United Kingdom Variety Lists / Plant Breeders’ Rights Technical Protocol for Official Examination of Distinctness, Uniformity and Stability (DUS)

Wheat

Triticum aestivum L.

December 2022
Section A – General Information

1 Purpose
1.1 This Protocol sets out the procedures for conducting tests and assessments in relation to official examinations of DUS, maintenance of reference stocks and verification of VCU submissions of varieties of winter and spring Wheat entered for Variety Lists (VL) Trials and Plant Breeders' Rights (PBR).

2 Scope
2.1 These procedures apply to all varieties of Wheat. Special procedures and responsibilities for Genetically Modified (GM) varieties are set out in Sections A5 and A6.

2.2 Except where specified in this protocol or authorised by the Animal and Plant Health Agency (APHA) Varieties and Seeds, only Variety List candidates, Plant Breeders' Rights candidates, candidates for Foreign Authorities and the reference varieties may be incorporated in the DUS tests.

3 Responsibilities
3.1 The growing tests and assessments in this protocol are carried out under the responsibility of the Secretary of State for Environment, Food and Rural Affairs, Scottish Ministers, Welsh Ministers and the Minister for Agriculture, Environment and Rural Affairs in Northern Ireland (the National Authorities).

3.2 They are supervised, on behalf of the National Authorities, by officials of the Testing Authorities, that is, APHA, the Scottish Government (SG), the Department of Agriculture, Environment and Rural Affairs (DAERA) and the Welsh Government (WG).

3.3 This protocol is authorised by the Plant Variety and Seeds Committee (PVSC). It cannot be amended without their approval. Requests and suggestions for amendment of the protocol should be put in writing to APHA Plant Varieties and Seeds, either directly or via the Test Centre.

3.4 The procedures are administered by:

Plant Varieties and Seeds
Animal and Plant Health Agency
Eastbrook
Shaftesbury Road
Cambridge
CB2 8DR
Email: pvs.helpdesk@apha.gov.uk
3.5 Test Centre

The DUS growing tests and assessments in this protocol are co-ordinated and carried out by:

NIAB
Barn 1 Park Farm
Villa Road
Impington
Cambridge Tel No. 01223 342200
CB24 9NZ Fax No. 01223 277602

3.6 The Test Centre is responsible for providing the appropriate facilities.

4 Non-Compliance with the Protocol

4.1 Where the protocol uses the word “must” for any action then failure to carry out this action will result in non-compliance. Where non-compliance occurs or there are concerns regarding the validity of any data or tests this must be reported to APHA. Where this protocol uses the word “should” for any action this is the method to be followed unless there are clear reasons which can be justified by the Test Centre as technically sound.

5 Responsibility for GM Releases

5.1 GM Release Consent Holders are responsible for GM releases. All parties involved in DUS work operating under a GM Release Consent must adhere to the instructions of the Release Consent Holder, where necessary, to comply with the relevant consent conditions. Where DUS protocol non-compliance occurs, this must be reported to the consent holder and the Test Centre who will notify APHA.

6 Procedures for GM Varieties

6.1 Applicants intending to enter GM candidates must consult APHA, well in advance of their application, about specific requirements under GM regulations.

6.2 The Test Centre must ensure that no test or trial sites are planted with GM candidates and/or varieties until APHA has given the specific clearances.
# 7 Associated Documents

The following documents are associated with this protocol

<table>
<thead>
<tr>
<th>Reference</th>
<th>Title</th>
</tr>
</thead>
<tbody>
<tr>
<td>UPOV TG/1/3</td>
<td>General Introduction to the Examination of Distinctness, Uniformity and Stability and the Development of Harmonised Descriptions of New Varieties of Plants (09.04.2002)</td>
</tr>
<tr>
<td>UPOV TGP/8/4</td>
<td>Trial Design and Techniques Used in the Examination of Distinctness, Uniformity and Stability (01.11.2019).</td>
</tr>
<tr>
<td>UPOV TGP/9/2</td>
<td>Examining Distinctness (29.10.2015).</td>
</tr>
<tr>
<td>UPOV TGP/10/2</td>
<td>Examining Uniformity (01.11.2019).</td>
</tr>
<tr>
<td>GB and NI National Lists</td>
<td>The Seeds (National Lists of Varieties) Regulations 2001 (as amended) and The Seeds (Variety Lists) Regulations (Northern Ireland) 2020</td>
</tr>
</tbody>
</table>
Section B – Application Requirements

1 Purpose
1.1 The purpose of this section is to identify the specific requirements for Variety Lists and/or Plant Breeders’ Rights applications, as appropriate.

2 Scope
2.1 These procedures apply to all applications.

3 Responsibilities
3.1 The applicants are responsible for ensuring that these procedures are complied with.

4 Receipt of Applications
4.1 The latest date for receipt of applications for acceptance of a variety onto the Variety List or for Plant Breeders’ Rights, which is set administratively by APHA, is stated on the GOV website (https://www.gov.uk/national-lists-of-agricultural-and-vegetable-crops). Applications received after this date may be considered for inclusion in the current year’s test and trials on a case-by-case basis.

4.2 The procedures for the submission of Variety List and Plant Breeders’ Rights applications, Technical Questionnaires (TQs) and for payment of administration fees can be obtained from APHA PVS at the address shown in Section A or on the GOV.UK website (https://www.gov.uk/national-lists-of-agricultural-and-vegetable-crops).

4.3 Applicants should note in the TQ, submitted with the application, any additional characteristics which may require examinations that are listed in the DUS characteristics section D, 5.2 or 5.3 (an additional fee may be required).

4.4 In the case of hybrid varieties the TQ must include details of the nature of the hybrid and all progenitor lines.

5 Receipt of Seed
5.1 The latest date for receipt of seed is stated in the Seed Gazette and is set administratively by APHA. Seed submissions received after this date will normally be refused. Instructions for the delivery of seed will be made available to applicants by APHA (https://www.gov.uk/national-lists-of-agricultural-and-vegetable-crops).

6 Seed Quality Requirements
6.1 The seed must satisfy the quality requirements for Basic Seed as laid down in the seed marketing legislation of the Devolved Administrations.
6.2 The seed must not be chemically treated. Seed treatment, if required, will be undertaken by the Test Centre. The chemicals applied and rates of application will be determined by the Test Centre.

7 Seed Quantity

7.1 Year 1

<table>
<thead>
<tr>
<th>Type</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Conventional Type</td>
<td>2 kg* with 1000 seed weight given.</td>
</tr>
<tr>
<td>2. Other Types</td>
<td>Contact APHA</td>
</tr>
</tbody>
</table>

This can be submitted as 1.5 kg bulk seed and 0.5 kg selected seed samples for the 2021/22 field season. From 1st August 2022 onward a split submission will not be accepted - a single 2 kg submission should be submitted.

There is a separate submission of seed for VCU trials in Year 1.

7.2 Year 2 and Further Year Submissions

None for DUS

A sample of 200g of seed will be drawn from VCU submissions in Year 2 and any further years to authenticate the submission (see Section E5). Applicants should refer to Trial Procedures for Official Examination of Value for Cultivation and Use (VCU) for cereals for seed requirements and Section E4 dealing with replacement seed of a variety.

7.3 Shortfall in Seed Quantities

Where insufficient seed is available in the first instance a further stock must be supplied in the following year which will be authenticated against the original submission. An additional charge may be applied. This must be agreed in advanced with APHA and the test centre.

7.4 Hybrid Wheat

In the case of hybrid wheats where insufficient seed stocks of parent lines are available, a minimum of 500g of each line should be supplied in the first instance. Further stocks should be supplied in the following year, and these will be authenticated against the original submission for which an additional charge may be applied. Where components of hybrids are on the GB and NI Variety Lists or are already in test, or have UK PBR, seed need not be supplied unless specifically requested.
8 Labelling Requirements, Including Provisions for GM Varieties

8.1 Applicants must clearly label their seed with the following information:

- Applicant
- AFP number (if known)
- Breeder’s Reference number or name
- Type of Seed (Bulk or Selected, DUS only)
- Quantity of seed and thousand seed weight
- In the case of hybrids, whether it is a parental line

8.1 All packages of GM material must be labelled clearly as “GMO” or “Genetically Modified Organism”.
Section C – Growing Test Procedures

1 Purpose
1.1 The purpose of this section is to provide details of the procedures used in the growing tests for DUS analysis.

2 Scope
2.1 These procedures apply to all varieties of Wheat.

3 Responsibilities
3.1 The Test Centre is responsible for conducting these procedures.

3.2 The Test Centre will be responsible for ensuring that no material supplied to them is used for any other purpose than the conduct of these procedures or the release of reference samples for authorised purposes. (See Section E7).

4 Reference Varieties
4.1 The principles governing the selection of reference varieties are set out in Appendix 1.

4.2 Seed of reference varieties will be supplied by the Test Centre.

5 Design of Tests
5.1 The Test Centre is responsible for selecting a suitable site which should be on ground that has not normally had a Wheat crop in the previous three years but may be less where the risk of contamination is negligible.

5.2 Crop husbandry should follow best local practice for all operations and particularly as regards cultivation, drilling, fertiliser and spray application, use of irrigation and control of pests and diseases.

5.3 The minimum duration of tests should normally be two independent growing cycles. Additional growing cycles may be approved by the National List and Seeds Committee (NLSC).

5.4 From information given in the TQ the candidate variety may be grown in field grown plots and compared with varieties which are in the same classification for the following characters; seasonal type, ear colour, presence or absence of scurs or awns and hairiness on external surface of lower glume.
5.5 Plots are sown from the submitted seed in each year of test as follows:

<table>
<thead>
<tr>
<th>Number of plots:</th>
<th>2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Number of Plants Examined/Variety:</td>
<td>2000 (approx.)</td>
</tr>
<tr>
<td>No. of plants /sq.m</td>
<td>200-300</td>
</tr>
<tr>
<td>Time of sowing (winter wheat)</td>
<td>October - November</td>
</tr>
<tr>
<td>Time of sowing (spring wheat)</td>
<td>February - April</td>
</tr>
</tbody>
</table>

5.6 In the case where two samples have been submitted (ie selected seed and bulk seed), all testing is conducted on the selected seed sample. A sample of 100g will be withdrawn from the bulk seed and sown side by side to the selected seed for authentication in the first year of test.

5.7 In the case of winter varieties, an additional plot is sown in late April during the first year of tests to examine the uniformity of the vernalisation response (characteristic ‘Seasonal Type’):

<table>
<thead>
<tr>
<th>Number of plots:</th>
<th>1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Number of Plants Examined/Variety:</td>
<td>minimum 500 plants (approx.)</td>
</tr>
<tr>
<td>No. of plants /m²</td>
<td>200-300</td>
</tr>
<tr>
<td>Time of sowing</td>
<td>Late April</td>
</tr>
</tbody>
</table>

5.8 At the end of the first recording year, reference varieties which are most similar to the candidate varieties are identified and may be sown in the second year of test.

5.9 At the end of the second year of tests, candidate varieties that are still not distinct may be grown in additional direct comparison plots. This requires approval from APHA, and an additional charge will be made to the applicant.

6 Records and Recording

6.1 All records and plot data should be in a form determined and validated by the Test Centre.

6.2 Characters, recording details and instructions are given in Section D. Any variant and abnormal plant or plants resulting from an adverse reaction to husbandry practice are recorded but excluded from analysis.

6.3 In the first test cycle, characters, as indicated in Section D, are recorded on all varieties in test and the data analysed to assess uniformity of the candidate varieties and to determine the most similar reference varieties for each candidate. (For details see Section G).
6.4 In the second test cycle, characters, as indicated in Section D, are assessed on all varieties in test and the data analysed and, together with those from the first test cycle, used to assess distinctness and uniformity of the candidate varieties. (For details see Section G).

6.5 If the Test Centre notices unusual or novel characters in candidate varieties they may be noted and a photographic record taken.

7 Communications with the Applicant

7.1 The Test Centre will notify the applicant or the agent of any DUS problems at the earliest practical opportunity as they arise during the growing season. All such notifications must be copied to APHA.

7.2 In the case of distinctness problems, if confidentiality considerations allow, the applicant should be informed which variety is similar and be invited to submit any information which may help to distinguish them.

7.3 If DUS problems arise, applicants will be invited to visit the DUS tests by arrangement so that the material can be examined (if appropriate) and discussions held with the Test Centre.

7.4 After each recording season the results are summarised and reported to the applicant and APHA by the Test Centre.
Section D – Summary of DUS Characteristics to be Assessed, Method of Assessment and Standards Applied

1 Purpose
1.1 The purpose of this section is to summarise the characteristics to be assessed.

2 Scope
2.1 This section summarises characteristics, states of expression, method of observation and standards required for DUS assessment.

3 Responsibilities
3.1 The Test Centre is responsible for co-ordinating the procedures in this summary of characteristics.

4 Organisation
4.1 The minimum duration of tests to assess characteristics is normally two independent growing cycles. Shorter durations may be applied for assessment of additional characteristics. Additional growing periods must be approved by the NLSC.

5 DUS Characteristics to be Assessed
5.1 Routine Characteristics
The following table summarises the DUS characteristics to be routinely examined.

Note:
* a characteristic which must be examined according to the UPOV Guidelines.
G a grouping characteristic

Type of observation of characteristics:
MG Single measurement of a group of plants or parts of plants
MS Measurement of a number of individual plants or parts of plants
VG Visual assessment by a single observation of a group of plants or parts of plants
VS Visual assessment by observation of individual plants or parts of plants
Number of plants or sample size for assessment

A  Sample size of 100 (See Section G 6.6)
B  Sample size of 2000
### 5.1 Wheat Characteristics Routinely Recorded in DUS Tests

<table>
<thead>
<tr>
<th>UPOV TG/3/12</th>
<th>Character</th>
<th>Material examined</th>
<th>Number of plants or sample size for assessment</th>
<th>Method of assessment and recording</th>
<th>States of expression</th>
<th>D Method and Minimum Distance required</th>
<th>U Method: standard applied</th>
</tr>
</thead>
<tbody>
<tr>
<td>1G</td>
<td>Seed: colour</td>
<td>Submitted seed</td>
<td>A</td>
<td>VG</td>
<td>1 white, 2 reddish, 3 purple, 4 bluish</td>
<td>1 state</td>
<td>1% @95%</td>
</tr>
<tr>
<td>2</td>
<td>Seed: colouration with phenol</td>
<td>Submitted seed</td>
<td>A</td>
<td>VG</td>
<td>1 absent or very light, 3 light, 5 medium, 7 dark, 9 very dark</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>3</td>
<td>Coleoptile: anthocyanin colouration</td>
<td>Submitted seed</td>
<td>A</td>
<td>VG</td>
<td>1 absent or very weak, 3 weak, 5 medium, 7 strong, 9 very strong</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>4*</td>
<td>Plant: growth habit</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 erect, 3 semi-erect, 5 intermediate, 7 semi-prostrate, 9 prostrate</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>5</td>
<td>Plants: frequency of plants with recurved flag leaves</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very low, 3 low, 5 medium, 7 high, 9 very high</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>6</td>
<td>Flag leaf: anthocyanin colouration of auricles</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very weak, 2 medium, 3 strong</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>7*</td>
<td>Time of ear emergence: (first spikelet visible on 50% of ears)</td>
<td>Field grown plot</td>
<td>B</td>
<td>MG</td>
<td>1 very early, 3 early, 5 medium, 7 late, 9 very late</td>
<td>2 days</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>UPOV TG/3/12</td>
<td>Character</td>
<td>Material examined</td>
<td>Number of plants or sample size for assessment</td>
<td>Method of assessment and recording</td>
<td>States of expression</td>
<td>D Method and Minimum Distance required</td>
<td>U Method: standard applied</td>
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<tr>
<td>8*</td>
<td>Flag leaf: glaucosity of sheath</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very weak 3 weak 5 medium 7 strong 9 very strong</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>9</td>
<td>Flag leaf: glaucosity of blade (lower side)</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very weak 3 weak 5 medium 7 strong 9 very strong</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>10*</td>
<td>Ear: glaucosity</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very weak 3 weak 5 medium 7 strong 9 very strong</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>11</td>
<td>Culm: glaucosity of neck</td>
<td>Field grown plot</td>
<td>B</td>
<td>VG</td>
<td>1 absent or very weak 3 weak 5 medium 7 strong 9 very strong</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>12G*</td>
<td>Lower glume: hairiness on external surface</td>
<td>Field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 absent 9 present</td>
<td>1 state</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>13*</td>
<td>Plant: length (stem, ear, awns and scurs)</td>
<td>Field grown plot</td>
<td>B</td>
<td>MG</td>
<td>1 very short 3 short 5 medium 7 long 9 very long</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>14*</td>
<td>Straw: pith in cross section (half way between base of ear and stem node below)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 thin 2 medium 3 thick or filled</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>15*</td>
<td>Ear: density</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 very lax 3 lax 5 medium 7 dense 9 very dense</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>UPOV TG/3/12</td>
<td>Character</td>
<td>Material examined</td>
<td>Number of plants or sample size for assessment</td>
<td>Method of assessment and recording</td>
<td>States of expression</td>
<td>D Method and Minimum Distance required</td>
<td>U Method: standard applied</td>
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<tr>
<td>16</td>
<td>Ear: length (excluding awns and scurs)</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 very short 3 short 5 medium 7 long 9 very long</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>17G*</td>
<td>Ear: scurs or awns</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 both absent 2 scurs present 3 awns present</td>
<td>1 state</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>18*</td>
<td>Ear: length of scurs or awns at tip of ear</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 very short 3 short 5 medium 7 long 9 very long</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>19G*</td>
<td>Ear: colour</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 white 2 coloured (red)</td>
<td>1 state</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>20</td>
<td>Ear: shape in profile</td>
<td>Samples from field grown plot</td>
<td>A for distinctness B for uniformity</td>
<td>VG</td>
<td>1 tapering 2 parallel sided 3 slightly clavate 4 strongly clavate 5 fusiform</td>
<td>2 states</td>
<td>0.3% @95%</td>
</tr>
<tr>
<td>21</td>
<td>Apical rachis segment: area of hairiness on convex surface</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 absent or very small 3 small 5 medium 7 large 9 very large</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>22</td>
<td>Lower glume: shoulder width (spikelet in mid-third of ear)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 very narrow 3 narrow 5 medium 7 broad 9 very broad</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>23</td>
<td>Lower glume: shoulder shape (spikelet in mid-third of ear)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 strongly sloping 3 slightly sloping 5 horizontal 7 slightly elevated 9 strongly elevated</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>UPOV TG/3/12</td>
<td>Character</td>
<td>Material examined</td>
<td>Number of plants or sample size for assessment</td>
<td>Method of assessment and recording</td>
<td>States of expression</td>
<td>D Method and Minimum Distance required</td>
<td>U Method: standard applied</td>
</tr>
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</tr>
<tr>
<td>24</td>
<td>Lower glume: length of beak (spikelet in mid-third of ear)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 very short 3 short 5 medium 7 long 9 very long</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>25*</td>
<td>Lower glume: shape of beak (spikelet in mid-third of ear)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 straight 3 slightly curved 5 moderately curved 7 strongly curved 9 geniculate</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>26</td>
<td>Lower glume: area of hairiness on internal surface (spikelet in mid-third of ear)</td>
<td>Samples from field grown plot</td>
<td>A</td>
<td>VG</td>
<td>1 very small 3 medium 5 very large</td>
<td>2 states</td>
<td>1% @95%</td>
</tr>
<tr>
<td>27G*</td>
<td>Seasonal type</td>
<td>Field grown plot</td>
<td>Winter and alternative types: 750 plant plot test sown in late spring Spring types: TQ declaration</td>
<td>VG</td>
<td>1 winter type 2 alternative type 3 spring type</td>
<td>1 state</td>
<td>1% @95%</td>
</tr>
</tbody>
</table>
5.2 Special Category Characteristics

These characters should only be used as a complement to confirm other morphological or physiological differences.

<table>
<thead>
<tr>
<th>UPOV TG/3/12</th>
<th>Character</th>
<th>Material examined</th>
<th>Number of plants or sample size for assessment</th>
<th>Method of assessment and recording</th>
<th>States of expression</th>
<th>D Method and Minimum Distance required</th>
<th>U Method: standard applied</th>
</tr>
</thead>
<tbody>
<tr>
<td>28</td>
<td>Glutenin composition: allele expression at locus Glu-A1</td>
<td>Submitted seed</td>
<td>20 grains for Distinctness 100 grains for Uniformity</td>
<td>Visual score</td>
<td>1 band 1 2 band 2 3 no band</td>
<td>1 state</td>
<td>1% @95% see note below</td>
</tr>
<tr>
<td>29</td>
<td>Glutenin composition: allele expression at locus Glu-B1</td>
<td>Submitted seed</td>
<td>20 grains for Distinctness 100 grains for Uniformity</td>
<td>Visual score</td>
<td>1 bands 6 + 8 2 bands 7 + 8 3 bands 7 + 9 4 band 7 (or 7 + 9 in the presence of bands 5 + 10 of char 30) 5 bands 13 + 16 6 bands 14 + 15 7 bands 17 + 18 8 band 20 9 bands 6.1 + 22</td>
<td>1 state</td>
<td>1% @95% see note below</td>
</tr>
<tr>
<td>30</td>
<td>Glutenin composition: allele expression at locus Glu-D1</td>
<td>Submitted seed</td>
<td>20 grains for Distinctness 100 grains for Uniformity</td>
<td>Visual score</td>
<td>1 bands 2+ 12 2 bands 3 + 12 3 bands 4 + 12 4 bands 5 + 10</td>
<td>1 state</td>
<td>1% @95% see note below</td>
</tr>
</tbody>
</table>

Note – allowance is made for the presence of biotypes
5.3 Additional Approved Characters
The following table summarises the additional characteristics which have been approved by the NLSC for Wheat.

<table>
<thead>
<tr>
<th>Type of expression</th>
<th>Characteristic</th>
<th>Growth Stage</th>
<th>Method of observation</th>
<th>States of expression</th>
<th>Example varieties</th>
<th>Note</th>
</tr>
</thead>
<tbody>
<tr>
<td>Quantitative</td>
<td>Lower lemma: beak shape</td>
<td>80-92</td>
<td>Visual score</td>
<td>Straight, Slightly curved, Moderately curved, Strongly curved, Geniculate</td>
<td>Soissons, Slejner, Sideral, Parade, Tara</td>
<td>1, 3, 5, 7, 9</td>
</tr>
<tr>
<td>Quantitative</td>
<td>Lower glume: external surface roughness</td>
<td>80-92</td>
<td>Visual score</td>
<td>Completely smooth, Slightly rough, Rough, Entire surface rough</td>
<td>Robigus, Gladiator, Portland</td>
<td>3, 5, 7, 9</td>
</tr>
</tbody>
</table>

5.4 New Additional DUS Characteristics
Applicants can suggest new characters on the TQ for testing DUS or after notification by the DUS Test Centre of distinctness problems (for procedures see Section F).
Section E – Reference Seed Stock Maintenance and VCU Seed Stock Authentication Procedures

1 Purpose
1.1 This section sets out the procedures for reference seed stock maintenance and VCU seed stock authentication (if applicable).

2 Scope
2.1 These procedures apply to all reference collection varieties and VCU seed submissions where the VCU seed has not been taken from the same bulk as the seed used for the DUS test.

3 Responsibilities
3.1 The Test Centre is responsible for conducting these procedures.

4 Procedures for Reference Seed Stock Maintenance
4.1 The DUS seed sample submitted with the successful or pending application is considered to be the definitive seed of the variety. In the case of a ‘selected seed’ sample and a ‘bulk seed’ sample being submitted, the ‘selected seed’ sample will be considered to be the definitive sample until the bulk sample has been authenticated (Section C). Subject to meeting the required quality standards (see Section B) the seed is dried and placed in storage under controlled and monitored refrigerated conditions as part of the official reference collection.

4.2 If during the normal tests there is any evidence that seed is deteriorating in storage, or that stocks are under 500 g, a request will be made to the maintainer asking for replacement seed (2 kg) of the variety. This replacement seed must be authenticated against the definitive seed.

4.3 Plots will be established from any replacement reference seed sample to be authenticated and compared visually with the definitive seed over a maximum of two recording seasons. Plots must be examined through all the growth stages from early growth habit to full harvest ripeness. If the new seed sample cannot be visually distinguished from the reference seed, it will be accepted as representing the variety. It will then be considered as the definitive seed and substituted for the existing definitive seed in the reference collection.
4.4 In the event of the replacement sample not meeting the required acceptance standards, an additional replacement sample will be requested. If the additional replacement sample does not meet the acceptance standard set out in 4.3, the variety will be deleted from the reference collection.

5 Procedures for VCU Seed Stock Authentication

5.1 A representative sub-sample of seven grains from the VCU seed submission are compared to a representative sub-sample of seven grains from the DUS seed submission (definitive seed) by electrophoresis using the SDS PAGE method. If the VCU seed sample tested by the electrophoresis method matches the DUS seed, it will be considered to represent the variety.

5.2 If the VCU seed sample does not match the DUS seed sample a further electrophoresis test will be carried out. If the VCU seed sample still does not match the DUS seed, side-by-side field plots of the two samples will be established and compared visually from early growth habit to full harvest ripeness.

5.3 If the VCU plot does not differ from the DUS plot in the comparison of field sown plots the VCU seed will be considered to represent the variety.

5.4 If the VCU plot can be visually distinguished from the definitive stock in the authentication plots then it will not be accepted as representing the variety.

6 Procedures for the Inclusion of New Common Knowledge Varieties into the Reference Collection

6.1 When a new variety enters into common knowledge such that it must be included in the reference collection, a request will be sent by the Test Centre to the Testing Authority which has added this variety to its Variety List for the supply of at least 200 g of seed of the definitive sample. This seed will then be used to validate a larger sample of seed from the breeder or the VCU seed sample.

7 Release of Reference Samples for Authorised Purposes

7.1 A maximum of 200 g of seed of reference samples can be supplied by the Test Centre, on request, to UK and UPOV, DUS Testing Authorities and UK and OECD Seed Certification Agencies. The recipient will be notified in writing that this material, or any material derived from it, must not be supplied to a Third party or used for any other purpose than as a reference for official DUS testing or seed certification.

7.2 Provision of reference samples, other than in 7.1, to any other parties must be authorised by APHA.
Section F – Procedures for Assessment of New Additional DUS Characters

1 Purpose
1.1 This Section sets out the procedures for assessment of new additional DUS characters for varieties of Wheat entered for Variety List trials and/or PBR.

2 Scope
2.1 These procedures apply to applications where additional DUS characteristics, which have not been previously approved by the NLSC, are requested for use in the examination of DUS.

3 Responsibilities
3.1 The Test Centre is responsible for communicating with the applicant to produce a proposed procedure for the conduct of new tests. This procedure must ensure that Distinctness, Uniformity and Stability requirements will be met.

3.2 All new additional characteristics must be authorised by the NLSC.

4 Reference Varieties
4.1 The reference varieties will include only those varieties from which the candidate variety is not distinct, as well as other varieties for control purposes.

4.2 Seed of reference varieties will be supplied by the Test Centre.

5 Procedures
5.1 Details of the proposed special test or assessments will be submitted to the NLSC to consider the feasibility of setting up a test acceptable to the UK Authorities. The applicant will be advised by APHA of arrangements and costs.

5.2 The NLSC will consider the results of the commissioned test or trial when reaching its recommendation on the granting of Plant Breeders’ Rights and/or Variety Listing.

5.3 Where the test for a character is approved by the NLSC it should be subsequently listed in Section D5.1, D5.2 or D5.3 as appropriate.
Section G – Procedures for DUS Decisions

1 Purpose
1.1 This section sets out the standards used to assess distinctness, uniformity and stability of varieties of Wheat.

2 Scope
2.1 These procedures apply to all varieties of Wheat entered for Variety Listing and/or Plant Breeders’ Rights tests and those being tested for Foreign Authorities.

3 Responsibilities
3.1 The Test Centre is responsible for applying the criteria for DUS, set out in this procedure.
3.2 The Test Centre is responsible for producing the DUS report in accordance with these procedures and for ensuring that they are in accordance with UPOV guidelines.

4 Reference Varieties
4.1 Appendix I sets out which varieties are considered as reference varieties for these procedures.

5 Distinctness
5.1 In accordance with associated document UPOV TG/1/3, varieties can be considered distinct where they have a different expression in a grouping character e.g. seasonal type; ear colour; presence or absence of awns; hairiness on external surface of lower glume.
5.2 The standard applied for distinctness over two years of test is a clear difference of one or two states in the expression of a characteristic in accordance with the table of characteristics given above in Section D.
5.3 Where varieties are grown in close proximity under the same conditions, and a direct comparison can be made, distinctness can be determined on the basis of visual observation. Characters are recorded using notes to represent states of expression (See Section D). In these circumstances, the basis for distinctness will be clearly recorded.
6 Uniformity

6.1 Uniformity is assessed for all characteristics used to establish Distinctness.

6.2 Uniformity is based on the assessment of off-types (variants) for visually observed characters.

6.3 The assessment of off-types is undertaken in both test cycles and the total should not exceed the number allowed using the population standards detailed below. Care is taken to ensure that the plants that are counted are not the result of any non-genetic factors such as environment, pest and disease.

6.4 In a sample size of 2,000 (characters marked as “B” in Section D), a population standard of 0.3% and an acceptance probability of at least 95% should be applied. For example, in a sample of 2000 plants, 10 off-types are allowed.

6.5 In a sample size of 100 (characters marked as “A” in Section D), a population standard of 1% and an acceptance probability of at least 95% should be applied. For example, in a sample of 100 plants or parts of plants, 3 off-types are allowed.

6.6 For characters marked as “A” in Section D, the assessment of uniformity can be carried out in two stages (with the exception of characteristics 2 and 3). In the first stage, 20 plants or parts of plants are examined. If no off types are observed the variety is declared uniform. If more than three off-types are found the variety is declared not to be uniform. If one to three off-types are observed, a further 80 plants or parts of plants should be examined.

6.7 Resubmissions

For all varieties, except hybrid varieties, a resubmission of plant material may be allowed for the second growing cycle if in the first growing cycle the number of off-types did not exceed 18 plants in a sample size of 2000 plants (population standard of 0.6% with an acceptance probability of at least 95%) or 9 plants or parts of plants in a sample size of 100 (population standard of 5% with and acceptance probability of at least 95%).

6.8 Hybrids

For the assessment of uniformity of hybrid varieties, a population standard of 10% and an acceptance probability of at least 95% should be applied. In case of characteristics indicated by “B”, the sample size for the assessment of uniformity may be reduced to 200 plants. In case of a sample size of 200 plants, 27 off-types are allowed. In case of a sample size of 100 plants or parts of plants, 15 off-types are allowed.
7 Stability

7.1 A variety is considered sufficiently stable when there is no evidence to indicate that it lacks uniformity or fails to conform to the essential characteristics of its description in different seed submissions or in different tests.

7.2 Hybrids may be considered to lack stability if there is evidence that their parental lines lack uniformity or fail to conform to the essential characteristics of their description.

7.3 For 3-way hybrids with segregating parental lines, the production and maintenance schemes of all parental lines must indicate that the final hybrid (candidate) can, in terms of its genetic constitution, be consistently reproduced in each cycle of propagation.

8 DUS Report and Variety Description

8.1 Upon completion of the DUS examination the DUS Summary report will be submitted to APHA by the specified date and will be discussed at the relevant DUS Test Centre Meeting. This report will specify all non-routine characteristics used for establishing distinctness.

8.2 The final DUS report, including the full variety description for positive reports, will be submitted to APHA by the specified date.
Appendix 1 – Reference Collection Varieties

1 Variety Listing and Plant Breeders’ Rights

1.1 The DUS reference collection, for any given category of plant variety comprises the following at the time when the application for the candidate is made:

1.2 All other candidate varieties already in DUS test in the UK or entering testing at the same time as the candidate.

1.3 All varieties with the UK PBR.

1.4 All varieties on the OECD variety list that are listed by countries with comparable climatic conditions to the UK.

1.5 All varieties protected under National PBR (UPOV contracting parties) with comparable climatic conditions to the UK.

1.6 Any varieties nominated by the applicant as being comparable i.e., known to be similar.

1.7 Any other varieties considered to be comparable i.e., known to be similar by the appropriate Test Centre or DUS Centre Group.

1.8 Other available comparable varieties in common knowledge.