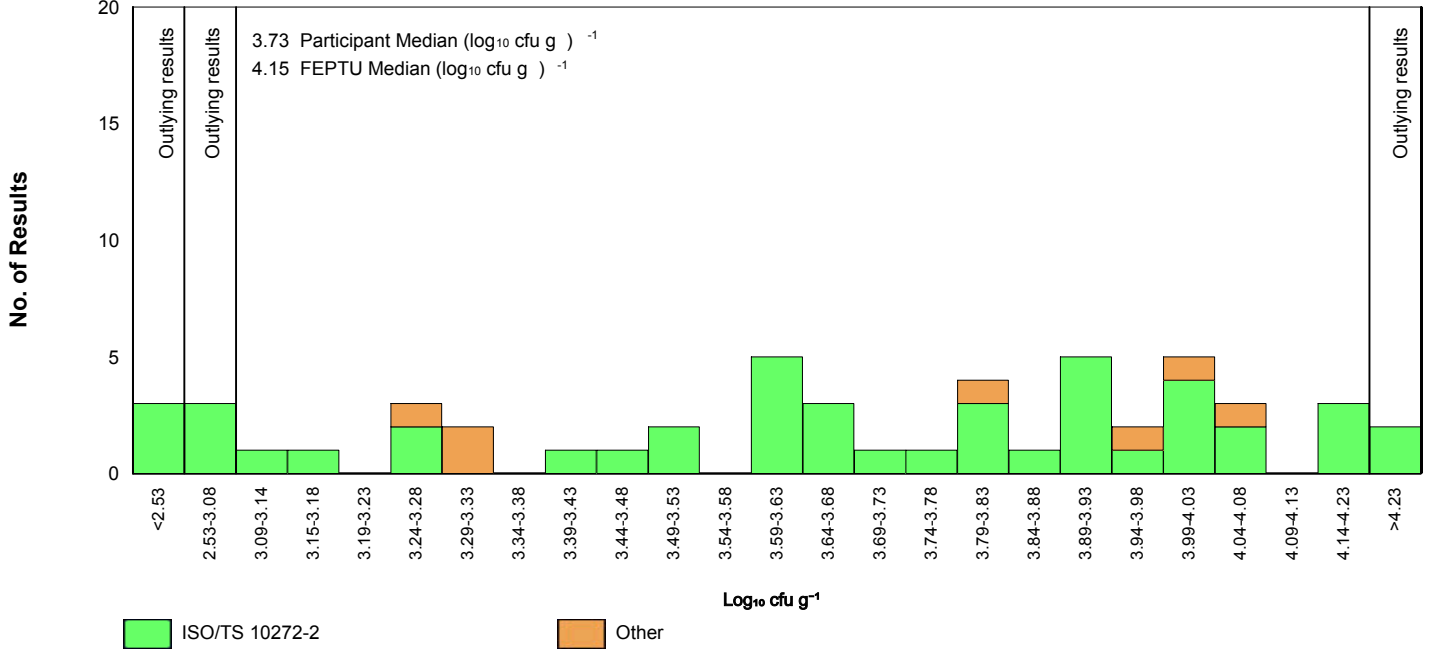
Method based presentation

S0625 : *Campylobacter* spp. enumeration

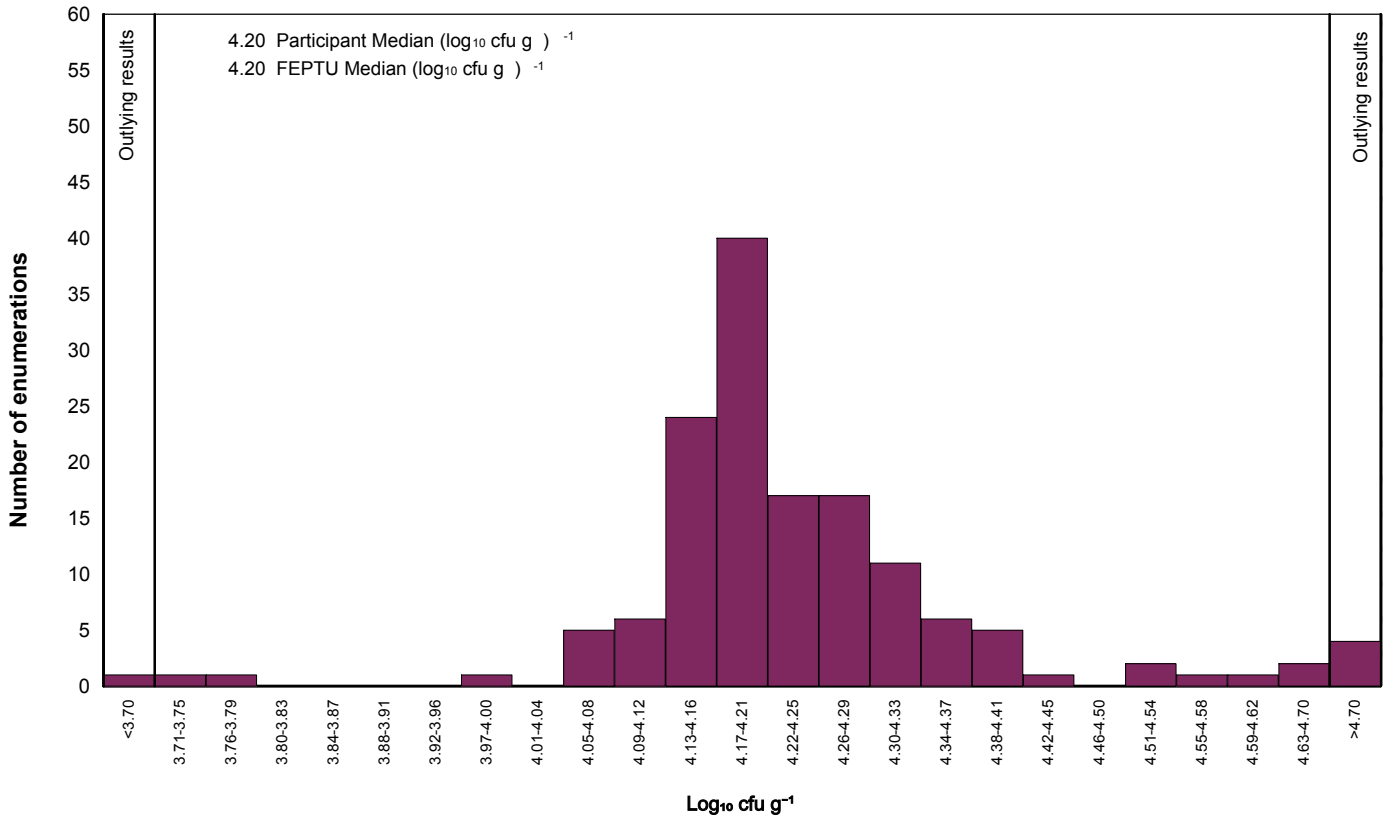
FEPTU Method: ISO/TS 10272-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (\log_{10} cfu g ⁻¹)	Robust S* (\log_{10} cfu g ⁻¹)	Range Reported (\log_{10} cfu g ⁻¹)
ISO/TS 10272-2	45	0	86	3.72	0.38	2.18 - 4.72
Other	7	1	13			-

Campylobacter spp. enumeration reported by participants - Sample S0625



Aerobic colony count reported by participants - Sample S0625



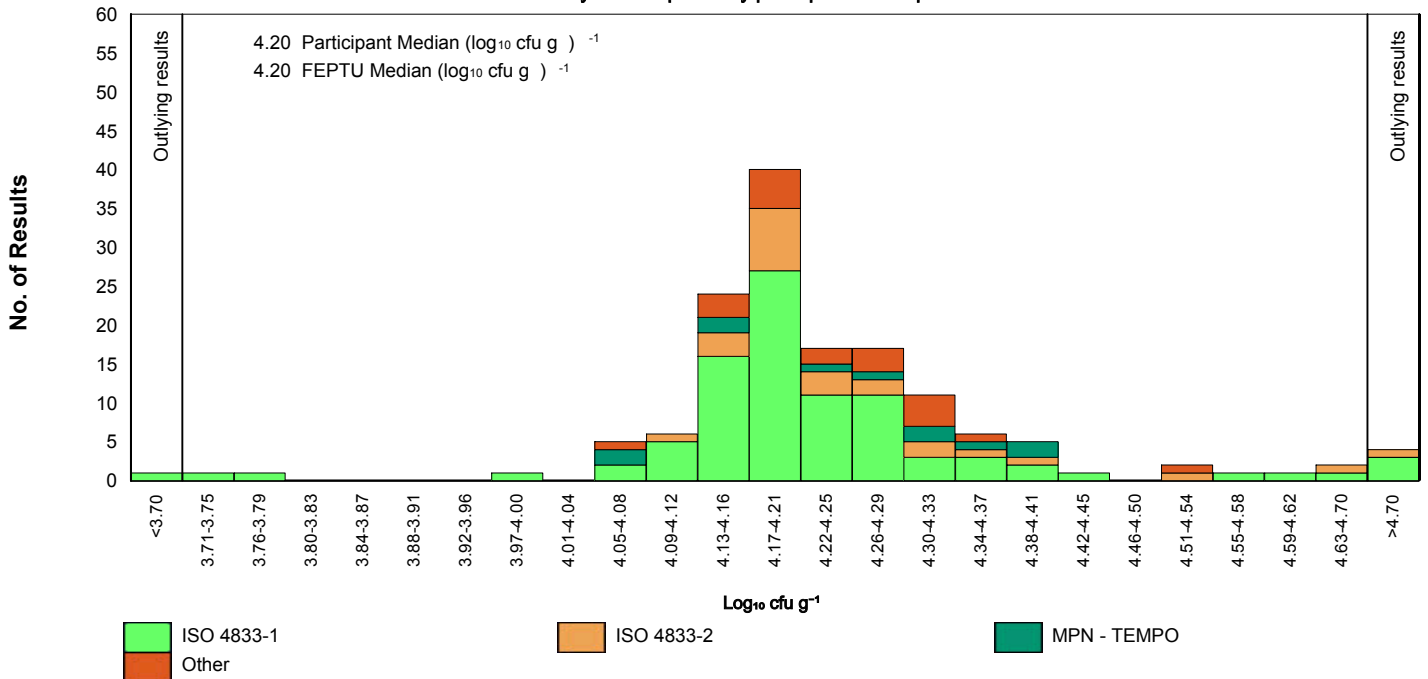
Method based presentation

S0625 : Aerobic colony count

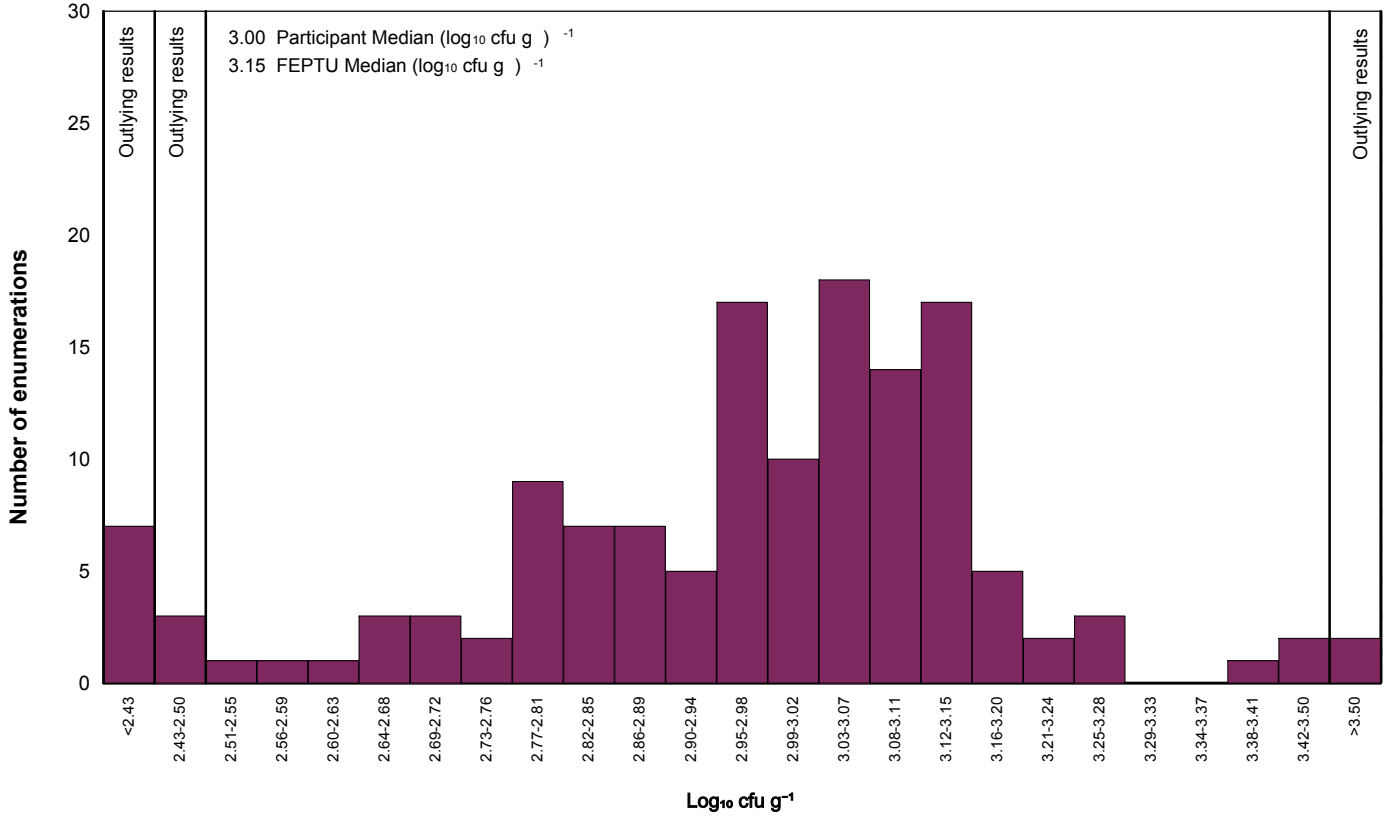
FEPTU Method: ISO 4833-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (\log_{10} cfu g ⁻¹)	Robust S* (\log_{10} cfu g ⁻¹)	Range Reported (\log_{10} cfu g ⁻¹)
ISO 4833-1	91	0	62	4.20	0.08	3.16 - 6.43
ISO 4833-2	24	0	16	4.21	0.08	4.11 - 5.28
MPN - TEMPO	11	0	7	4.25	0.14	4.04 - 4.40
Other	20	0	13	4.22	0.09	4.04 - 4.51

Aerobic colony count reported by participants - Sample S0625



Enterobacteriaceae reported by participants - Sample S0625



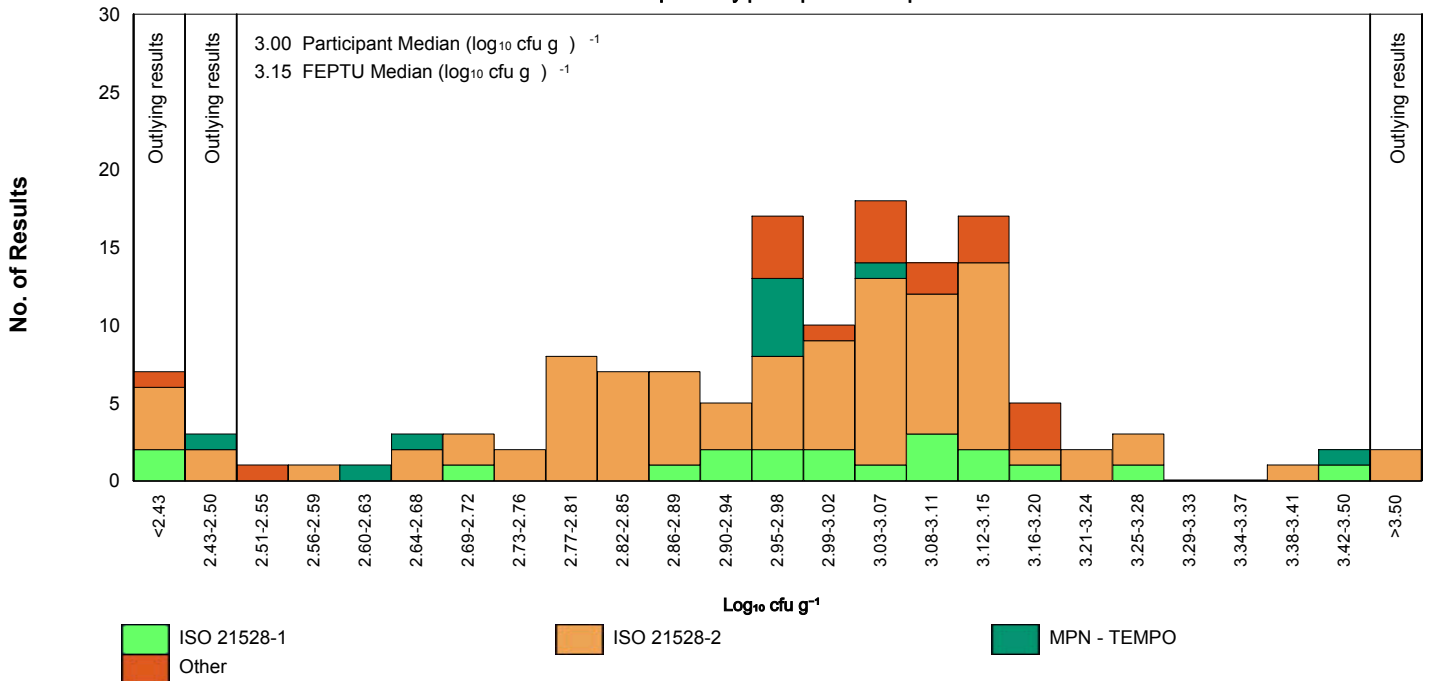
Method based presentation

S0625 : Enterobacteriaceae

FEPTU Method: ISO 21528-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (\log_{10} cfu g ⁻¹)	Robust S* (\log_{10} cfu g ⁻¹)	Range Reported (\log_{10} cfu g ⁻¹)
ISO 21528-1	19	0	13	3.00	0.17	1.65 - 3.49
ISO 21528-2	91	0	65	3.00	0.17	2.11 - 4.11
MPN - TEMPO	10	0	7	2.97	0.08	2.48 - 3.49
Other	19	0	13	3.04	0.11	2.00 - 3.18

Enterobacteriaceae reported by participants - Sample S0625



Sample S0625

<i>Campylobacter</i> spp. Method	<i>Campylobacter</i> spp. Media	<i>Campylobacter</i> spp. Enrichment	No. Participants detected	No. Participants not detected
ISO 10272-1	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	26	0
ISO 10272-1	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA); Chromogenic agar - please state	Bolton Broth	5	0
ISO 10272-1	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA); Other	Bolton Broth	11	0
ISO 10272-1	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA); Preston agar	Bolton Broth	6	0
ISO 10272-1; PCR	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	5	0
Other	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	5	0

Sample S0625

<i>Campylobacter</i> spp. enumeration Method	<i>Campylobacter</i> spp. enumeration Media	<i>Campylobacter</i> spp. enumeration Incubation	Count reported	Count censored values
	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		0	0
ISO/TS 10272-2	Chromogenic agar - please state		1	0
ISO/TS 10272-2	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		41	1
ISO/TS 10272-2	Other		2	0
ISO/TS 10272-2	Preston agar		1	0
Other	Chromogenic agar - please state		2	1
Other	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		4	0
Other	Other		1	0

Sample S0625

<i>Salmonella</i> spp. Method	<i>Salmonella</i> spp. Media	<i>Salmonella</i> spp. Enrichment	No. Participants detected	No. Participants not detected
ISO 6579	Brilliant green agar (BGA); Xylose lysine deoxycolate agar (XLD)	Muller-Kauffman tetrathionate/novobiocin ; Rappaport-Vassiliadis with soya	5	0
ISO 6579	Brilliant green agar (BGA); Xylose lysine deoxycolate agar (XLD)	Rappaport-Vassiliadis with soya; Muller-Kauffman tetrathionate/novobiocin	13	0
ISO 6579	Chromogenic agar - please state; Xylose lysine deoxycolate agar (XLD)	Muller-Kauffman tetrathionate/novobiocin ; Rappaport-Vassiliadis with soya	5	0
ISO 6579	Chromogenic agar - please state; Xylose lysine deoxycolate agar (XLD)	Rappaport-Vassiliadis with soya; Muller-Kauffman tetrathionate/novobiocin	5	0
ISO 6579	Other; Xylose lysine deoxycolate agar (XLD)	Muller-Kauffman tetrathionate/novobiocin ; Rappaport-Vassiliadis with soya	6	0
ISO 6579	Other; Xylose lysine deoxycolate agar (XLD)	Rappaport-Vassiliadis with soya; Muller-Kauffman tetrathionate/novobiocin	5	0
ISO 6579	Xylose lysine deoxycolate agar (XLD); Brilliant green agar (BGA)	Muller-Kauffman tetrathionate/novobiocin ; Rappaport-Vassiliadis with soya	8	0
ISO 6579	Xylose lysine deoxycolate agar (XLD); Brilliant green agar (BGA)	Rappaport-Vassiliadis with soya; Muller-Kauffman tetrathionate/novobiocin	9	0
Other	Chromogenic agar - please state	Other	14	0

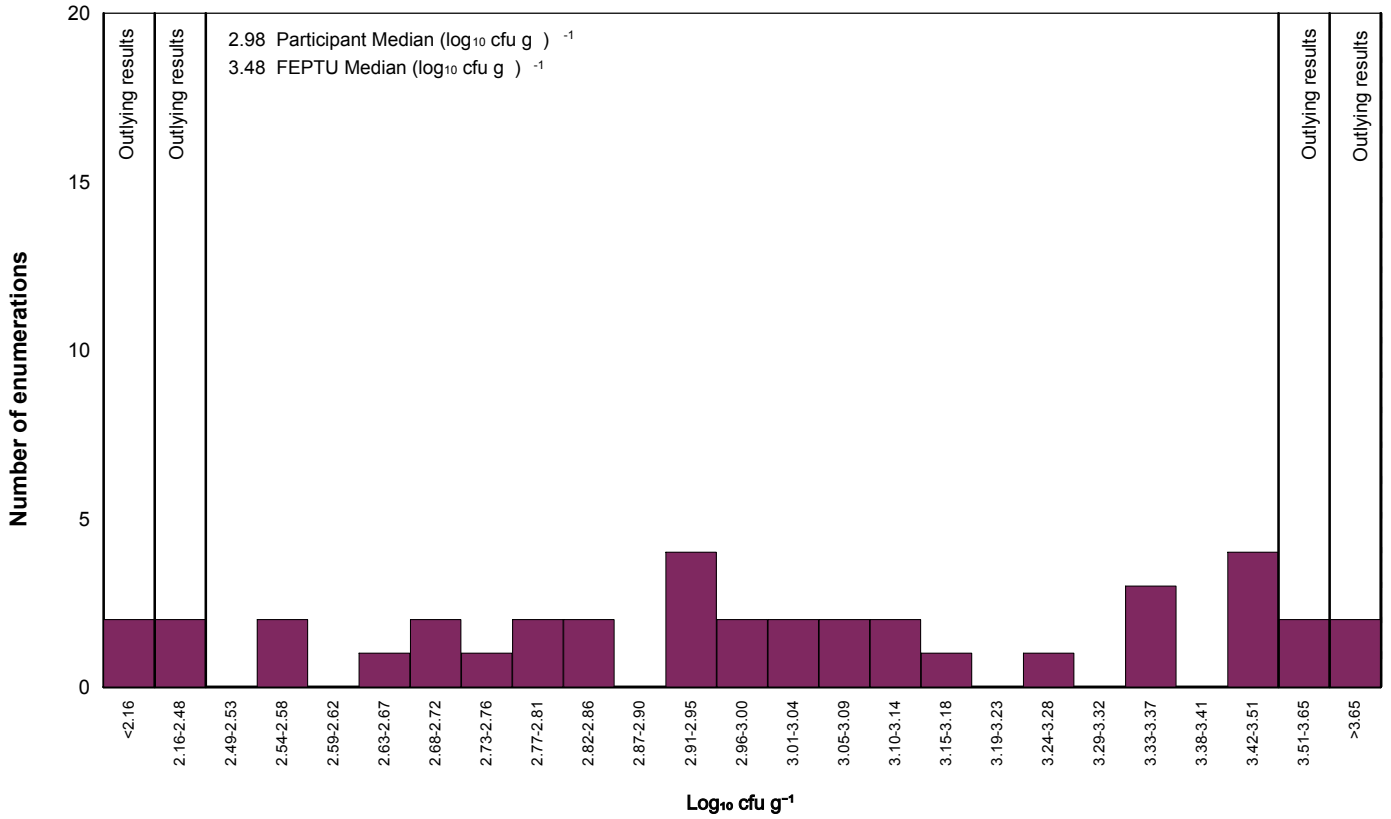
Sample S0625

Aerobic colony count Method	Aerobic colony count Media	Aerobic colony count Incubation	Count reported	Count censored values
	Other		0	0
ISO 4833-1	Petrifilm TM	30°C/48h	2	0
ISO 4833-1	Petrifilm TM	30°C/72h	1	0
ISO 4833-1	Petrifilm TM	Other	1	0
ISO 4833-1	Plate count agar	30°C/48h	8	1
ISO 4833-1	Plate count agar	30°C/48h; 30°C/72h	1	0
ISO 4833-1	Plate count agar	30°C/72h	77	1
ISO 4833-1	Plate count agar	30°C/72h; 30°C/48h	0	1
ISO 4833-1	Plate count agar	Other	1	0
ISO 4833-1; ISO 4833-2	Plate count agar	30°C/72h	0	0
ISO 4833-2	Milk plate count agar	Other	1	0
ISO 4833-2	Petrifilm TM; Plate count agar	30°C/48h	0	0
ISO 4833-2	Plate count agar	30°C/48h	13	0
ISO 4833-2	Plate count agar	30°C/72h	10	0
MPN - TEMPO	Other	30°C/48h	8	0
MPN - TEMPO	Other	Other	3	0
Other	Other	30°C/72h	1	0
Other	Other	37°C/24h	1	0
Other	Petrifilm TM	30°C/48h	1	0
Other	Petrifilm TM	Other	5	0
Other	Plate count agar	30°C/48h	3	0
Other	Plate count agar	30°C/72h	1	0
Other	Plate count agar	Other	8	0

Sample S0625

<i>Enterobacteriaceae</i> Method	<i>Enterobacteriaceae</i> Media	<i>Enterobacteriaceae</i> Incubation	Count reported	Count censored values
	VRBGA (Violet red bile glucose agar)	37°C/24h	1	0
ISO 21528-1		37°C/24h	0	0
ISO 21528-1	Petrifilm TM	Other	1	0
ISO 21528-1	VRBGA (Violet red bile glucose agar)	30°C/24h	1	0
ISO 21528-1	VRBGA (Violet red bile glucose agar)	37°C/24h	17	1
ISO 21528-2	Chromogenic agar - please state	37°C/24h	1	0
ISO 21528-2	Petrifilm TM	37°C/24h	2	0
ISO 21528-2	VRBGA (Violet red bile glucose agar)	30°C/24h	2	0
ISO 21528-2	VRBGA (Violet red bile glucose agar)	37°C/24h	83	2
ISO 21528-2	VRBGA (Violet red bile glucose agar)	Other	3	0
MPN - TEMPO	Other	37°C/24h	1	0
MPN - TEMPO	Other	Other	8	0
MPN - TEMPO	VRBGA (Violet red bile glucose agar)	37°C/24h	1	0
Other			0	0
Other	Chromogenic agar - please state	37°C/24h	5	0
Other	Other	37°C/24h	1	0
Other	Petrifilm TM	37°C/24h	3	0
Other	Petrifilm TM	Other	1	0
Other	VRBGA (Violet red bile glucose agar)	37°C/24h	7	0
Other	VRBGA (Violet red bile glucose agar)	Other	2	0

Campylobacter spp. enumeration reported by participants - Sample S0626



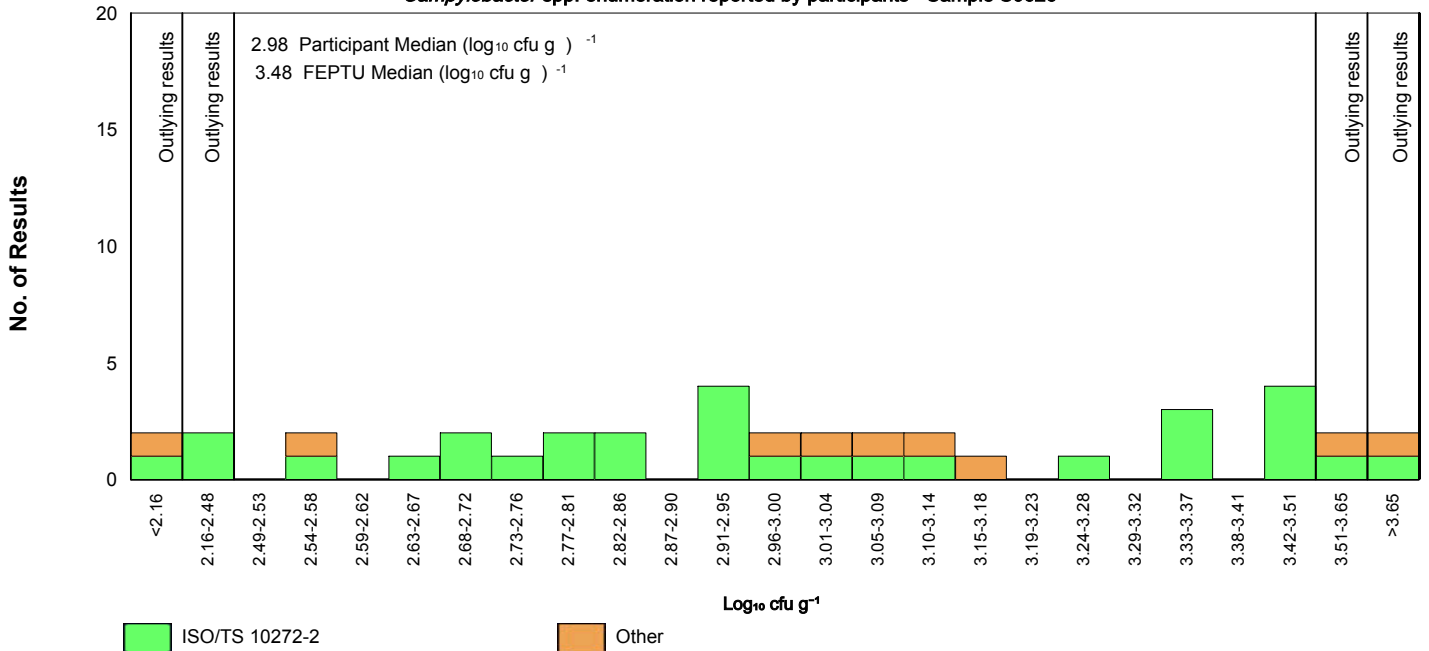
Method based presentation

S0626 : *Campylobacter* spp. enumeration

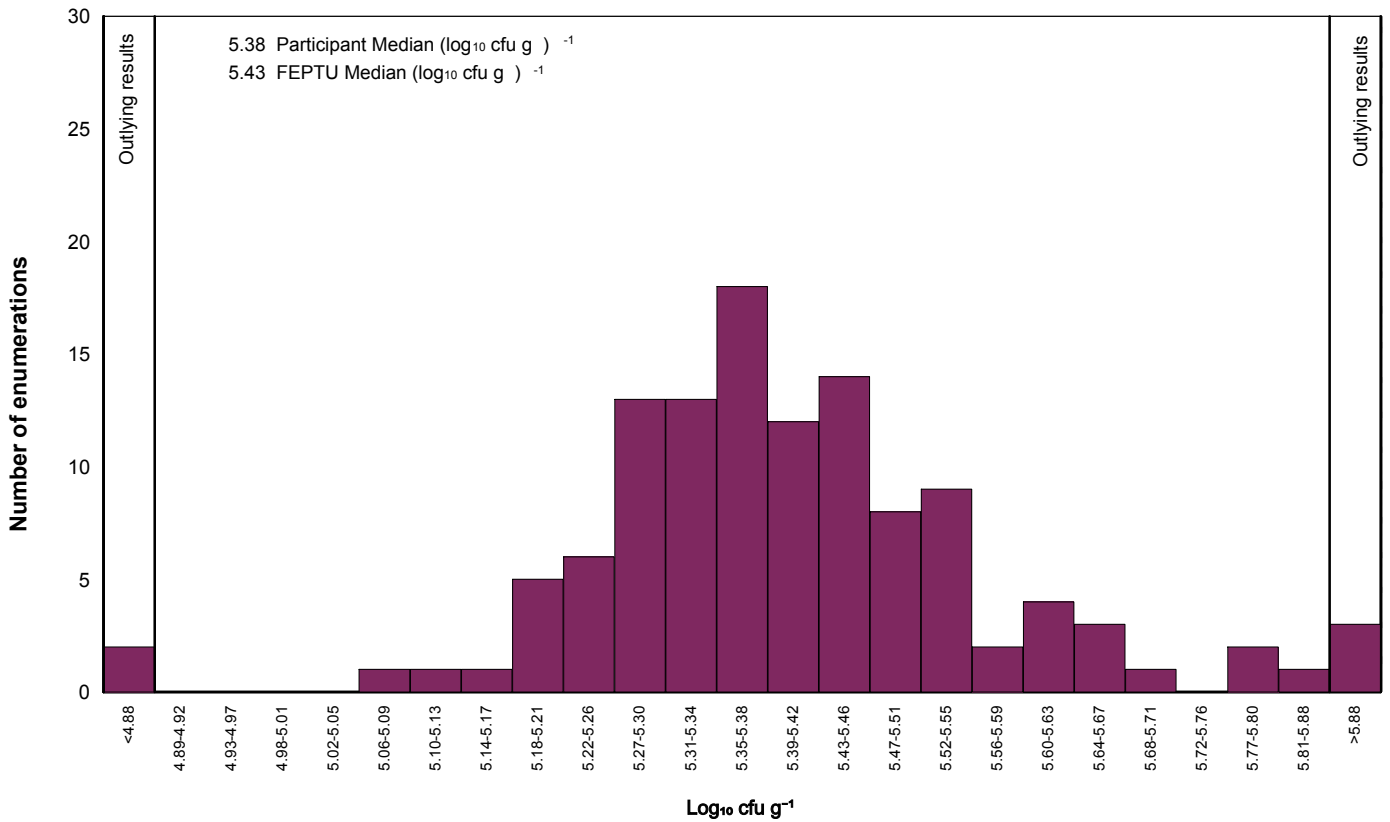
FEPTU Method: ISO/TS 10272-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (Log ₁₀ cfu g ⁻¹)	Robust S* (Log ₁₀ cfu g ⁻¹)	Range Reported (Log ₁₀ cfu g ⁻¹)
ISO/TS 10272-2	30	0	76	2.93	0.39	1.96 - 4.23
Other	9	1	23			-

Campylobacter spp. enumeration reported by participants - Sample S0626



Aerobic colony count reported by participants - Sample S0626



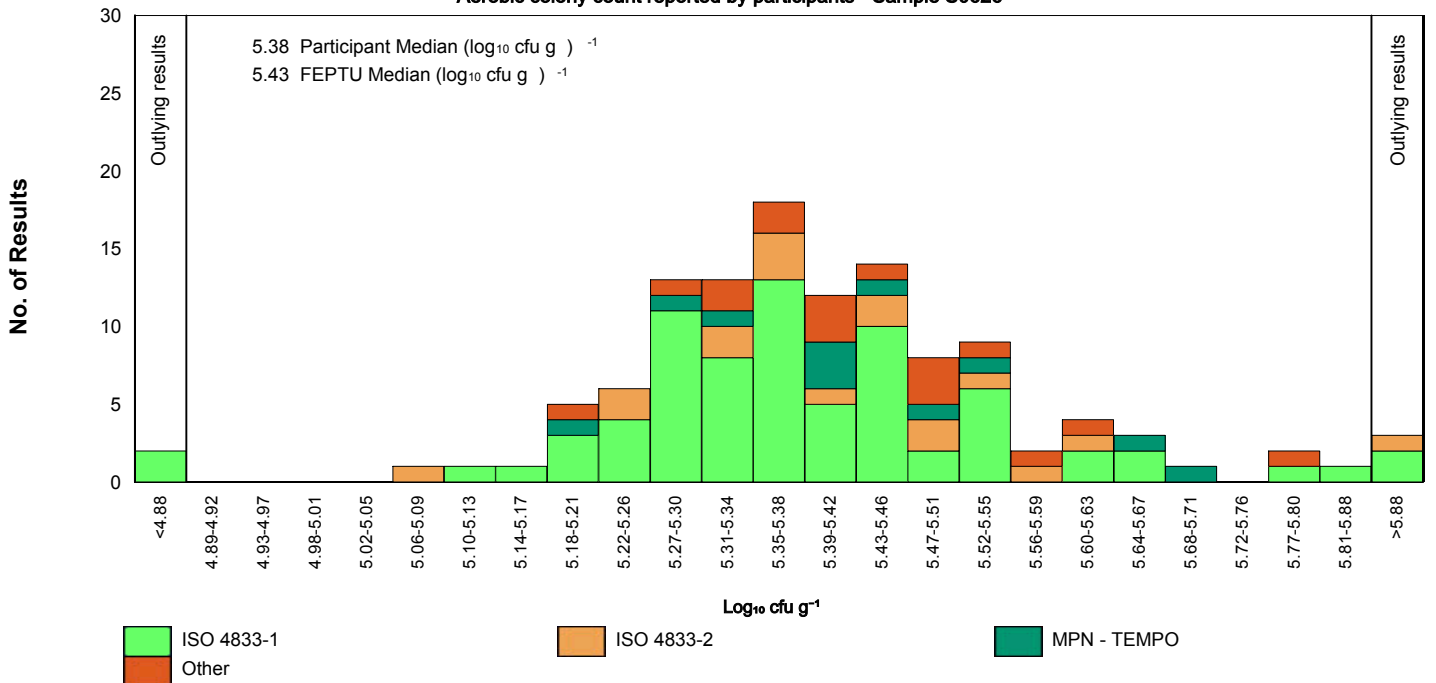
Method based presentation

S0626 : Aerobic colony count

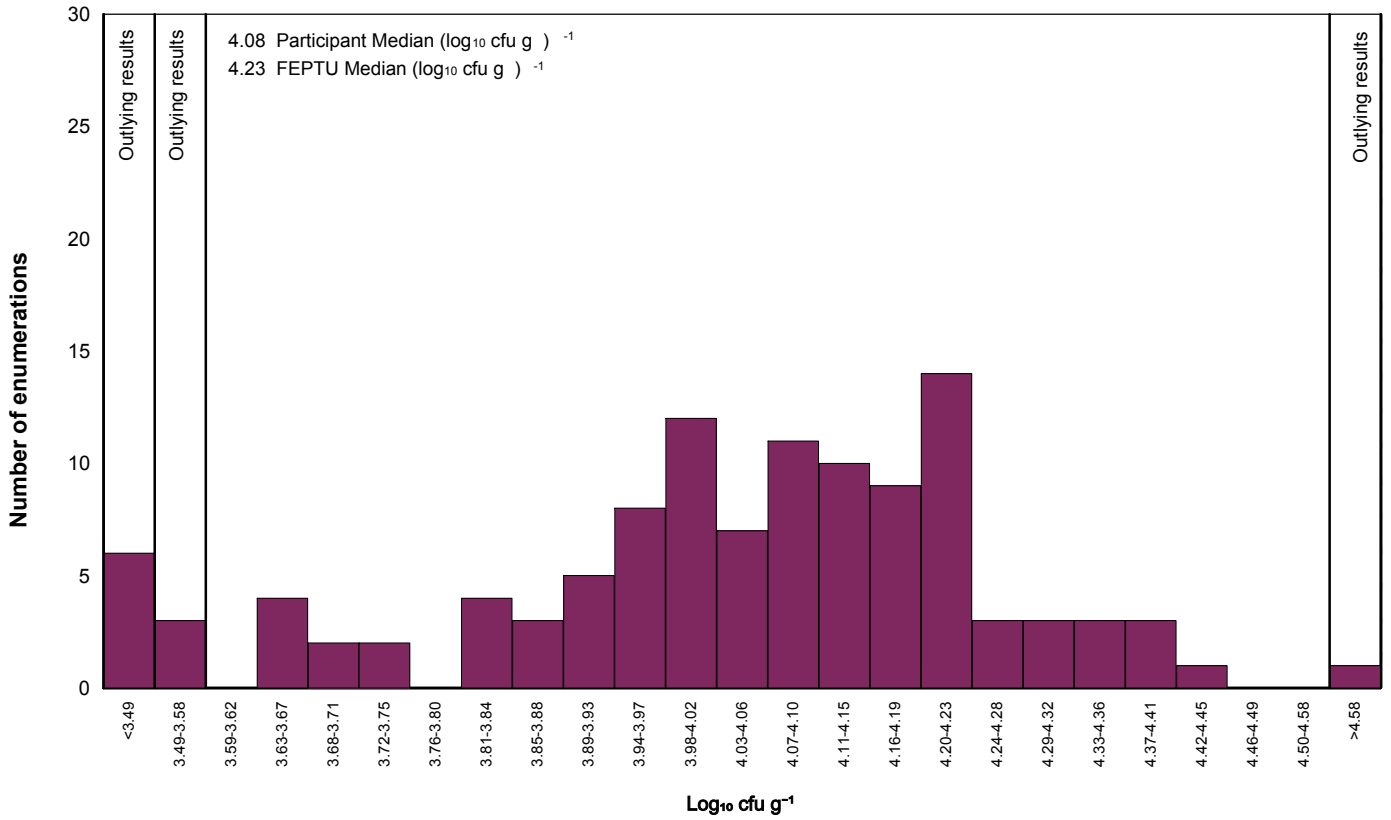
FEPTU Method: ISO 4833-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (\log_{10} cfu g ⁻¹)	Robust S* (\log_{10} cfu g ⁻¹)	Range Reported (\log_{10} cfu g ⁻¹)
ISO 4833-1	74	0	62	5.36	0.13	4.38 - 7.43
ISO 4833-2	17	0	14	5.40	0.15	5.06 - 6.36
MPN - TEMPO	11	0	9	5.40	0.13	5.18 - 5.69
Other	17	0	14	5.41	0.13	5.19 - 5.76

Aerobic colony count reported by participants - Sample S0626



Enterobacteriaceae reported by participants - Sample S0626



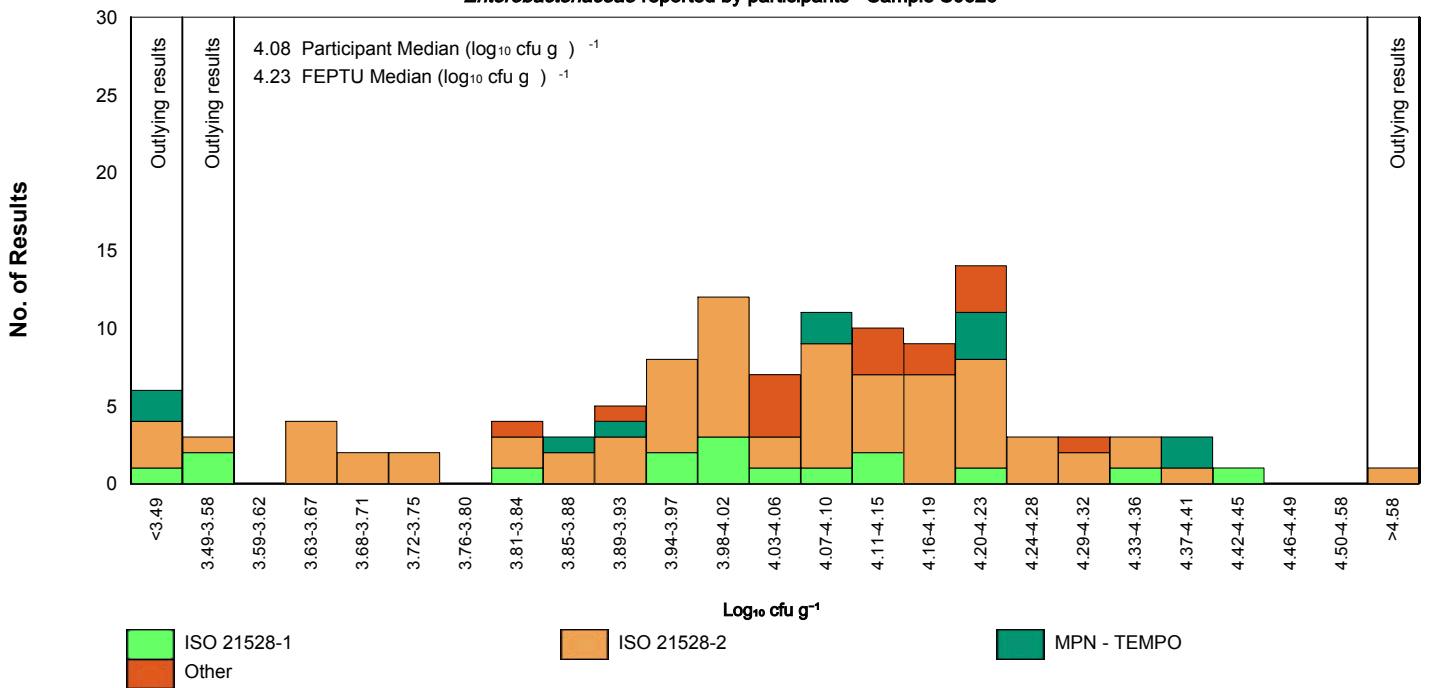
Method based presentation

S0626 : Enterobacteriaceae

FEPTU Method: ISO 21528-2

Method	Number of Results	Excluded Results	Percentage of the total	Median (\log_{10} cfu g ⁻¹)	Robust S* (\log_{10} cfu g ⁻¹)	Range Reported (\log_{10} cfu g ⁻¹)
ISO 21528-1	16	0	14	3.99	0.23	3.36 - 4.45
ISO 21528-2	72	1	63	4.06	0.19	3.19 - 5.07
MPN - TEMPO	11	0	9	4.08	0.29	2.04 - 4.40
Other	15	0	13	4.11	0.11	3.83 - 4.30

Enterobacteriaceae reported by participants - Sample S0626



Sample S0626

<i>Campylobacter</i> spp. Method	<i>Campylobacter</i> spp. Media	<i>Campylobacter</i> spp. Enrichment	No. Participants detected	No. Participants not detected
ISO 10272-1	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	18	1
ISO 10272-1	Other; Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	9	0
ISO 10272-1	Preston agar; Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)	Bolton Broth	5	0

Sample S0626

<i>Campylobacter</i> spp. enumeration Method	<i>Campylobacter</i> spp. enumeration Media	<i>Campylobacter</i> spp. enumeration Incubation	Count reported	Count censored values
	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		0	0
ISO/TS 10272-2	Chromogenic agar - please state		1	0
ISO/TS 10272-2	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		26	1
ISO/TS 10272-2	Other		2	0
ISO/TS 10272-2	Preston agar		1	0
Other	Chromogenic agar - please state		4	0
Other	Modified charcoal cefoperazone deoxycholate agar (mCCDA/CCDA)		4	0
Other	Other		1	1

Sample S0626

<i>E.coli</i> O157 Method	<i>E.coli</i> O157 Media	<i>E.coli</i> O157 Enrichment	No. Participants detected	No. Participants not detected
ISO 16654	Cefixime tellurite sorbitol MacConkey agar (CT-SMAC)	Modified Tryptone soya	8	1
ISO 16654	Cefixime tellurite sorbitol MacConkey agar (CT-SMAC); Chromogenic agar - please state	Modified Tryptone soya	7	0

Sample S0626

Aerobic colony count Method	Aerobic colony count Media	Aerobic colony count Incubation	Count reported	Count censored values
	Other		0	0
ISO 4833-1			0	0
ISO 4833-1	Petrifilm TM	30°C/48h	2	0
ISO 4833-1	Petrifilm TM	30°C/72h	1	0
ISO 4833-1	Petrifilm TM	Other	1	0
ISO 4833-1	Plate count agar	22°C/72h	1	0
ISO 4833-1	Plate count agar	30°C/48h	7	1
ISO 4833-1	Plate count agar	30°C/48h; 30°C/72h	1	1
ISO 4833-1	Plate count agar	30°C/72h	60	0
ISO 4833-1	Plate count agar	Other	1	0
ISO 4833-2	Milk plate count agar	Other	1	0
ISO 4833-2	Petrifilm TM; Plate count agar	30°C/48h	0	0
ISO 4833-2	Plate count agar	30°C/48h	11	0
ISO 4833-2	Plate count agar	30°C/72h	5	0
ISO 4833-2; ISO 4833-1	Plate count agar	30°C/72h	0	0
MPN - TEMPO	Other	30°C/48h	8	0
MPN - TEMPO	Other	Other	3	0
Other	Other	37°C/24h	1	0
Other	Petrifilm TM	30°C/48h	1	0
Other	Petrifilm TM	Other	5	0
Other	Plate count agar	30°C/48h	3	0
Other	Plate count agar	30°C/72h	1	0
Other	The data in FEPTU reports is confidential	Other		

Sample 30626

<i>Enterobacteriaceae</i> Method	<i>Enterobacteriaceae</i> Media	<i>Enterobacteriaceae</i> Incubation	Count reported	Count censored values
ISO 21528-1		37°C/24h	0	0
ISO 21528-1	Petrifilm TM	Other	1	0
ISO 21528-1	VRBGA (Violet red bile glucose agar)	30°C/24h	1	0
ISO 21528-1	VRBGA (Violet red bile glucose agar)	37°C/24h	14	1
ISO 21528-2	Chromogenic agar - please state	37°C/24h	1	0
ISO 21528-2	Petrifilm TM	37°C/24h	2	0
ISO 21528-2	VRBGA (Violet red bile glucose agar)	30°C/24h	2	0
ISO 21528-2	VRBGA (Violet red bile glucose agar)	37°C/24h	64	2
ISO 21528-2	VRBGA (Violet red bile glucose agar)	Other	3	0
MPN - TEMPO	Other	37°C/24h	1	0
MPN - TEMPO	Other	Other	9	0
MPN - TEMPO	VRBGA (Violet red bile glucose agar)	37°C/24h	1	0
Other			0	0
Other	Chromogenic agar - please state	37°C/24h	3	0
Other	Other	37°C/24h	1	0
Other	Petrifilm TM	37°C/24h	3	0
Other	Petrifilm TM	Other	1	0
Other	VRBGA (Violet red bile glucose agar)	37°C/24h	7	0

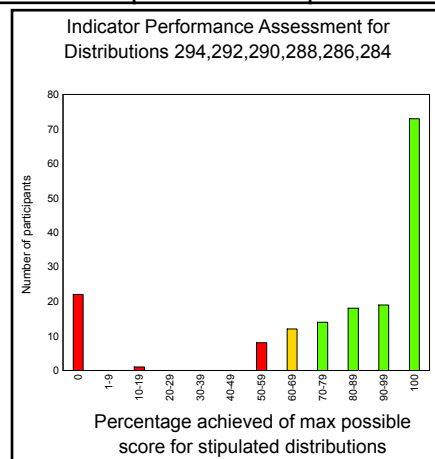
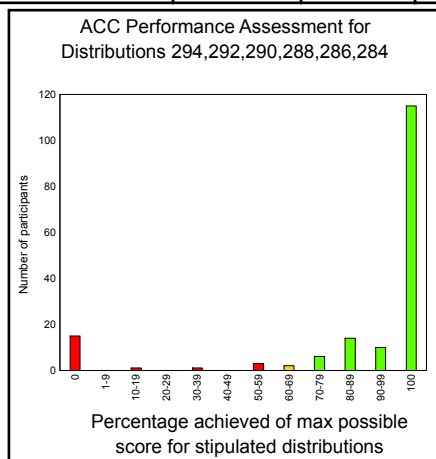
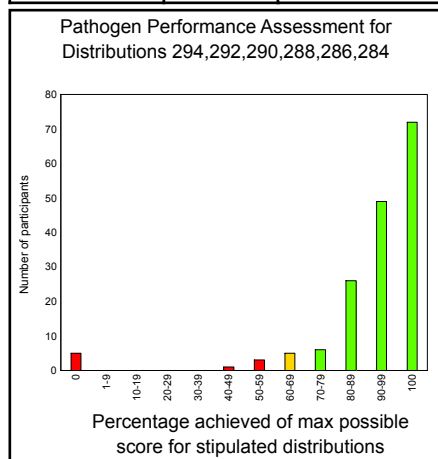
Performance Assessment Sheet

Participants are reminded that to take advantage of the performance assessment overtime tool provided in the reports they need to take part in more than one distribution a year.

Performance assessments are designed to identify laboratories with on-going problems with their examinations and are undertaken after every distribution. Scores are allocated to results reported for every sample to help assess participants' performance.

Cumulative scores are calculated for every participant, for all examination types, for the current and previous five distributions. Participants' cumulative scores for each of the examination types are compared with the maximum possible scores after every distribution.

Distribution	Sample	Examination	Your score	Your %	Sample	Examination	Your score	Your %
294	S0625	Pathogen			S0626	Pathogen		
	S0625	ACC			S0626	ACC		
	S0625	Indicator			S0626	Indicator		
292	S0621	Pathogen			S0622	Pathogen		
	S0621	ACC			S0622	ACC		
	S0621	Indicator			S0622	Indicator		
290	S0617	Pathogen			S0618	Pathogen		
	S0617	ACC			S0618	ACC		
	S0617	Indicator			S0618	Indicator		
288	S0613	Pathogen			S0614	Pathogen		
	S0613	ACC			S0614	ACC		
	S0613	Indicator			S0614	Indicator		
286	S0609	Pathogen			S0610	Pathogen		
	S0609	ACC			S0610	ACC		
	S0609	Indicator			S0610	Indicator		
284	S0605	Pathogen			S0606	Pathogen		
	S0605	ACC			S0606	ACC		
	S0605	Indicator			S0606	Indicator		



Performance Assessment Comment:

Laboratories that achieve less than 70% of the maximum possible score are likely to be experiencing significant problems with their examinations and are advised to

- refer to the relevant sample reports for specific comments
- refer to the website guidance documents: <https://www.gov.uk/government/collections/external-quality-assessment-ega-and-proficiency-testing-pt-for-food-water-and-environmental-microbiology>
- contact the organisers for advice.

General distribution comments:

Participants are reminded if you do not examine a specific parameter you must return your results as 'Not examined' as this impacts the overall scores awarded.

General comments on methods:

Participants that did not provide information on the method and testing conditions, their data is not included in the method graphs and tables. This information is useful; therefore participants are encouraged to complete these details.

Method based presentation tables for enumeration results

Participants are advised if less than 10 laboratories report an enumeration result for a method, no data is shown for the Median, Robust SD and the Range Reported.

Method, media and enrichment/incubation tables

Participants are asked to note:

- that for pathogen detection parameters, the data presented in the tables on the specific method used, is only shown when five or more laboratories have reported a result for that specific method
- that the count shown in the 'Count censored values' column includes data from those laboratories that reported:
 - a censored value
 - a result reported as detected or not detected
 - method data with no results reported.

Participants are reminded that the method data presented in this way has some limitations and seeks to identify trends in the results rather than assess specific method details.

Trend analysis

Plotting your PT results over a period of time can help to identify potential problems. Download the updated trend analysis spreadsheet one week after this report has been issued:

<https://www.gov.uk/government/publications/standard-scheme-trend-analysis>

