

COMMITTEE ON CARCINOGENICITY OF CHEMICALS IN FOOD, CONSUMER PRODUCTS AND THE ENVIRONMENT (COC)

Second draft updated Guidance Statement (G04): The Use of Biomarkers in Carcinogenic Risk Assessment

1. The COC has periodically published guidelines for the evaluation of chemicals for carcinogenicity. The first guidance was published in 1982 and has undergone several updates since then to reflect advances in development and validation of methods for assessing risk of chemical carcinogenicity.
2. These updates included the separation of the overall guidance into individual documents during 2012 – 2014 to allow faster revisions to be made in the case of rapidly developing areas. This included a separate document addressing the use of biomarkers in carcinogenic risk assessment (G04).
3. Guidance statement G04 was last updated in 2018 and, as part of the rolling review process for COC documents, is being considered for update again. A first draft updated version of G04 was presented to COC in March 2021 (CC/2021/02).
4. The paper presented here is a second draft of the updated guidance statement, amended in line with discussions at the meeting in March 2021.

Questions for the Committee

5. Members are asked to:
 - i. Comment on the proposed updates presented in the second draft updated statement.
 - ii. Highlight any updates or new areas that are not currently covered.
 - iii. Comment on whether the Committee summary is appropriate.

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Committee on Carcinogenicity of Chemicals in Food, Consumer Products and the Environment (COC)

COC Guidance Statement G04 – version 1.2 draft 0.b

The Use of Biomarkers in Carcinogenic Risk Assessment

<https://www.gov.uk/government/groups/committee-on-carcinogenicity-of-chemicals-in-food-consumer-products-and-the-environment-coc>

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COC Guidance Statement G04 v1.2 draft 0.b

COMMITTEE ON CARCINOGENICITY OF CHEMICALS IN FOOD, CONSUMER PRODUCTS AND THE ENVIRONMENT

The Use of Biomarkers in Carcinogenic Risk Assessment

Introduction

1. The focus of this guidance statement are biomarkers of toxicological relevance, defined as “any substance, structure or process that can be measured in an organism, related to a specific exposure or effect or which can influence the incidence of the effect” (Choi et al., 2015). Biomarkers can provide valuable information to aid chemical risk assessment processes and are used as investigative tools in both animal and human studies which aim to evaluate carcinogenic hazards and risk.

2. For the purposes of this document, biomarkers will be broadly characterised as those of *exposure* and those of *effect*, although the distinction between these two is not always clear-cut. Biomarkers in the context of carcinogenicity can mean proof of exposure to a carcinogen, detection of a reaction product or an indication that a preliminary genotoxic event or actual DNA damage has occurred. Other types of biomarkers exist, for example biomarkers of susceptibility, which were initially introduced as interpretative aids to epidemiological investigations of chemically-induced carcinogenesis.

- *Biomarkers of exposure* - “an exogenous substance or its metabolite or the product of an interaction between a xenobiotic agent and some target molecule or cell that is measured in a compartment within an organism” (Laideira and Viegas, 2016). Biomarkers of exposure are further divided into those reflecting ‘internal dose’ and those reflecting ‘effective dose’. The concentration of a chemical (or metabolite) in blood following exposure is a basic measure of the internal dose, indicating the likely level of chemical (or metabolite) at the target site. The effective dose is a more accurate measurement of the exposure levels associated with the target molecule, structure or cell itself (Laideira and Viegas, 2016) (refer to paragraphs X and X).
- *Biomarkers of effect* - “a measurable biochemical, physiological, behavioural or other alteration within an organism that, depending upon the magnitude,

can be recognised as associated with an established or possible health impairment or disease” (IPCS, 1993^[BG1]), for example measures of chromosome damage, related to carcinogenicity (refer to paragraphs X and X).

- *Biomarkers of susceptibility* - A biomarker of susceptibility may be defined as an indicator of an inherent or acquired ability of an organism to respond to the challenge of exposure to a chemical (Manno) (refer to paragraphs XX to XX).

3. The over-arching summary Guidance Statement ([G01](#)) provides the Committee’s views on the general principles relating to carcinogenic hazard and risk assessment and a background to the individual components of the risk assessment process and how these are integrated. This statement aims to provide detail of how biomarkers are utilised within the individual components of the risk assessment process.

4. The Committee recommends a four-stage approach to the risk assessment of chemical carcinogenicity which is based on the widely adopted paradigm proposed by the National Academy of Sciences (US National Academy of Sciences, 1983). This is summarised as follows:

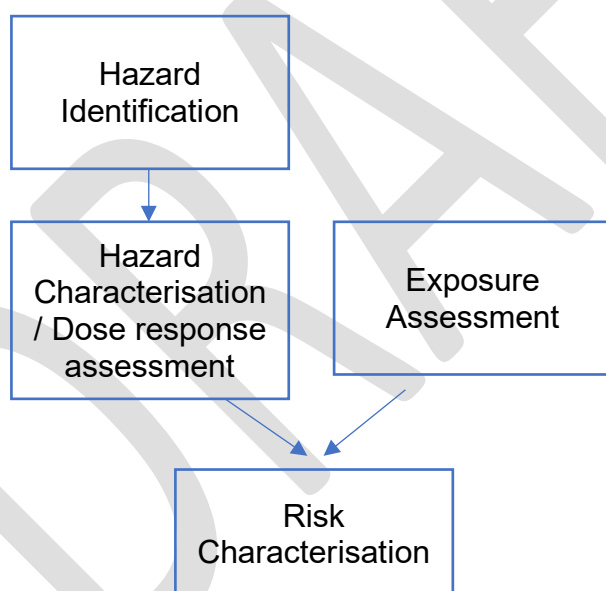


Figure 1: Four stage evaluation strategy for the risk assessment process of carcinogenic hazard

5. In recent discussions, COC has expressed the aspiration to move away from traditional risk assessment approaches for potential carcinogens, to a more “holistic approach” encompassing consideration of the effects of chemicals on all stages of cancer development (COC doc to ref when available).

6. Within exposure assessment, biomarkers can be used (usually) to establish recent exposures to, and uptake of, actual or putative carcinogens in human populations or experimental animals. Within hazard assessment, biomarkers may be

used to quantitatively associate a dose or exposure with either a precursor carcinogenic effect or the probability of a disease outcome. In this context, biomarkers can provide specific evidence that a chemical has the potential to cause a carcinogenic effect and may also provide information on mode of action. Therefore, biomarkers provide a range of possible measurements from systemic exposure through to resulting causal events in the process of carcinogenesis.

7. When utilising biomarker data, it is important to consider that there is usually a long latency period between first exposure to the carcinogen and the clinical onset of cancer. Currently, established biomarkers of exposure often represent recent exposure but some which show organ or tissue retention can be used to assess long-term exposure.

8. Biomarkers are powerful tools for investigating the mode of carcinogenic action (MOA) and can be incorporated into animal studies for this purpose. Indeed, biomarkers, where a clear rationale for the alteration of the level of biomarker with the underlying latent variable, can be useful to discern mechanisms of action. Conversely, knowledge of MOA may help in the development of better biomarkers for use in human exposure scenarios. It should be noted however, that any biomarker used in animal studies must have relevance to humans and not represent an animal (species)-specific response (see for example, Bartsch et al., 2018).

Biomarker characterisation and validation

9. The Committee has a role in evaluating the entire spectrum of biomarkers including the development, validation and practicality of new techniques and the applicability and interpretation of well-established methods.

10. Biomarkers must be appropriately characterised and validated before conclusions are drawn from their use. There are a number of criteria that should be considered when selecting and validating suitable biomarkers for use in human biomonitoring studies (Vorkamp et al., 2021). For example, the general criteria used for the evaluation of the most suitable exposure biomarkers (EB) and matrix (M) for the current European initiative, HBM4EU for carcinogens and non-carcinogens, include:

- Specificity – concentration of the exposure biomarker in the selected matrix should exclusively reflect exogenous exposure and is a consequence of environmental and/or occupational exposure. It is noted that for chemicals with very limited data, exclusivity may not be achievable, however concentrations should be a correct indication of exposure.
- Biological sensitivity - the measured concentration of the EB/M correlates strongly with the substance intake dose. Variations of EB/M concentration reflect the variation of exposure to the substance of interest. For chemicals with very limited data, the measured concentration of the EB/M is an acceptable indication of the substance intake dose.

- Half-life - the EB/M should preferably have a half-life sufficiently long to avoid an excessive intra-individual variability in EB/M concentration measurement.
- Stability after sample collection - cryo-preservability of EB/M is sufficient to guarantee a high stability during storage in the biobank or analyse the sample as soon as possible.
- Matrix availability and sample collection - the sample collection of the relevant matrix is not considered too invasive. Easy collection and transportation of the required amount of sample with a validated sampling protocol is beneficial. It is advantageous if it is possible to determine more than one EB in the same matrix where it is relatively easy to obtain a sufficient sample volume for a required number of samples.
- Measurement validity - the EB/M concentration in the sample is not likely to be altered by contamination with a ubiquitous parent substance from the environment preceding and during the analysis. Variations in matrix composition can be easily corrected for (e.g. creatinine in urine, lipids in serum). Sample contamination by a ubiquitous parent substance might occur, but the level of contamination is low compared to expected levels and special precautions can be applied to minimize the amount of contamination.

11. Biomarkers used in animal studies must also be suitably characterised and validated and this should be based on the principles detailed for human biomarkers. In relation to biomarkers, the STrengthening the Reporting of OBservational studies in Epidemiology – Molecular Epidemiology (STROBE-ME) initiative (Gallo et al., 2011) provides). standardised guidelines and a ‘checklist’ for the reporting of biomarker and molecular epidemiology studies (see <http://www.equator-network.org/reporting-guidelines/strobe-me/>, accessed June 2021).

Strategic uses of HBM

12. Probably the most well-developed use of HBM has been in occupational settings where exposures to a chemical of particular concern might be relatively high. Here, routine HBM might be more informative about risk than air monitoring and, various types of reference values used for risk management might exist for the chemical of concern. In the general population, HBM is often used to inform on exposure to chemical of particular concern and also, for changes over time (increase or decrease) for substances of interest related to industrial or consumer usage to existing or newly-introduced substances (Bevan et al., 2017).

13. Ongoing national and international surveillance programmes such as the US National Health and Nutrition Examination Survey (NHANES), the Canada Health Measures Survey (CHMS) and the German Environmental Survey (GerES) typically use well-established biomonitoring techniques (e.g. biomarkers which are known to reflect exposure to the chemical of interest, standardized sampling methods and verified analytical techniques) to collect information on population exposures to environmental hazards that are known to be significant to public health. As

biomonitoring does not generally determine exposure sources and routes of exposure, environmental monitoring remains crucial (Ladeira and Viegas, 2016).

14. A Biomonitoring Equivalent (BE) is an estimated concentration or range of concentrations of an environmental chemical in humans that is consistent with existing health-based guidance values such as the Tolerable Daily Intake (TDI) or reference dose or concentration (RfD, RfC). It provides a way of interpreting biomonitoring data in the context of these values (Hays et al., 2008; LaKind et al., 2008). It is envisaged that they will be useful for understanding and prioritising risk management practices and will enable the available biomonitoring data to be utilised more fully. However, to date, there is limited information on the use of BEs for estimating chemical exposure in the context of carcinogenesis (Faure et al., 2020).

15. Human biomonitoring guidance values (HBM-GVs) are being derived by the European Human Biomonitoring Initiative referred to as HBM4EU. There is currently a diversity in the derivation of health-based guidance values for both the general population and for occupational exposure. The HBM4EU initiative aims to increase confidence in HBM-GVs derived using a harmonised, systematic and generally accepted strategy for the derivation of HBM-GVs at the European level (Vorkamp et al., 2021).

Biomarkers of exposure

16. The objective of human exposure assessment is to estimate probable exposure by determining exposure routes, source, magnitude and duration of contact with the chemical of concern. However, epidemiological studies can have limitations related to measurement of exposure to carcinogens over long periods, and exposure assessment is frequently identified as the main area of uncertainty in the overall risk assessment process. Although the alternative approach of personal monitoring (e.g. dermal patch studies) provides a way to measure exposure directly, assumptions need to be made about the relationship between results from short-term sampling and predicted long-term exposure. Approaches used in exposure assessment and the characterisation of uncertainties and variability in the resulting estimates have been extensively reviewed elsewhere (Ladeira and Viegas, 2016).

17. Biomarkers of exposure can indicate the presence of a carcinogenic compound or its biological interactions. This is achieved by assaying levels of the chemical, a metabolite or a reaction product in blood, urine, saliva, cerebrospinal fluid, or other biological samples. In this way, exposure biomarkers can provide direct evidence of human exposure to a carcinogen through internal dose. In addition, any factors that may impact on target organ concentrations, such as individual phenotype should be taken into consideration.

18. Where a relationship can be established between biomarker levels and external and internal dose, data from exposure biomarkers can be used to calculate the initial external exposure. It is important that a relationship can be established to counter the potential impact of interfering factors. Physiologically Based Pharmacokinetic (PBPK) models are valuable tools to help define safe external levels of chemicals based on internal doses at target organs in experimental

animals, humans and organisms used in environmental risk assessment (Paini et al., 2021).

19. Biomarkers such as adducts (DNA or protein) are important in understanding the kinetics and potential biological interactions of a chemical, for example if it is capable of interacting with DNA. Many biomarkers of exposure are short-lived and provide short- to medium-term indications of internal exposure. Adipose tissue (AT) is also recognised as a significant site of lipophilic toxicant bioaccumulation, thereby reflecting cumulative exposure. It has been suggested that AT plays a major role in the storage and overall toxicokinetics of hydrophobic xenobiotic persistent organic pollutants (POPs), which has both positive and negative consequences. The sequestration of POPs can be both protective, i.e. by removing them from the blood stream but are potentially harmful if released from AT, for example during rapid periods of weight loss (Jackson et al., 2017).

20. Biomonitoring, the direct measurement of a chemical or its metabolites in biological samples, has been widely used within the risk assessment process, for both carcinogens and non-carcinogens. Validated biomarkers of internal exposure have been identified for a wide range of environmental chemicals and metabolites, including: metals; polyaromatic hydrocarbons (PAHs); polychlorinated dibenzodioxins (PCDD), polychlorinated biphenyls (PCB); phthalates; pesticides; aromatic amines; perfluorinated substances; tobacco smoke components; and volatile organic compounds (VOCs) (Jeddi et al., 2021). Biomarkers of exposure can be used in animal studies to provide important information which can contribute to carcinogenic MOA investigations. For example, investigations of the carcinogenic potential of acrylamide utilising DNA and haemoglobin (Hb) adduct data (Virk-Baker et al., 2014; Xu et al., 2014).

DNA adducts_[BG2]

21. DNA adducts, where DNA is covalently bound to a chemical moiety, characterise the first stage of DNA damage and provide a marker of exposure to reactive chemicals or their metabolites. The presence of DNA adducts may demonstrate systemic exposure to specific target tissues. Their measurement can be used in human biomonitoring studies investigating environmental exposures to chemicals. DNA adducts can be measured in peripheral blood lymphocytes (PBLs), exfoliated cells, such as from the urothelium or buccal mucosa, and in tissue biopsy samples.

22. DNA adducts are commonly used as biomarkers of exposure when investigating exposure to PAHs from sources such as tobacco smoking (Phillips, 2005; Veglia et al., 2003_[RB3]; Ewa and Danuta, 2017; Ma et al., 2019), environmental pollution (Farmer et al., 2003; Singh et al., 2007; Ewa and Danuta, 2017; Totsuka et al., 2021) or occupational exposure (Lee et al., 2003; Taioli et al., 2007; Ewa and Danuta, 2017). The epidemiological link between aflatoxin B1 exposure and hepatocellular carcinoma development is strongly supported by investigations using DNA adducts as biomarkers of exposure (Rundle, 2006; Wogan et al., 2011). In addition, aflatoxin biomarkers have also been used as predictive values for cancer outcome to be used as short-term indicators for intervention trials (Kensler et al.,

2003). [RB4] Exposure to acrylamide is strongly associated with the production of DNA adducts *in vitro* and in experimental animals but the correlation is less clear in humans (Xu et al., 2014, Li et al., 2016). The mode of action of aristolochic acid, a naturally occurring component of plants of the *Aristolochia* family associated with nephropathy and urothelial cancer, has been investigated using DNA adducts and specific DNA adducts have been identified as a biomarker in an exposed population (Jadot et al., 2017).

23. The biological significance of DNA adducts has been considered by ECETOC and ILSI/HESI workshops (Pottenger et al., 2009; Sander et al., 2005). Both reached the general consensus that DNA adducts had an important role in the risk assessment process and in establishing mode of carcinogenic action. However, adducts vary greatly in their mutagenic potency and it is not possible to establish a generic level below which there is no adverse biological response. Understanding the role of processes such as DNA repair, cell turnover and death is critical to establishing the significance of adducts in the generation of mutagenic lesions and the subsequent development of a tumour. Accordingly, association of an adduct with a disease does not automatically indicate causality, although there is considerable evidence indicating that they can inform epidemiological investigations with regard to causation. It has also been proposed that DNA adducts can be useful biomarkers of cumulative exposure, representing cumulative unrepaired DNA damage (Kwack et al., 2014).

24. Frameworks and guidance have been developed by ILSI-HESI workgroups with a view to standardising methodological approaches and for data presentation and interpretation. An organisational approach for the assessment of DNA adduct data outlines how information which defines and characterises the DNA adduct (e.g. type of adduct, frequency, persistence, type of repair process) should be integrated with other relevant data, such as dosimetry, toxicity, toxicokinetics, genotoxicity, and tumour incidence, to inform on the chemical MOA. DNA adducts are considered biomarkers of exposure, whereas gene mutations and chromosomal alterations represent biomarkers of early biological effects but are also potential bio-indicators of the carcinogenic process (COM. 2021). DNA adduct data are most effectively utilised when viewed in the context of other information within the risk assessment framework.

25. The presence of endogenous DNA adducts at very low levels ($< 1 \times 10^6$ nucleosides) necessitates highly specific and sensitive methods of identification and quantitation. Early studies utilised ^{32}P -postlabelling, mass spectrometry (MS), immunoassay and fluorescence detection (Himmelstein et al., 2009). More recently, advances in mass spectrometry have also led to the development of DNA adductome analysis, an emerging method that simultaneously screens for multiple DNA adducts and provides relevant structural information (Totsuka et al., 2021). MS is made more sensitive through pre-separation or enrichment of samples and by combining with other techniques such as liquid-chromatography (LC-MS) and gas chromatography (GC-MS) (Liu and Wang, 2015). The most sensitive marker of DNA damage is considered to be the phosphorylation of histone H2A variant H2AX at Ser 139 (γ -H2AX). This correlates well with each double-strand break (DSB) and can be used to

examine both DNA damage and the subsequent repair of the DNA lesion. γ -H2AX can be detected by immunoblotting and immunostaining using microscopic or flow cytometric detection (Ji et al., 2017).

Protein (Haemoglobin or Albumin) adducts

26. Adducts of chemicals with proteins such as Hb and albumin can also be used as biomarkers of exposure to carcinogens (Hartwig et al., 2020). Occupational exposure to 1,3-butadiene and styrene have been effectively investigated using Hb-adduct methodology (Vacek et al., 2010; Boysen et al., 2012; Ogawa et al., 2006). Acrylamide exposure in humans has been successfully monitored by measuring Hb adducts of acrylamide itself or its metabolite glycidamide (Vikstrom et al., 2012). Similarly, albumin adducts of aflatoxin have been detected in exposed populations (McCoy et al., 2008) and biomonitoring of arylamines and nitroarenes utilises albumin adducts (Sabbionu and Turesky, 2017).

Biomarkers of Effect

27. Biomarkers of a key event implicated in a carcinogenic mode of action may be used to characterise the hazard. With regard to carcinogenicity, the most commonly studied biomarkers of effect measure genotoxicity endpoints such as chromosomal changes (Albertini et al., 2000; Bonassi et al., 2005 and 2011). It is important to recognise that, in some instances, these biomarkers of effect may only be indicative of immediate alterations and may not represent injury resulting in actual impairment of health or disease. Biomarkers of effect are frequently not specific to a given exposure or a specific agent and are influenced by sources of inter- and intra-individual variability, including species, sex and age. The relationship between exposure (acute, subacute, or chronic), the biomarker of effect, and carcinogenic event must be established in order to determine validity (Jeddi et al., 2021).

Genotoxicity Biomarkers

28. The Committee on Mutagenicity of Chemicals in Food, Consumer Products and the Environment (COM) Guidance on a Strategy for Genotoxicity Testing of Chemicals (COM, 2021) defines three possible types of genetic damage following exposure to chemicals, namely gene mutation, changes to chromosome structure (i.e. clastogenicity) and number (i.e. aneuploidy). As there is currently no single validated assay that can provide comprehensive information on all three types of genetic damage, it is important that testing is carried out with several different assays to provide comprehensive coverage of the mutagenic potential of a chemical. Assays may be classified on the basis of genetic endpoints (e.g. gene mutation, clastogenicity, aneugenicity and tests for DNA damage) or by consideration of the different phylogenetic levels (e.g. bacteria, and mammalian cells) represented and also in mammals by the tissues or target organs studied.

29. Cytogenetic endpoints such as micronuclei (MN) and chromosome aberrations (CA) are considered to be biomarkers of early carcinogenic effect and are thought to be predictive for cancer risk in humans (Bonassi et al., 2011; Wang et al., 2012; Bonassi et al., 2016; Barth et al., 2017; Vodenkova et al., 2015; Hopf

et al., 2019). Sampling of blood and the preparation and analysis of PBLs /buccal mucosa epithelial cells for MN or PBLs for CA are techniques often used in occupational and environmental biomonitoring studies (Mahmoodi et al., 2017; Brucker et al., 2019).

30. The comet assay detects single strand breaks and alkali-labile lesions in DNA using PBLs and is well established as a biomonitoring tool for occupational and environmental exposures (Azqueta et al., 2020). However, its value for predicting cancer is not yet known because it has not been investigated in prospective cohort studies. An understanding of the covariates influencing inter-individual background levels is also critical in the design of such studies (Azqueta et al., 2020) and a role of genotype is also implicated in this variability (Koppen et al., 2017).

31. 8-Hydroxy-2'-deoxyguanosine (8-OHdG) is a non-specific marker of oxidative damage to DNA developed as a biomarker of biochemical effect. 8-OHdG levels can be assessed using PBLs and, as oxidised DNA repair products are excreted, they can also be assayed in biofluids such as urine (Loft et al., 2012a). 8-OHdG levels have been widely used in studies ranging from those examining workers occupationally exposed to PAHs to those occupationally exposed to air pollution (Brucker et al., 2019). There is good evidence that increases in this biomarker correlate with exposure to potential mutagens and these increases are broadly in accordance with comet results (Brucker et al., 2019). Whilst there is good experimental evidence that 8-OH-dG has potential as a biomarker of effect, its reliability is still being evaluated and is the subject of extensive research.[RB6][BG7][RB8]

32. Epigenetic features are increasingly considered sensitive to environmental exposures, and therefore could serve as an important biomarker of effect (Sanchez-Guerra et al., 2015; Shukla et al., 2019). Epigenetic modifications are flexible genomic parameters that can change genome function under exogenous influences. To date, epigenetic changes include DNA methylation, histone modifications, and miRNAs that affect post-transcriptional regulation, without changes to nucleotide sequences. It has been proposed that environmental factors could influence epigenetic processes, leading to epigenetic reprogramming and contributing to the development of several diseases (Ji et al., 2016).

33. Despite progress in the identification of biomarkers, gene mutation-based approaches still face considerable challenges as cancer evolves from a complex interplay between environment and host. Therefore, identification of an epigenomic signature might be useful for early diagnosis with the potential to reduce the environmental-associated disease burden (Ceccaroli et al., 2015).

34. Evidence that genotoxicity biomarkers are indicative of cancer risk in humans is not extensive. Furthermore, the presence of genotoxicity biomarkers does not inform on the precise nature of the chemical exposure which has occurred to give rise to the measured endpoint.

Non – genotoxicity biomarkers

35. Non-genotoxic carcinogens (NGCs) demonstrate a broader mechanistic variety than genotoxic carcinogens, altering epigenetics, the endocrine system, apoptotic signalling, cell proliferation, and/or gap-junctional intercellular communication. In addition, simultaneous alteration of multiple pathways is often required to prompt non-genotoxic oncogenesis (Wilde et al., 2018). Examples of NGCs include 1,4-dichlorobenzene that acts as a tumour promoter, 17 β -oestradiol that is an endocrine-modifier, 2,3,7,8-tetrachlorodibenzo-p-dioxin that is a receptor-mediator, cyclosporine which is an immunosuppressant or metals such as arsenic and beryllium that induce tissue-specific toxicity and inflammatory responses (Hernández, et al., 2009). [RB9]

Molecular [BG10] Epidemiology [BG11] in Cancer Risk Assessment

36. Epidemiologists are currently challenged with measuring the biological effect of an exposure occurring at low doses, whilst recognising the effect of a single component in a mixture and highlighting the link between genetic and environment factors in the etiology of cancer. Population-based studies involving environmental and occupational exposure, infectious agents, personal susceptibility factors, and acquired genetic factors may identify high-risk populations that are likely to develop cancer; additionally, such studies are very informative and significant in designing future community-based health initiatives.

37. Molecular epidemiology is simply defined as the application of the techniques of molecular biology to the study of populations, with a particular focus on the investigation of disease prevalence (Vineis et al., 2005). The term encompasses the use of biomarkers to investigate the events and potential mechanisms which occur during the process of carcinogenicity, from initial exposure to disease (also called the 'meet in the middle approach'; Vineis and Perera, 2007; Vineis and Chadeau-Hyam, 2011;).

38. Developments in this field have been underpinned by the improvement of genetic and molecular techniques identifying environmental and genetic risk factors in the aetiology of cancer. There is a large body of literature which describes the development of potential new biomarkers of exposure and effect and discusses the usefulness and limitations of biomarker measurement (e.g. Ceccaroli et al, 2015).

Biomarkers of Susceptibility

39. Evidence suggests that genetic susceptibility may play a role in an individual's response to exogenous and environmental exposures. Consequently, a number of studies have explored the interactions between genetics and exposures in the aetiology of disease (for example, Kelly and Vineis, 2014).

40. The use of detailed PBPK models for interpreting biomonitoring data also allows for the modelling of different sources of interindividual variability of the absorption, distribution, metabolism and excretion processes, such as body weight, age, genetic polymorphisms in xenobiotic metabolic pathways, excretion and

elimination rates and others. The previously so-called confounders or uncertainty factors can be treated as analysable variables which reflect variations in the susceptibility within a population that is exposed to environmental pollutants (Ladiera and Viegas, 2016).

41. Several large genome-wide association studies (GWAS) have assessed genetic variation in the aetiology of different cancers, e.g. under the International Agency for Research on Cancer (for a list of publications from the study see <http://epic.iarc.fr/research/activitiesbyresearchfields/geneticepidemiology.php>, accessed 14/06/21) and the US National Cancer Institute (for a list of publications from the study see <https://epi.grants.cancer.gov/gameon/>, accessed 14/06/21). The Committee has considered GWAS previously and interactions between genotype and chemicals in the environment. It was concluded that, whilst such data are useful, it would be difficult to use the derived information for the risk assessment of specific chemical carcinogens at the current stage of technique development without a clearer understanding of the functional links and biological relevance of each genotype (COC, 2011).

Omics technologies

42. The development of omics technologies (genomics, proteomics, metabolomics) to investigate gene and protein changes following chemical exposure, and its use in toxicological risk assessment has previously been reviewed in detail by the COT, COC and COM (COT, COC and COM, 2004; COT, 2012). In addition, COM is currently preparing an updated evaluation of the application of transcriptomics and next generation sequencing to genotoxicity and carcinogenicity assessment.

42. The conceptual term ‘exposome’ has been developed as a means to consider an individual’s exposures over the entire life course from conception until death (Wilde et al., 2005; Vineis et al., 2020; Huhn et al., 2021). It has been described as “the totality of internal human exposure with regards to exogenous chemicals, their biotransformation products, and endogenous chemicals sensitive to various environmental exposures and potentially involved in signaling pathways” (Escher et al., 2017).

43. Understanding the relation of external to internal exposures is a central aspect of an exposome assessment, however at present there is no consensus on how to assess the exposome. A number of major ongoing projects investigating approaches to implement the exposome concept with a focus on environmental chemicals have been developed, including: Human Early Life Exposure (HELIX); Health and Environment-wide Associations based on Large Population Surveys (HEALS); EXPOsOMICS; and HBM4EU. All of these projects related their research to existing infrastructures and data available in different European cohorts with the aim of comparing health outcomes and exposure information and to generate cohort-related biosamples and exposome data (Huhn et al., 2021).

44. Tools such as Exposome-Explorer (<http://exposome-explorer.iarc.fr>; accessed 14/06/2021) are also available. This is the first database dedicated to the collation of biomarkers of exposure to environmental risk factors for diseases and aims to provide comprehensive data on all known biomarkers of exposure to dietary factors, pollutants, and contaminants measured in population studies (Neveu et al., 2017).

45. The application of omics technologies to carcinogenicity evaluation was previously considered by the COC as part of its discussions on alternatives to the use of the 2-year rodent bioassay for carcinogenicity risk assessment (COC Guidance Statement [G07](#) Part C). However, as a result of discussions to move away from traditional risk assessment approaches for potential carcinogens, G07 is undergoing substantial revision which will include an updated evaluation of the use of omics technologies.

46. Epigenetics, heritable changes in gene expression which are independent of changes in DNA sequence are being increasingly recognised as part of the process of carcinogenesis (Barrow and Michels, 2014). Epigenetic mechanisms include changes in DNA methylation. There is evidence that some chemical exposures result in epigenetic modifications which could impact on the induction of cancer and may act as historical biomarkers of exposure (Verma, 2015). The possibility of use of epigenetic change as a biomarker of exposure has been discussed at the joint COC, COM and COT meeting in October 2017 where use of epigenetics in chemical risk assessment was discussed (COC, COM, COT, 2019).

47. Non-coding RNAs (ncRNAs) constitute the majority of the human transcribed genome and have been implicated in many pathological conditions, especially cancer (Chan and Tay, 2018). One subclass of ncRNAs, miRNAs, which have a role in the regulation of translation of protein from mRNA, have been explored for development as a biomarker of effect. These miRNAs are differentially expressed in many cancer types and found in the circulation (Brase et al., 2010; Calin and Croce, 2006; Mitchell et al., 2008; Mo et al., 2012). miRNA species are coded from regions of the genome that can be under epigenetic control and can be differentially methylated in cancer (Chuang and Jones, 2007; Li et al., 2012; Lujambio et al., 2008). This raises the possibility that epigenetic change resulting from carcinogen exposure may lead to altered miRNA expression via differential methylation and that this could be a biomarker of historical carcinogen exposure and arbiter of potential future effect (Vrijens et al., 2015; Krewski et al., 2020).

48. The use of ncRNAs as potential biomarkers in regulatory toxicology was discussed at an ECETOC workshop, the summary of which noted “To make available ncRNAs as biomarkers for regulatory toxicology and RA, normal and adverse ncRNA profiles and dose-response relationships of effects should be determined, and ncRNA expression profiles should be linked to phenotypic alterations. Further, it should be determined whether ncRNA levels in specific body fluids reflect levels in specific target tissues. Even though a number of research projects demonstrated a lack of toxicologically relevant uptake and activity of ingested RNAs, bioavailability of ingested ncRNAs and potential impacts to the consuming organism may merit further investigation” (ECETOC, 2016).

Summary

49. A biomarker, in the context of chemicals and carcinogenicity, is defined as an observable change related to the carcinogenic process following a specific exposure or effect.
50. In cancer risk assessment, biomarkers can be utilised for hazard identification and characterisation, for exposure assessment and to inform the MOA.
51. The relationship between the biomarker and the carcinogenic response should be established.
52. Biomonitoring studies should fulfil pre-defined criteria and human-specific biomarkers should be appropriately characterised and validated. Particular attention should be given to ascertaining the stability and half-life of the biomarker and how these impact on the interpretation of epidemiological data.
53. Biomarkers of exposure include DNA and protein adducts, MN and CA. Biomarkers of effect include genotoxicity biomarkers such as MN and CA, and the indicator of oxidative damage, 8-OHdG.
54. The Committee maintains an on-going awareness of the development of newer techniques including molecular epidemiology methods, omics technologies and the emergence of the exposome. However, many of the techniques are still experimental and are useful only for deriving qualitative measurements or information contributing to MOA investigations. It is not currently possible to provide specific guidance on their use in a quantitative capacity.
55. The Committee continues to evaluate the usefulness of the entire spectrum of biomarker techniques including the applicability and interpretation of well-established methods.

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Date TBC

References[RB12]

- Agundez, J.A. (2008). Polymorphisms of human N-acetyltransferases and cancer risk. *Current drug metabolism* 9, 520-531.
- Albertini, R.J., Anderson, D., Douglas, G.R., Hagmar, L., Hemminki, K., Merlo, F., Natarajan, A.T., Norppa, H., Shuker, D.E., Tice, R., *et al.* (2000). IPCS guidelines for the monitoring of genotoxic effects of carcinogens in humans. International Programme on Chemical Safety. *Mutat Res* 463, 111-172.
- Angerer, J., Ewers, U., and Wilhelm, M. (2007). Human biomonitoring: state of the art. *Int J Hyg Environ Health* 210, 201-228.
- Athersuch, T. (2016). Metabolome analyses in exposome studies: Profiling methods for a vast chemical space. *Arch Biochem Biophys* 589, 177-86.
- Azqueta, A., Ladeira, C., Giovannelli, L. *et al.* (2020) Application of the comet assay in human biomonitoring: An hCOMET perspective. *Mutation Research/Reviews in Mutation Research* 783, 108288.
- Barrow, T.M., Michels, K.B. (2014) Epigenetic epidemiology of cancer. *Biochem Biophys Res Commun* 455, 70-83.
- Barth, A., Brucker, N., Moro, A. M., Nascimento, S., Goethel, G., Souto, C., Garcia, S. C. (2017). Association between inflammation processes, DNA damage, and exposure to environmental pollutants. *Environmental Science and Pollution Research*, 24(1), 353–362.
- Bartsch, R., Brinkmann, B., Jahnke, G., Laube, B., Lohmann, R., Michaelsen, S., Neumann, I., Greim, H. (2018) Human relevance of follicular thyroid tumors in rodents caused by non-genotoxic substances. *Regulatory Toxicology and Pharmacology*, 98, 199-208,
- Battershill, J.M., Burnett, K., and Bull, S. (2008). Factors affecting the incidence of genotoxicity biomarkers in peripheral blood lymphocytes: impact on design of biomonitoring studies. *Mutagenesis* 23, 423-437.
- Boffetta, P., Winn, D.M., Ioannidis, J.P., Thomas, D.C., Little, J., Smith, G.D., Coglian, V.J., Hecht, S.S., Seminara, D., Vineis, P., *et al.* (2012). Recommendations and proposed guidelines for assessing the cumulative evidence on joint effects of genes and environments on cancer occurrence in humans. *International journal of epidemiology* 41, 686-704.
- Bonassi, S., and Au, W.W. (2002). Biomarkers in molecular epidemiology studies for health risk prediction. *Mutat Res* 511, 73-86.

Bonassi, S., El-Zein, R., Bolognesi, C., and Fenech, M. (2011). Micronuclei frequency in peripheral blood lymphocytes and cancer risk: evidence from human studies. *Mutagenesis* 26, 93-100.

Bonassi, S., Milic, M., & Neri, M. (2016). Frequency of micronuclei and other biomarkers of DNA damage in populations exposed to dusts, asbestos and other fibers. A systematic review. *Mutation Research*, 770, 106–118.

Bonassi, S., Ugolini, D., Kirsch-Volders, M., Stromberg, U., Vermeulen, R., and Tucker, J.D. (2005). Human population studies with cytogenetic biomarkers: review of the literature and future perspectives. *Environ Mol Mutagen* 45, 258-270.

Bonassi, D., El-zein, R., Bolognesi, C., Fenech, M. (2011) Micronuclei frequency in peripheral blood lymphocytes and cancer risk: evidence from human studies. *Mutagenesis*, 26, 93–100

Boysen, G., Georgieva, N.I., Bordeerat, N.K., Sram, R.J., Vacek, P., Albertini, R.J., and Swenberg, J.A. (2012). Formation of 1,2:3,4-diepoxybutane-specific hemoglobin adducts in 1,3-butadiene exposed workers. *Toxicol Sci* 125, 30-40.

Brase, J.C., Wuttig, D., Kuner, R., and Sultmann, H. (2010). Serum microRNAs as non-invasive biomarkers for cancer. *Molecular cancer* 9, 306.

Brucker, N., do Nascimento, S.N., Bernardini, L., Charao, M.F., Garcia, S.C. (2020) Biomarkers of exposure, effect, and susceptibility in occupational exposure to traffic-related air pollution: A review. *Journal of Applied Toxicology*, 40, 722-736.

Bull, S., Fletcher, K., Boobis, A.R., and Battershill, J.M. (2006). Evidence for genotoxicity of pesticides in pesticide applicators: a review. *Mutagenesis* 21, 93-103.

Calin, G.A., and Croce, C.M. (2006). MicroRNA signatures in human cancers. *Nat Rev Cancer* 6, 857-866.

Ceccaroli, C., Pulliero, A., Geretto, M. and Izzotti, A. (2015) Molecular fingerprints of environmental carcinogens in human cancer. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 33, 188-228.

Chadeau-Hyam, M., Athersuch, T.J., Keun, H.C., De Iorio, M., Ebbels, T.M., Jenab, M., Sacerdote, C., Bruce, S.J., Holmes, E., and Vineis, P. (2011). Meeting-in-the-middle using metabolic profiling - a strategy for the identification of intermediate biomarkers in cohort studies. *Biomarkers* 16, 83-88.

Chan JJ, Tay Y. (2018) Noncoding RNA:RNA Regulatory Networks in Cancer. *International Journal of Molecular Sciences*, 19, 1310.

Choi 2015

Chuang, J.C., and Jones, P.A. (2007). Epigenetics and microRNAs. *Pediatric research* 61, 24R-29R.

COC (2011) Interaction between genotype and chemicals in the environment on the induction of cancer in risk assessment. In COT, COM and COC Joint Annual Report 2011, p53-54.

COC, COM, COT (2019) Statement from a joint Committee workshop on the use of epigenetics in chemical risk assessment. Available from:

<https://www.gov.uk/government/publications/epigenetics-in-chemical-risk-assessment-joint-committee-statement>

Collins, A., Koppen, G., Valdiglesias, V., Dusinska, M., Kruszewski, M., Møller, P., Rojas, E., Dhawan, A., Benzie, I., Coskun, E., Moretti, M., Speit, G., Bonassi, S., ComNet project (2014). The comet assay as a tool for human biomonitoring studies: the ComNet project. *Mutat Res Rev Mutat Res* 759, 27-39.

COM (2006). Statement on risk factors affecting the formation of chromosomal aberrations and micronuclei in peripheral blood lymphocytes. COM/06/S3-December 2006. Link:

<http://webarchive.nationalarchives.gov.uk/20140506144244/http://www.iacom.org.uk/statements/documents/pbl0603.pdf> (accessed 27/04/17)

COT, COC and COM (2004) Joint statement on the use of toxicogenomics in toxicology. Available from:

<https://cot.food.gov.uk/cotstatements/cotstatementsyrs/cotstatements2004/toxicogenomics> (accessed 13/07/2017)

COT (2012) COT statement on the use of toxicogenomics data in risk assessment. COT/2012/02. Available from:

<https://cot.food.gov.uk/cotstatements/cotstatementsyrs/cotstatements2012/646694> (accessed 13/07/2017)

Dellarco, V.L., McGregor, D., Berry, S.C., Cohen, S.M., and Boobis, A.R. (2006). Thiazopyr and thyroid disruption: case study within the context of the 2006 IPCS Human Relevance Framework for analysis of a cancer mode of action. *Crit Rev Toxicol* 36, 793-801.

Dong, L.M., Potter, J.D., White, E., Ulrich, C.M., Cardon, L.R., and Peters, U. (2008). Genetic susceptibility to cancer: the role of polymorphisms in candidate genes. *JAMA : the journal of the American Medical Association* 299, 2423-2436.

ECETOC (2009). Use of Markers for Improved Retrospective Exposure Assessment in Epidemiology Studies. Workshop Report No. 14. Available from:

<http://www.ecetoc.org/publication/workshop-report-14-use-of-markers-for-improved-retrospective-exposure-assessment-in-epidemiology-studies/> (accessed 13/07/2017)

ECETOC (2016) Noncoding RNAs and Risk Assessment Science. Workshop Report No. 32. Available from: <http://www.ecetoc.org/publication/workshop-report-no-32-noncoding-rnas-risk-assessment-science-3-4-march-2016-malaga/> (accessed 02/05/17)

Escher BI, Hackermuller J, Polte T, Scholz S, Aigner A, Altenburger R, Bohme A, Bopp SK, Brack W, Busch W, Chadeau-Hyam M, Covaci A, Eisentrager A, Galligan JJ, Garcia-Reyero N, Hartung T, Hein M, Herberth G, Jahnke A, Kleinjans J, Kluver N, Krauss M, Lamoree M, Lehmann I, Luckenbach T, Miller GW, Muller A, Phillips DH, Reemtsma T, Rolle-Kampczyk U, Schuurmann G, Schwikowski B, Tan YM, Trump S, Walter-Rohde S, Wambaugh JF (2017) From the exposome to mechanistic understanding of chemical-induced adverse effects. *Environ Int* 99:97–106. <https://doi.org/10.1016/j.envint.2016.11.029>

Ewa B, Danuta MŠ. Polycyclic aromatic hydrocarbons and PAH-related DNA adducts. *J Appl Genet.* 2017 Aug;58(3):321-330. doi: 10.1007/s13353-016-0380-3. Epub 2016 Dec 12. PMID: 27943120; PMCID: PMC5509823.

Farmer, P.B., Singh, R., Kaur, B., Sram, R.J., Binkova, B., Kalina, I., Popov, T.A., Garte, S., Taioli, E., Gabelova, A., *et al.* (2003). Molecular epidemiology studies of carcinogenic environmental pollutants. Effects of polycyclic aromatic hydrocarbons (PAHs) in environmental pollution on exogenous and oxidative DNA damage. *Mutat Res* 544, 397-402.

Faure, S., Noisel, N., Werry, K., Karthikeyan, S., Aylward, L.L., St-Amand, A. (2020) Evaluation of human biomonitoring data in a health risk based context: An updated analysis of population level data from the Canadian Health Measures Survey. *International Journal of Hygiene and Environmental Health*, 223, 267-280

Fenech, M., and Bonassi, S. (2011). The effect of age, gender, diet and lifestyle on DNA damage measured using micronucleus frequency in human peripheral blood lymphocytes. *Mutagenesis* 26, 43-49.

Fenech, M., Holland, N., Chang, W.P., Zeiger, E., and Bonassi, S. (1999). The HUman MicroNucleus Project--An international collaborative study on the use of the micronucleus technique for measuring DNA damage in humans. *Mutat Res* 428, 271-283.

Gallo, V., Egger, M., McCormack, V., Farmer, P.B., Ioannidis, J.P., Kirsch-Volders, M., Matullo, G., Phillips, D.H., Schoket, B., Stromberg, U., *et al.* (2011). STrengthening the Reporting of OBservational studies in Epidemiology - Molecular Epidemiology (STROBE-ME): An extension of the STROBE statement. *Mutagenesis* 27, 17-29.

Gallo, V., Khan, A., Gonzales, C., Phillips, D.H., Schoket, B., Gyorffy, E., Anna, L., Kovacs, K., Moller, P., Loft, S., *et al.* (2008). Validation of biomarkers for the study of environmental carcinogens: a review. *Biomarkers* 13, 505-534.

Garcia-Closas, M., Malats, N., Silverman, D., Dosemeci, M., Kogevinas, M., Hein, D.W., Tardon, A., Serra, C., Carrato, A., Garcia-Closas, R., *et al.* (2005). NAT2 slow acetylation, GSTM1 null genotype, and risk of bladder cancer: results from the Spanish Bladder Cancer Study and meta-analyses. *Lancet* 366, 649-659.

Harris, R.M., Williams, T.D., Waring, R.H. and Hodges, N.J. (2015). Molecular basis of carcinogenicity of tungsten alloy particles. *Toxicol Appl Pharmacol* 283, 223-233.

Hartwig, A., Arand, M., Epe, B., Guth, S., Jahnke, G., Lampen, A., Martus, H.-J., Monien, B., Rietjens, I.M.C.M., Schmitz-Spanke, S., Schrieffer-Schwemmer, G., Steingberg, P., Eisenbrand, G. (2020) Mode of action-based risk assessment of genotoxic carcinogens. *Archives of Toxicology*, 94, 1787–1877

Hays, S.M., Aylward, L.L., LaKind, J.S., Bartels, M.J., Barton, H.A., Boogaard, P.J., Brunk, C., DiZio, S., Dourson, M., Goldstein, D.A., *et al.* (2008). Guidelines for the derivation of Biomonitoring Equivalents: report from the Biomonitoring Equivalents Expert Workshop. *Regul Toxicol Pharmacol* 51, S4-15.

Hernández, L.G., van Steeg, H., Luijten, M., van Benthem, J. (2009) Mechanisms of non-genotoxic carcinogens and importance of a weight of evidence approach. *Mutation Research/Reviews in Mutation Research*, 682, 94-109

Himmelstein, M.W., Boogaard, P.J., Cadet, J., Farmer, P.B., Kim, J.H., Martin, E.A., Persaud, R., and Shuker, D.E. (2009). Creating context for the use of DNA adduct data in cancer risk assessment: II. Overview of methods of identification and quantitation of DNA damage. *Crit Rev Toxicol* 39, 679-694.

Hochstenbach, K., van Leeuwen, D.M., Gottschalk, R.W., Gmuender, H., Stolevik, S.B., Nygaard, U.C., Lovik, M., Granum, B., Namork, E., van Loveren, H., *et al.* (2012). Transcriptomic fingerprints in human peripheral blood mononuclear cells indicative of genotoxic and non-genotoxic carcinogenic exposure. *Mutat Res*.

Hogervorst, J.G., Baars, B.J., Schouten, L.J., Konings, E.J., Goldbohm, R.A., and van den Brandt, P.A. (2010). The carcinogenicity of dietary acrylamide intake: a comparative discussion of epidemiological and experimental animal research. *Crit Rev Toxicol* 40, 485-512.

Holsapple, M.P., Pitot, H.C., Cohen, S.M., Boobis, A.R., Klaunig, J.E., Pastoor, T., Dellarco, V.L., and Dragan, Y.P. (2006). Mode of action in relevance of rodent liver tumors to human cancer risk. *Toxicol Sci* 89, 51-56.

Hopf, N.B., Bolognesi, C., Danuser, B., Wild, P. (2019) Biological monitoring of workers exposed to carcinogens using the buccal micronucleus approach: A systematic review and meta-analysis. *Mutation Research/Reviews in Mutation Research*, 781, 11-29,

Huhn, S., Escher, B.I., Krauss, M., Scholz, S., Hackermüller, J., Altenburger, R. (2021) Unravelling the chemical exposome in cohort studies: routes explored and steps to become comprehensive. *Environmental Sciences Europe*, 33, 17.

IPCS (1993) *Environmental Health Criteria 155: Biomarkers and risk assessment: Concepts and principles*. Geneva, World Health Organization, International Programme on Chemical Safety, 82 pp.

IPCS (1999). *Principles for the assessment of risks to human health from exposure to chemicals*. Geneva, World Health Organization, International Programme on Chemical Safety (*Environmental Health Criteria 210*).

IPCS (2001). *Biomarkers In Risk Assessment: Validity and Validation*. Geneva, World Health Organization, International Programme on Chemical Safety (*Environmental Health Criteria 222*).

Jackson E, Shoemaker R, Larian N, Cassis L. (2018) *Adipose Tissue as a Site of Toxin Accumulation*. *Compr Physiol*. 7, 1085-1135. Erratum in: *Compr Physiol*. 2018, 8, 1251.

Jadot, I., Decleves, A.-E., Nortier, J. and Caron, N. (2017). An integrated view of aristolochic acid nephropathy: update of the literature. *Int J Mol Sci* 18, 297

Jarabek, A.M., Pottenger, L.H., Andrews, L.S., Casciano, D., Embry, M.R., Kim, J.H., Preston, R.J., Reddy, M.V., Schoeny, R., Shuker, D., *et al.* (2009). Creating context for the use of DNA adduct data in cancer risk assessment: I. Data organization. *Crit Rev Toxicol* 39, 659-678.

Zare Jeddi M, Hopf NB, Viegas S, Price AB, Pains A, van Thriel C, Benfenati E, Ndaw S, Bessems J, Behnisch PA, Leng G, Duca RC, Verhagen H, Cubadda F, Brennan L, Ali I, David A, Mustieles V, Fernandez MF, Louro H, Pasanen-Kase R. (2021) Towards a systematic use of effect biomarkers in population and occupational biomonitoring. *Environ Int*. 146:106257.

Ji J, Zhang Y, Redon CE, Reinhold WC, Chen AP, Fogli LK, *et al.* (2017) Phosphorylated fraction of H2AX as a measurement for DNA damage in cancer cells and potential applications of a novel assay. *PLoS ONE* 12(2): e0171582.

Karahalil, B., Bohr, V., and Wilson, D., 3rd (2012). Impact of DNA polymorphisms in key DNA base excision repair proteins on cancer risk. *Human & experimental toxicology* 31, 981-1005.

Kensler, T.W., Qian, G.S., Chen, J.G. and Groopman, J.D. (2003) Translational strategies for cancer prevention in liver. *Nat Rev Cancer* 3, 321-329.

Klaunig, J.E., Babich, M.A., Baetcke, K.P., Cook, J.C., Corton, J.C., David, R.M., DeLuca, J.G., Lai, D.Y., McKee, R.H., Peters, J.M., *et al.* (2003). PPARalpha

agonist-induced rodent tumors: modes of action and human relevance. *Crit Rev Toxicol* 33, 655-780.

Klaunig, J.E., and Kamendulis, L.M. (2005). Mechanisms of acrylamide induced rodent carcinogenesis. *Advances in experimental medicine and biology* 561, 49-62.

Koppen, G., Azqueta, A., Pourrut, B., Brunborg, G., Collins, A.R. and Langie, S.A. (2017). The next three decades of the comet assay: a report of the 11th International Comet Assay Workshop. *Mutagenesis* 32, 397-408.

Kwack SJ, Kim DY, Kim YJ, Roh TH, Choi SM, Lim DS, Shin HS, Kim HS, Lee BM. Potential application of benzo(a)pyrene-associated adducts (globin or lipid) as blood biomarkers for target organ exposure and human risk assessment. *J Toxicol Environ Health A*. 2014;77(22-24):1491-501. doi: 10.1080/15287394.2014.955904. PMID: 25343297.

Krewski, D., Andersen, M.E., Tyshenko, M.G. *et al.* Toxicity testing in the 21st century: progress in the past decade and future perspectives. *Arch Toxicol* **94**, 1–58 (2020). <https://doi.org/10.1007/s00204-019-02613-4>

Ladiera and Viegas 2016

LaKind, J.S., Aylward, L.L., Brunk, C., DiZio, S., Dourson, M., Goldstein, D.A., Kilpatrick, M.E., Krewski, D., Bartels, M.J., Barton, H.A., *et al.* (2008). Guidelines for the communication of Biomonitoring Equivalents: report from the Biomonitoring Equivalents Expert Workshop. *Regul Toxicol Pharmacol* 51, S16-26.

Lee, K.H., Ichiba, M., Zhang, J., Tomokuni, K., Hong, Y.C., Ha, M., Kwon, H.J., Koh, S.B., Choi, H.R., Park, C.G., *et al.* (2003). Multiple biomarkers study in painters in a shipyard in Korea. *Mutat Res* 540, 89-98.

Li, D., Wang, P., Liu, Y., Hu, X., Chen, F. (2016) Metabolism of Acrylamide: Interindividual and Interspecies Differences as Well as the Application as Biomarkers. *Curr Drug Metab* 17, 317-26.

Li, X.Q., Guo, Y.Y., and De, W. (2012). DNA methylation and microRNAs in cancer. *World journal of gastroenterology : WJG* 18, 882-888.

Loft, S., Danielsen, P., Løhr, M., Jantzen, K., Hemmingsen, J.G., Roursgaard, M., Karotki, D.G., Møller, P. (2012a) Urinary excretion of 8-oxo-7,8-dihydroguanine as biomarker of oxidative damage to DNA. *Arch Biochem Biophys* 518, 142-50.

Loft, S., Svoboda, P., Kawai, K., Kasai, H., Sorensen, M., Tjønneland, A., Vogel, U., Møller, P., Overvad, K., and Raaschou-Nielsen, O. (2012b). Association between 8-oxo-7,8-dihydroguanine excretion and risk of lung cancer in a prospective study. *Free Radic Biol Med* 52, 167-172.

Lujambio, A., Calin, G.A., Villanueva, A., Ropero, S., Sanchez-Cespedes, M., Blanco, D., Montuenga, L.M., Rossi, S., Nicoloso, M.S., Faller, W.J., *et al.* (2008). A

microRNA DNA methylation signature for human cancer metastasis. *Proc Natl Acad Sci U S A* **105**, 13556-13561.

Ma B, Stepanov I, Hecht SS. Recent Studies on DNA Adducts Resulting from Human Exposure to Tobacco Smoke. *Toxics*. 2019; 7(1):16.
<https://doi.org/10.3390/toxics7010016>

Mahmoodi, M., Soleyman-Jahi, S., Zendehdel, K., *et al.* (2017). Chromosomal aberrations, sister chromatid exchanges, and micronuclei in lymphocytes of oncology department personnel handling anti-neoplastic drugs. *Drug Chem. Toxicol.* **40**, 235-240.

Manno M., Viau C., in collaboration with, Cocker J., Colosio C., Lowry L., *et al.*, Biomonitoring for occupational health risk assessment (BOHRA), *Toxicol. Lett.*, 2010, 192, 3–16.

Marczynski, B., Rihs, H.P., Rossbach, B., Holzer, J., Angerer, J., Scherenberg, M., Hoffmann, G., Bruning, T., and Wilhelm, M. (2002). Analysis of 8-oxo-7,8-dihydro-2'-deoxyguanosine and DNA strand breaks in white blood cells of occupationally exposed workers: comparison with ambient monitoring, urinary metabolites and enzyme polymorphisms. *Carcinogenesis* **23**, 273-281.

McCoy, L.F., Scholl, P.F., Sutcliffe, A.E., Kieszak, S.M., Powers, C.D., Rogers, H.S., Gong, Y.Y., Groopman, J.D., Wild, C.P., and Schleicher, R.L. (2008). Human aflatoxin albumin adducts quantitatively compared by ELISA, HPLC with fluorescence detection, and HPLC with isotope dilution mass spectrometry. *Cancer Epidemiol Biomarkers Prev* **17**, 1653-1657.

Mitchell, P.S., Parkin, R.K., Kroh, E.M., Fritz, B.R., Wyman, S.K., Pogosova-Agadjanyan, E.L., Peterson, A., Noteboom, J., O'Briant, K.C., Allen, A., *et al.* (2008). Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* **105**, 10513-10518.

Mo, M.H., Chen, L., Fu, Y., Wang, W., and Fu, S.W. (2012). Cell-free Circulating miRNA Biomarkers in Cancer. *Journal of Cancer* **3**, 432-448.

Neveu, V., Moussy, A., Rouaix, H., Wedekind, R., Pon, A., Knox, C., Wishart, D.S., Scalbert, A. (2017) Exposome-Explorer: a manually-curated database on biomarkers of exposure to dietary and environmental factors. *Nucleic Acids Res* **45**, D979–D984.

National Research Council, Biological markers in environmental health research. Committee on Biological Markers of the National Research Council, *Environ. Health Perspect.*, 1987, **74**, 3–9.

OECD (2017). Overview on genetic toxicology TGs. OECD Series on Testing and Assessment 238. Available from: https://www.oecd-ilibrary.org/environment/overview-on-genetic-toxicology-tgs_9789264274761-en

Ogawa, M., Oyama, T., Isse, T., Yamaguchi, T., Murakami, T., Endo, Y., and Kawamoto, T. (2006). Hemoglobin adducts as a marker of exposure to chemical substances, especially PRTR class I designated chemical substances. *J Occup Health* 48, 314-328.

Paini A., Tan Y.-M., Sachana M., Worth A. (2021) Physiologically Based Kinetic (PBK) models are valuable tools to help define safe external levels of chemicals based on internal doses at target organs in experimental animals, humans and organisms used in environmental risk assessment. *Computational Toxicology*, 18, 100163.

Peters, A., Hoek, G., and Katsouyanni, K. (2012). Understanding the link between environmental exposures and health: does the exposome promise too much? *J Epidemiol Community Health* 66, 103-105.

Phillips, D.H. (2005). DNA adducts as markers of exposure and risk. *Mutat Res* 577, 284-292.

Pottenger, L.H., Carmichael, N., Banton, M.I., Boogaard, P.J., Kim, J., Kirkland, D., Phillips, R.D., van Benthem, J., Williams, G.M., and Castrovinci, A. (2009). ECETOC workshop on the biological significance of DNA adducts: summary of follow-up from an expert panel meeting. *Mutat Res* 678, 152-157.

Rappaport, S.M., and Smith, M.T. (2010). Epidemiology. Environment and disease risks. *Science* 330, 460-461.

Rundle, A. (2006). Carcinogen-DNA adducts as a biomarker for cancer risk. *Mutat Res* 600, 23-36.

Sabbioni, G., and Turesky, R.J. (2017). Biomonitoring Human Albumin Adducts: The Past, the Present, and the Future. *Chem Res Toxicol* 30, 332-366.

Sander, M., Cadet, J., Casciano, D.A., Galloway, S.M., Marnett, L.J., Novak, R.F., Pettit, S.D., Preston, R.J., Skare, J.A., Williams, G.M., *et al.* (2005). Proceedings of a workshop on DNA adducts: biological significance and applications to risk assessment Washington, DC, April 13-14, 2004. *Toxicol Appl Pharmacol* 208, 1-20.

Sanderson, S., Salanti, G., and Higgins, J. (2007). Joint effects of the N-acetyltransferase 1 and 2 (NAT1 and NAT2) genes and smoking on bladder carcinogenesis: a literature-based systematic HuGE review and evidence synthesis. *American journal of epidemiology* 166, 741-751.

Sharma A., Singh K., Almasan A. (2012) Histone H2AX Phosphorylation: A Marker for DNA Damage. In: Bjergbæk L. (eds) DNA Repair Protocols. *Methods in Molecular Biology (Methods and Protocols)*, vol 920. Humana Press, Totowa, NJ. https://doi.org/10.1007/978-1-61779-998-3_40

Singh, R., Sram, R.J., Binkova, B., Kalina, I., Popov, T.A., Georgieva, T., Garte, S., Taioli, E., and Farmer, P.B. (2007). The relationship between biomarkers of oxidative DNA damage, polycyclic aromatic hydrocarbon DNA adducts, antioxidant status and genetic susceptibility following exposure to environmental air pollution in humans. *Mutat Res* 620, 83-92.

Taioli, E., Sram, R.J., Binkova, B., Kalina, I., Popov, T.A., Garte, S., and Farmer, P.B. (2007). Biomarkers of exposure to carcinogenic PAHs and their relationship with environmental factors. *Mutat Res* 620, 16-21.

Totsuka Y, Watanabe M, Lin Y. New horizons of DNA adductome for exploring environmental causes of cancer. *Cancer Sci.* 2021;112(1):7-15. doi:10.1111/cas.14666

US NAS - National Academy of Science (1983). Risk assessment in the Federal Government: Managing the process. National Research Council, Committee on the Institutional Means for Assessment of Risks to Public Health. National Academy Press, Washington, DC.

Vacek, P.M., Albertini, R.J., Sram, R.J., Upton, P., and Swenberg, J.A. (2010). Hemoglobin adducts in 1,3-butadiene exposed Czech workers: female-male comparisons. *Chem Biol Interact* 188, 668-676.

Veglia, F., Matullo, G., and Vineis, P. (2003). Bulky DNA adducts and risk of cancer: a meta-analysis. *Cancer Epidemiol Biomarkers Prev* 12, 157-160.

Verma, M. (2015) The Role of Epigenomics in the Study of Cancer Biomarkers and in the Development of Diagnostic Tools. *Adv Exp Med Biol* 867, 59-80.

Vikstrom, A.C., Warholm, M., Paulsson, B., Axmon, A., Wirfalt, E., and Tornqvist, M. (2012). Hemoglobin adducts as a measure of variations in exposure to acrylamide in food and comparison to questionnaire data. *Food Chem Toxicol*.

Vineis, P., Brennan, P., Canzian, F., Ioannidis, J.P., Matullo, G., Ritchie, M., Stromberg, U., Taioli, E., and Thompson, J. (2008). Expectations and challenges stemming from genome-wide association studies. *Mutagenesis* 23, 439-444.

Vineis, P., and Chadeau-Hyam, M. (2011). Integrating biomarkers into molecular epidemiological studies. *Curr Opin Oncol* 23, 100-105.

Vineis, P., Chadeau-Hyam, M., Gmuender, H., Gulliver, J., Herceg, Z., Kleinjans, J., Kogevinas, M., Kyrtopoulos, S., Nieuwenhuijsen, M., Phillips, D.H., Probst-Hensch, N., Scalbert, A., Vermeulen, R., Wild, C.P. EXPOsOMICS Consortium. (2017) The exposome in practice: Design of the EXPOsOMICS project. *Int J Hyg Environ Health* 220, 142-151.

Vineis P., Matullo G., Berwick M. (2005) Molecular Epidemiology. In: Ahrens W., Pigeot I. (eds) Handbook of Epidemiology. Springer, Berlin, Heidelberg.
https://doi.org/10.1007/978-3-540-26577-1_28

Vineis, P., and Perera, F. (2000). DNA adducts as markers of exposure to carcinogens and risk of cancer. *Int J Cancer* 88, 325-328.

Vineis, P., and Perera, F. (2007). Molecular epidemiology and biomarkers in etiologic cancer research: the new in light of the old. *Cancer Epidemiol Biomarkers Prev* 16, 1954-1965.

Vineis P, Robinson O, Chadeau-Hyam M, Dehghan A, Mudway I, Dagnino S. What is new in the exposome? *Environ Int.* 2020 Oct;143:105887. doi: 10.1016/j.envint.2020.105887. Epub 2020 Jun 30. PMID: 32619912.

Virk-Baker MK, Nagy TR, Barnes S, Groopman J. Dietary acrylamide and human cancer: a systematic review of literature. *Nutr Cancer.* 2014;66(5):774-90. doi: 10.1080/01635581.2014.916323. Epub 2014 May 29. PMID: 24875401; PMCID: PMC4164905.

Vorkamp, K., Castaño, A., Antignac, J.-P., Boada, L.D., Cequier, E., Covaci, A., López, M.E., Haug, L.S., Kasper-Sonnenberg, M., Koch, H.M., Luzardo, O.P., Osīte, A., Rambaud, L., Pinorini, M.-T., Sabbioni, G., Thomsen, C. (2021) Biomarkers, matrices and analytical methods targeting human exposure to chemicals selected for a European human biomonitoring initiative. *Environment International*, 146,

Vrijens, K., Bollati, V., Nawrot, T.S. (2015). MicroRNAs as potential signatures of environmental exposure or effect: a systematic review. *Environ Health Perspect* 123, 399-411.

Vodenkova, S., Polivkova, Z., Musak, L., Smerhovsky, Z., Zoubkova, H., Sytarova, S., Kavcova, E., Halasova, E., Vodickova, L., Jiraskova, K., Svoboda, M., Ambrus, M., Hemminki, K., Vodicka, P. (2015) Structural chromosomal aberrations as potential risk markers in incident cancer patients. *Mutagenesis*, 30, 557-563

Wang, Y., Yang, H., Li, L., Wang, H., Xia, X., and Zhang, C. (2012). Biomarkers of chromosomal damage in peripheral blood lymphocytes induced by polycyclic aromatic hydrocarbons: a meta-analysis. *International archives of occupational and environmental health* 85, 13-25.

Wild, C.P. (2005). Complementing the genome with an "exposome": the outstanding challenge of environmental exposure measurement in molecular epidemiology. *Cancer Epidemiol Biomarkers Prev* 14, 1847-1850.

Wild, C.P. (2012). The exposome: from concept to utility. *Int J Epidemiol.* 41 24-32

Wilde, E.C., Chapman, K.E., Stannard, L.M. et al. A novel, integrated in vitro carcinogenicity test to identify genotoxic and non-genotoxic carcinogens using human lymphoblastoid cells. Arch Toxicol 92, 935–951 (2018).
<https://doi.org/10.1007/s00204-017-2102-y>

Wogan, G.N., Kensler, T.W., and Groopman, J.D. (2011). Present and future directions of translational research on aflatoxin and hepatocellular carcinoma. A review. Food Addit Contam Part A Chem Anal Control Expo Risk Assess, 1-9.

Xu, Y., Cui, B., Ran, R., Liu, Y., Chen, H., Kai, G., Shi, J. (2014) Risk assessment, formation, and mitigation of dietary acrylamide: current status and future prospects. Food Chem Toxicol 69, 1-12.

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